



United States  
Department of  
Agriculture

Forest  
Service

Agriculture  
Handbook 660

87

# Guide to the Olethreutine Moths of Midland North America (Tortricidae)

William E. Miller

DISKENT SERIAL RECORDS

SEP 16 '87

USDA  
NATL. AGRIC. LIBRARY  
GAINESVILLE, FL  
SEP 16 1987

894234

United States  
Department of  
Agriculture

Forest  
Service

Agriculture  
Handbook 660

# **Guide to the Olethreutine Moths of Midland North America (Tortricidae)**

William E. Miller

Formerly, Chief Insect Ecologist  
U.S. Department of Agriculture, Forest Service  
North Central Forest Experiment Station  
St. Paul, MN

Currently, Adjunct Professor  
Department of Entomology  
University of Minnesota — St. Paul

July 1987

To three pioneering tortricidologists:

James Brackenridge Clemens, 1829?–1867  
Thomas de Grey Walsingham, Sixth Earl, 1843–1919  
Carl Heinrich, 1880–1955

## Contents

	<i>Page</i>
Introduction .....	1
Vegetation of the Region .....	3
Subfamily Olethreutinae: Definition and Diagnosis .....	4
Structural Characters .....	5
Both Sexes .....	5
Male .....	6
Female .....	7
Generic Diagnosis .....	8
Keys to Genera .....	9
Males .....	9
Females .....	11
Species Diagnosis .....	13
Systematics .....	14
Tribe Olethreutini .....	14
Genus <i>Episimus</i> .....	14
Genus <i>Bactra</i> .....	14
Genus <i>Endopiza</i> .....	15
Genus <i>Lobesia</i> .....	17
Genus <i>Endothenia</i> .....	17
Genus <i>Aterpia</i> .....	19
Genus <i>Eumarozia</i> .....	19
Genus <i>Zomaria</i> .....	20
Genus <i>Apotomis</i> .....	21
Genus <i>Pseudosciaphila</i> .....	22
Genus <i>Orthotaenia</i> .....	23
Genus <i>Phaecasiophora</i> .....	24
Genus <i>Olethreutes</i> .....	25
Genus <i>Hedya</i> .....	35
Genus <i>Evora</i> .....	37
Tribe Eucosmini .....	37
Genus <i>Rhyacionia</i> .....	37
Genus <i>Retinia</i> .....	39
Genus <i>Barbara</i> .....	40
Genus <i>Spilonota</i> .....	41
Genus <i>Phaneta</i> .....	41
Genus <i>Eucosma</i> .....	48
Genus <i>Pelochrista</i> .....	54
Genus <i>Epiblema</i> .....	55
Genus <i>Notocelia</i> .....	60
Genus <i>Suleima</i> .....	61
Genus <i>Sonia</i> .....	62
Genus <i>Gypsonoma</i> .....	62
Genus <i>Proteoteras</i> .....	64
Genus <i>Zeiraphera</i> .....	65
Genus <i>Pseudexentera</i> .....	66
Genus <i>Gretchena</i> .....	70
Genus <i>Rhopobota</i> .....	72
Genus <i>Epinotia</i> .....	73
Genus <i>Catastega</i> .....	77

Genus <i>Ancylis</i> .....	78
Genus <i>Hystriophora</i> .....	83
Tribe Grapholitini .....	83
Genus <i>Dichrorampha</i> .....	83
Genus <i>Talponia</i> .....	85
Genus <i>Pammene</i> .....	85
Genus <i>Eucosmomorpha</i> .....	86
Genus <i>Larisa</i> .....	87
Genus <i>Sereda</i> .....	87
Genus <i>Grapholita</i> .....	88
Genus <i>Corticivora</i> .....	90
Genus <i>Cydia</i> .....	91
Genus <i>Ecdytolopha</i> .....	95
Genus <i>Pseudogalleria</i> .....	95
References Cited .....	97
Species Index .....	102
Host Index .....	104

## Introduction

Tortricidae comprise the fourth largest family of North American Lepidoptera, and the subfamily Olethreutinae contains three-fourths of the species. Larvae of this large subfamily feed on buds, shoots, stems, leaves, flowers, fruit, seed, and roots of herbs, shrubs, and trees. Some familiar common names of olethreutine species in Midland North America are blackheaded fireworm, boxelder twig borer, cherry fruitworm, codling moth, European pine shoot moth, eyespotted bud moth, hickory shuckworm, oriental fruit moth, pea moth, spruce needle-miner, and strawberry leafroller (Werner 1982). Midland North America refers here to Michigan, Wisconsin, and Minnesota.

The available handbooks on North American Olethreutinae appeared more than a half century ago (Forbes 1923; Heinrich 1923b, 1926). The need for modern coverage of the subfamily has grown as new species, synonyms, and errors have emerged. Some genera have recently been reviewed, such as *Zeiraphera* (Mutuura and Freeman 1966), *Rhyacionia* (Powell and Miller 1978), and *Corticivora* (Brown 1984). But only works of broader scope comparable to the modern handbooks on Eurasian olethreutines (Benander 1950, Bentinck and Diakonoff 1968, Bradley *et al.* 1979, Hannemann 1961, Kuznetsov 1978, and others) adequately serve some purposes. This Guide treats adults, but larval hosts are also given. Works on larval taxonomy of North American as well as Eurasian species are available (MacKay 1959, 1962; Swatschek 1958).

Past family-level works might lead one to conclude erroneously that the olethreutine fauna in Midland North America is scant. Fernekes (1906) listed only 28 species in 9 genera for Milwaukee County, WI. The material on which Heinrich's (1923b, 1926) revisions were based included specimens from Michigan, Wisconsin, and Minnesota for only 30 species in 17 genera. Huang (1962) reviewed the Grapholitini (formerly Laspeyresiini) of Wisconsin, one of three olethreutine tribes, and recorded only 11 species in 5 genera. The present Guide includes 286 species or species complexes in 47 genera, which are 35 percent and 80 percent, respectively, of these olethreutine categories known in North America (Powell 1983).

This Guide began to be assembled two decades ago. Designed as a compact working tool and reference, it emphasizes comprehensive regional coverage, diagnostics of genera and species, accurate species identities, and sources of further systematics as well as biological literature. Potential users are detection and survey entomologists in a region where agriculture and forestry are immensely important; researchers in systematics, biology, and faunistics; some educators; some students; and not least, some avocational entomologists. Much of the specimen base underlying the Guide is due to dedicated recreational collecting and curating.

Species were included only if one or more examined adults originated in Michigan, Wisconsin, or Minnesota. The Guide serves as a record of species occurrence in each State of the region. The search for study material was extensive, and nearly 7,000 pertinent specimens were assembled. Some characteristics of the sample are summarized by State in table 1. Michigan's first rank in both species abundance measures is probably due more to the large proportion of sample specimens originating there than to other factors. More specimens might

**Table 1—Distribution of the *olethreutine* sample by States**

State	Percentage of region in State <sup>1</sup>	Percentage of specimens originating in State <sup>2</sup>	Percentage of regional species found in State <sup>3</sup>	Percentage of regional species found exclusively in State
Michigan	30	65	90	28
Wisconsin	28	20	51	2
Minnesota	42	15	50	6
Total	100	100		

<sup>1</sup>Total of 495,441 km<sup>2</sup> of land area in the three-State region, excluding inland waters.

<sup>2</sup>Total of 6,936 specimens in the Michigan–Wisconsin–Minnesota sample.

<sup>3</sup>Total of 286 species and species complexes found in the three-State region.

add species records appreciably in Wisconsin and Minnesota, but not appreciably in the region as a whole.

The study material is located in source institutions as follows:

	Percentage of sample
Michigan State University Entomology Museum, East Lansing	47
University of Michigan Museum of Zoology, Ann Arbor	15
University of Minnesota Entomology Museum, St. Paul	15
University of Wisconsin Entomology Museum, Madison	13
Essig Museum of Entomology, University of California, Berkeley	6
Others	4
(Field Museum of Natural History, Chicago; Illinois Natural History Survey, Urbana; National Museum of Natural History, Washington, DC; American Museum of Natural History, New York; Los Angeles County Museum of Natural History, Los Angeles; E.G. Voss collection, Ann Arbor, MI; Canadian National Collection of Insects, Ottawa)	
Total	100

Nearly all the study material was captured at lights. The most frequent collectors in Michigan were S. Moore, W.C. Stinson, R.R. Dreisbach, W.W. Newcomb, J.P. Donahue, M.C. Nielsen, J.H. Newman, and R.L. Fischer; in Wisconsin, H.M. Bower, R.D. Shenefelt, W.E. Sieker, E.F. Legner, J.T. Eschle, and E.R. Oatman; and in Minnesota, O. Lugger, W.E. Miller, and D.G. Denning.

Nearly 2,600 genitalia were prepared for identification and illustration, using the method of Clarke (1941). Each preparation bears the name of one of the following preparators: B.J. Thompson, C.D. Waddell, V. Adams, P. Jacus, R.B. Moore, J.A. Blackwell, P. Brigham, K. Lindstrom, K.A. Kohn, C.W. Taylor, L. Holtz, D. Hagler, J.T. Eschle, L.K. Miller, W.E. Miller, M.A. Menke, K. Hosfield, A.C. Collison, J.M. Muggli, S.M. Galeota, H. Scroggins, M.M. Doohar, and L.S. Bauer. Voucher specimens are not otherwise labeled. Genitalia were drawn by U. Daigle. Wings were prepared and drawn by M.G. Pogue. All drawings were made firsthand from specimens.

In addition to the persons already named, I am indebted to R.L. Fischer, T.E. Moore, P.J. Clausen, Steven Krauth, J.A. Powell, the late Henry Dybas, G.L. Godfrey, D.R. Davis, F.H. Rindge, J.P. Donahue, W.S. Craig, J.R. Heitzman, and the late T.N. Freeman for specimen loans; to R.L. Brown, J.A. Powell, and R.W. Hodges for suggesting ways to improve the manuscript; to R.L. Brown for identifying *Epinotia* specimens to reflect his new revision of the genus; to R.L. Kuehn and G.G. Ahlstrand for scanning electron photomicrography; and to Phyllis Moline for painstakingly typing the manuscript.



## Vegetation of the Region

Climate, soils, physiography, and other environmental features of Midland North America are treated in many atlases. Merz (1979) briefly summarizes some background features. Much of the region's environment and history is integrated in its vegetation. Moreover, because olethreutines are seldom polyphagous (Powell 1980), their species diversity in the region can be expected to correlate with the region's vegetative diversity. Larval hosts are reported for 71 percent of the species and species complexes treated in this Guide. However, host profiles for very few species are exhaustive; for most, host information is still fragmentary or lacking.

Küchler's (1964) potential or original vegetation provides a convenient way to characterize the vegetation. Twelve potential vegetation units occupy significant areas of Michigan, Wisconsin, and Minnesota (table 2). Bluestem prairie leads at 15.8 percent of vegetated area, followed by northern hardwoods, pine forest, and others. Exhaustive generic or specific components of units can be found in Küchler (1964). None of the three States has a profile of unit areas similar to that of the three together. The combined profile reflects both forest and plains biomes. With the fewest forest units, and the largest prairie and spruce-fir units, Minnesota is the most plainslike and boreal of the three States. These characteristics, together with large total area, suggest why, despite the smaller proportion of specimens originating in Minnesota, it had more exclusive species occurrences than Wisconsin (table 1).

**Table 2—Potential vegetation of Michigan, Wisconsin, and Minnesota as calculated from areas mapped in Küchler (1964)**

Vegetation unit	Percentage of vegetated area			Total
	Michigan	Wisconsin	Minnesota	
Bluestem prairie	0.1	1.4	14.3	15.8
Northern hardwoods	5.9	9.2	0	15.1
Pine forest	5.7	3.8	5.1	14.6
Oak savanna	0	7.6	4.4	12.0
Maple-basswood forest	0	3.4	6.3	9.7
Spruce-fir forest	0	0.7	6.9	7.6
Northern hardwoods-fir forest	5.6	1.6	0	7.2
Oak-hickory forest	6.6	0	0	6.6
Conifer bog	0.9	1.6	3.7	6.2
Beech-maple forest	1.9	0	0	1.9
Elm-ash forest	1.9	0	0	1.9
Floodplain forest	0	0.1	1.3	1.4
Total				100.0

## Subfamily Olethreutinae: Definition and Diagnosis

Forewing 4 to 20 mm long.

Head rough-scaled above. Antenna less than two-thirds forewing length, scales of flagellar segments originating in a well-defined ring. Labial palpus rough-scaled, third segment usually short and horizontal. Ocelli and chaetosemata present.

Metanotal scales longer and paler than mesonotal scales, restricted to lateral areas. Tibiae with all spurs present. Forewing with 11 or 12 veins:  $R_4$  and  $R_5$  separate, stalked, or united;  $M_2$  and  $M_3$  separate, approximate, or connate;  $Cu_2$  originating before distal three-fourths of discal cell (except *Evora*); 1A usually atrophied, 2A and 3A united for more than half their length distally; remaining veins separate.

Hindwing with 7 or 8 veins; Sc- $R_1$  free;  $R_s$  and  $M_1$  approximate toward base or stalked (except *Dichrorampha*);  $M_3$  and  $Cu_1$  separate, connate, stalked, or united; cubital vein of discal cell with pecten on upper side toward base (except *Sereda* and some *Cydia*).

Valva of male genitalia strongly sclerotized except for a basal opening, usually with differentiated cucullus; gnathos membranous to strongly sclerotized, usually fused with lower surface of the tuba analis; fultura superior absent; processus basalis rudimentary or absent. Uncus present or absent.

Abdominal sternum 7 of female simple to deeply emarginate around ostium bursae, or inflected and overlapping it. Lamella antevaginalis and lamella postvaginalis present or absent. Apophyses anteriores and sterigma unconnected. Ductus bursae variable in length, nonsclerotized, partly sclerotized, or entirely sclerotized; corpus bursae without a signum, with a single signum or two signa.

Olethreutinae appears to be monophyletic. Synapomorphies include restriction of scale origins to one partial or complete ring on antennal flagellar segments, presence of a basal opening in male valval sclerotization, and lack of connection between female apophyses anteriores and sterigma.

Information in this section was drawn chiefly from Heinrich (1923b), Horak (1984), Obratsov (1958), and Razowski (1976). The second- and last-named authors review the history of tortricid classification, and the former evaluates the characters used.

Adult Olethreutinae originating in Midland North America can also be diagnosed with keys to families of Lepidoptera such as that in Borror *et al.* (1981).

## Structural Characters

All characters used in keys, in subfamilial and generic diagnoses, and many characters otherwise mentioned in this Guide are summarized below. An effort was made to select characters whose variability could readily be resolved to binary states, and characters for which a given state applies to the species of a genus without exception. This reduces instances where the user must resolve inexact terms like "sometimes" or "often." When not clearly established by previous authors, such characters were established here by examining the Midland North American representatives.

Characters and states are illustrated where species having them appear in the Systematics section. The characters and page numbers of illustrations are grouped below according to sex and body part. The illustrations represent any degree of manifestation, not necessarily extremes.

Venation can usually be revealed by touching a capillary tube of solvent such as xylol to the wings while light is directed through them; the solvent evaporates quickly and does not harm the specimen. Genitalic characters can sometimes be revealed by brushing away scaling, but more elaborate preparation (Clarke 1941) may be necessary. Nomenclature of wing veins and genitalia follows Borror *et al.* (1981) and Klots (1970), respectively. Venation character states pertain to vein origins unless otherwise specified.

Most structural characters are practically invariant, but venation may be an exception. The keys and diagnoses are based on sample sizes that may not encompass the extensive variation in venation that Tripp (1954) found in one species. To minimize risk of identification errors, the user should consider co-occurring nonvenational character states and use more than one specimen whenever possible.

In many genera, species are distinguishable more often by shape or outline of the male valva and the female ostium bursae and associated structures, than by any other structures. Small differences in orientation of valvae on microslides can lead to perceived differences in valval outline; instances are noted by Diakonoff (1964:67) and Powell and Miller (1978:10). In this Guide, drawings are of valvae mounted as near flat as attainable without distortion. The user should ensure that valvae for comparison are similarly oriented.

### Both Sexes

	<i>Page</i>
<i>Antenna</i>	
Scale origins on flagellar segments restricted to one partial or complete ring . . . . .	67
<i>Thorax</i>	
With posterior tuft . . . . .	23
<i>Forewing</i>	
With raised scale tufts . . . . .	65
Termen concave . . . . .	78
Termen convex . . . . .	38
Apex falcate . . . . .	80
Vein R <sub>1</sub> originating at or near middle of discal cell . . . . .	37
R <sub>2</sub> originating nearer R <sub>3</sub> than R <sub>1</sub> . . . . .	57
Upper internal vein of discal cell originating between R <sub>1</sub> and R <sub>2</sub> . . . . .	57
R <sub>3</sub> and R <sub>4</sub> approximate . . . . .	35

R <sub>4</sub> and R <sub>5</sub> united .....	62
M <sub>2</sub> and M <sub>3</sub> connate .....	39
M <sub>2</sub> , M <sub>3</sub> , and Cu <sub>1</sub> approximate at termen .....	62
Cu <sub>2</sub> originating before distal two-thirds of discal cell .....	67
Cu <sub>2</sub> originating at or beyond distal three-fourths of discal cell .....	37
<i>Hindwing</i>	
Veins Rs and M <sub>1</sub> approximate toward base or connate .....	38
M <sub>2</sub> and M <sub>3</sub> approximate, or M <sub>2</sub> appreciably bent at base .....	18
M <sub>2</sub> and M <sub>3</sub> remote .....	33
M <sub>3</sub> and Cu <sub>1</sub> separate .....	16
M <sub>3</sub> and Cu <sub>1</sub> connate .....	18
Cubital vein with pecten on upper side .....	15
<b>Male</b>	
<i>Antenna</i>	
Notched near base .....	41
<i>Legs</i>	
Hindmost tibia with dilated and tufted scaling .....	24
Hindmost tibia with basal hair pencil .....	31
<i>Forewing</i>	
With costal fold. Grant (1978) examined the anatomy of the costal fold and associated hair pencils .....	58
<i>Hindwing</i>	
Anal margin modified .....	36
Anal margin with lobe .....	31
Upper surface with melanic sex scaling. Brown and P.R. Miller (1983) examined structure and function of these scales .....	64
<i>Abdomen</i>	
Segment 1 with paired ventrolateral papilliform scale pockets .....	17
Segments 6 and 7 with modified dorsal hair tufts beneath scaling .....	86
Segment 8 with paired lateral tufts. Baker and Cardé (1979) demonstrated the courtship function of these tufts .....	89
<i>Genitalia</i>	
Uncus present or developed .....	78
Uncus bifid .....	78
Uncus bifurcate .....	72
Socius or hamus present .....	39
Socius finger- or ribbonlike and not heavily sclerotized .....	39
Anellus loosely surrounding aedeagus .....	78
Valva with basal opening in sclerotization .....	21
Valva divided .....	83
Valval sacculus densely clothed with spinelike setae .....	76
Valva with rudimentary clasper .....	63
Mid-point of valval neck nearer base than apex of valva .....	59
Cross-sectional width of valva at mid-point of neck relative to lineal sacculus width .....	88
Valval Sc <sub>1</sub> at or near base of cucullus .....	23
Valval Sc <sub>1</sub> on neck projection separate from cucullus .....	17
Valval Sc <sub>2</sub> on a projecting digitus .....	20
Valval outer surface with spinelike setae .....	19
Valval cucullus with thick spinelike seta projecting from lower margin .	55
Valval cucullus lacking thick spinelike seta; may have thin spinelike seta projecting from lower margin .....	50
Vesica of aedeagus with cornuti .....	60
Vesica with some cornuti nondeciduous .....	60

## Female

### *Abdomen*

Sternum 7 emarginate posteriorly .....	86
Sternum 7 deeply emarginate .....	68
Sternum 7 inflected and overlapping ostium bursae .....	71
Sternum 7 with median bilobed projection over ostium bursae .....	41

### *Genitalia*

Apophyses anteriores (apo ant) and sterigma unconnected .....	67
Lamella antevaginalis (lam antevag) conical, and lamella postvaginalis (lam postvag) absent .....	78
Lamella antevaginalis wider than lamella postvaginalis in anteroposterior orientation .....	19
Lamella postvaginalis narrow, recessed, and ostium bursae elongate .....	65
Ductus bursae sclerotized with or without minor discontinuities from near genital opening to two-thirds or more its length .....	24
Corpus bursae with sides sclerotized .....	72
Corpus bursae with two signa .....	54
Signa thornlike .....	87
Signa finlike .....	54
Signa scobinate .....	23
Signa bladeliike .....	80
Signa pocketlike .....	18

### *Ovipositor*

Papillae anales with ventral extensions .....	49
---	----

## Generic Diagnosis

The generic diagnoses appearing in the Systematics section utilize only those characters and character states that vary among the genera of a given tribe. For example, presence or absence of vesical cornuti is used to help diagnose genera of Olethreutini because cornuti occur in some genera and not in others; this character is not used for genera of Eucosmini because cornuti occur in all of them. This policy maximizes the usefulness of each character, avoids burdening diagnoses with excess characters, and reduces repetition. In contrast, the keys use all available characters because the keys are not divided into sections by tribe.

The failure of the female key to separate three groups of three to eight genera indicates a need for comparative studies to redefine categories or uncover new diagnostic characters. In these groups, the species illustrations should suffice to identify genera as well as species. One-third of the genera are monotypic or thus far are represented by only one species in Midland North America. Identifying these genera also identifies their species.

Generic diagnoses contain a Comments section with information about the following topics: monotypy; characters that apply less than genus-wide if any; total number of Nearctic species, based mainly on Powell (1983); extra-Nearctic occurrence of species, based on Benander (1950), Bentinck and Diakonoff (1968), Bradley *et al.* (1979), Hannemann (1961), and Kuznetsov (1978); known time of introduction if introduced; literature concerning substantial reviews of Nearctic species, and generic treatments; unconfirmed reports of species in Michigan, Wisconsin, and Minnesota; difficulties that may be encountered in diagnosing species; and species that are considered complexes. For genera with no literature citations, the treatments by Heinrich (1923b, 1926) are still applicable.

## Keys to Genera

Shortest-path keys for use with adults of each sex were generated almost entirely by computer (Pankhurst 1970). They apply chiefly to the Midland North American fauna and may be inadequate for use elsewhere or for use with species yet to be discovered in the region. To locate illustrations of key characters, see the Structural Characters section.

### Males

		Page
1. Antenna notched near base	<i>Spilonota</i>	41
Not so		2
2. Valva of genitalia divided	<i>Hystricophora</i>	83
Not so		3
3. Hindwing cubitus vein without pecten on upper side		4
Not so		5
4. Cross-sectional width of valva at mid-point of neck one-eighth or less lineal sacculus width	<i>Sereda</i>	87
Not so	<i>Cydia</i> (part)	91
5. Socius or hamus of genitalia present		6
Not so		41
6. Tibia of hindmost leg with scaling dilated and tufted	<i>Phaecasiophora</i>	24
Not so		7
7. Hindwing upper surface with melanic sex scaling or costal hair pencil	<i>Proteoteras</i>	64
Not so		8
8. Thorax with posterior tuft		9
Not so		18
9. Forewing upper internal vein of discal cell originating between veins $R_1$ and $R_2$		10
Not so	<i>Zomaria</i>	20
10. Hindwing veins $R_s$ and $M_1$ approximate toward base or connate		11
Not so	<i>Endothenia</i>	17
11. Forewing $Cu_2$ originating at or beyond distal three-quarters of discal cell	<i>Evora</i>	37
Not so		12
12. Tibia of hindmost leg with basal hair pencil		13
Not so		17
13. Forewing $R_3$ and $R_4$ approximate		14
Not so		16
14. Hindwing anal margin with lobe	<i>Olethreutes</i> (part)	25
Not so		15
15. Forewing $Cu_2$ originating before distal two-thirds of discal cell	<i>Hedya</i>	35
Not so	<i>Olethreutes</i> (part)	25
16. Forewing $Cu_2$ originating before distal two-thirds of discal cell	<i>Orthotaenia</i>	23
Not so	<i>Apotomis</i>	21
17. Hindwing anal margin modified	<i>Eumarozia</i>	19
Not so	<i>Pseudosciaphila</i>	22
18. Hindwing anal margin modified	<i>Eucosmomorpha</i>	86
Not so		19

	<i>Page</i>
19. Hindwing Rs and M <sub>1</sub> approximate toward base or connate . . . . .	20
Not so . . . . .	36
20. Forewing upper internal vein of discal cell originating between R <sub>1</sub> and R <sub>2</sub> . . . . .	21
Not so . . . . .	<i>Rhopobota</i> 72
21. Forewing veins R <sub>4</sub> and R <sub>5</sub> united . . . . .	<i>Sonia</i> 62
Not so . . . . .	22
22. Hindwing veins M <sub>3</sub> and Cu <sub>1</sub> connate . . . . .	<i>Episimus</i> 14
Not so . . . . .	23
23. Forewing veins M <sub>2</sub> and M <sub>3</sub> connate . . . . .	24
Not so . . . . .	25
24. Valva of genitalia with rudimentary clasper . . . . .	<i>Retinia</i> 39
Not so . . . . .	<i>Barbara</i> 40
25. Socius of genitalia finger- or ribbonlike and not heavily sclerotized . . . . . . . .	26
Not so . . . . .	31
26. Forewing with costal fold . . . . .	27
Not so . . . . .	<i>Phaneta</i> 41
27. Vesica of aedeagus with some cornuti nondeciduous . . . . .	<i>Notocelia</i> 60
Not so . . . . .	28
28. Valval cucullus of genitalia with thick spinelike seta projecting from lower margin . . . . .	<i>Pelochrista</i> 54
Valval cucullus with or without thin spinelike seta projecting from lower margin . . . . .	29
29. Valva of genitalia with rudimentary clasper . . . . .	<i>Epiblema</i> (part) 55
Not so . . . . .	30
30. Mid-point of valval neck distinctly nearer base than apex of valva . . . . .	<i>Epiblema</i> (part) 55
Not so . . . . .	<i>Eucosma</i> 48
31. Forewing apex falcate . . . . .	<i>Ancylis</i> 78
Not so . . . . .	32
32. Valval sacculus of genitalia densely clothed with spinelike setae . . .	33
Not so . . . . .	<i>Zeiraphera</i> 65
33. Uncus of genitalia developed . . . . .	34
Not so . . . . .	35
34. Anellus loosely surrounding aedeagus . . . . .	<i>Catastega</i> 77
Not so . . . . .	<i>Epinotia</i> 73
35. Forewing with raised scale tufts . . . . .	<i>Gretchena</i> 70
Not so . . . . .	<i>Pseudexentera</i> 66
36. Forewing R <sub>4</sub> and R <sub>5</sub> united . . . . .	<i>Suleima</i> 61
Not so . . . . .	37
37. Forewing upper internal vein of discal cell originating between R <sub>1</sub> and R <sub>2</sub> . . . . .	38
Not so . . . . .	<i>Gypsonoma</i> 62
38. Hindwing M <sub>2</sub> and M <sub>3</sub> approximate, or M <sub>2</sub> appreciably bent at base . . . . .	<i>Bactra</i> 14
Not so . . . . .	39
39. Forewing termen concave . . . . .	<i>Talponia</i> 85
Not so . . . . .	40
40. Abdominal segment 8 with paired lateral tufts . . . . .	<i>Larisa</i> 87
Not so . . . . .	<i>Corticivora</i> 90
41. Forewing M <sub>2</sub> and M <sub>3</sub> connate . . . . .	<i>Rhyacionia</i> 37
Not so . . . . .	42
42. Hindwing Rs and M <sub>1</sub> approximate toward base or connate . . . . .	43
Not so . . . . .	<i>Dichrorampha</i> 83



		Page
43.	Abdominal segment 8 with paired lateral tufts . . . . . <i>Grapholita</i>	88
	Not so . . . . .	44
44.	Valva of genitalia with spinelike setae on outer surface . . . . . <i>Aterpia</i>	19
	Not so . . . . .	45
45.	Vesica of aedeagus with cornuti . . . . .	46
	Not so . . . . .	49
46.	Forewing upper internal vein of discal cell originating between $R_1$ and $R_2$ . . . . .	47
	Not so . . . . .	48
47.	Abdominal segments 6 and 7 with modified dorsal hair tufts beneath scaling . . . . . <i>Pammene</i>	85
	Not so . . . . . <i>Cydia</i> (part)	91
48.	Forewing termen concave . . . . . <i>Pseudogalleria</i>	95
	Not so . . . . . <i>Ecdytolopha</i>	95
49.	Thorax with posterior tuft . . . . .	50
	Not so . . . . . <i>Cydia</i> (part)	91
50.	Valva of genitalia with $Sc_1$ at or near base of cucullus . . . . . <i>Lobesia</i>	17
	Not so . . . . . <i>Endopiza</i>	15

## Females

1.	Hindwing cubital vein with pecten on upper side . . . . .	2	
	Not so . . . . . <i>Sereda</i>	87	
2.	Hindwing veins $Rs$ and $M_1$ approximate toward base or connate . . . . .	3	
	Not so . . . . .	29	
3.	Forewing upper internal vein of discal cell originating between $R_1$ and $R_2$ . . . . .	4	
	Not so . . . . .	26	
4.	Forewing $R_4$ and $R_5$ united . . . . . <i>Sonia</i>	62	
	Not so . . . . .	5	
5.	Hindwing $M_2$ and $M_3$ approximate, or $M_2$ appreciably bent at base . . . . .	6	
	Not so . . . . .	25	
6.	Hindwing $M_3$ and $Cu_1$ connate . . . . .	7	
	Not so . . . . .	16	
7.	Thorax with posterior tuft . . . . .	8	
	Not so . . . . . <i>Episimus</i>	14	
8.	Forewing $Cu_2$ originating at or beyond distal three-quarters of discal cell . . . . .		
	Not so . . . . . <i>Evora</i>	37	
9.	Ductus bursae sclerotized with or without minor discontinuities from near genital opening to two-thirds or more its length . . . . .		
	Not so . . . . . <i>Phaecasiophora</i>	24	
	Not so . . . . .	10	
10.	Corpus bursae with two signa . . . . .	11	
	Not so . . . . .	14	
11.	Forewing $R_3$ and $R_4$ approximate . . . . .	12	
	Not so . . . . . <i>Apotomis</i>	21	
12.	Forewing $Cu_2$ originating before distal two-thirds of discal cell . . . . .	13	
	Not so . . . . . <i>Eumarozia</i>	19	
13.	Signa of corpus bursae bladelike . . . . . <i>Pseudosciaphila</i>	22	
	Not so . . . . . <i>Hedya</i>	35	
14.	Forewing $R_3$ and $R_4$ approximate . . . . .	15	
	Not so . . . . . <i>Orthotaenia</i>	23	
15.	Hindwing $M_2$ and $M_3$ remote . . . . . <i>Aterpia</i>	19	
	Not so . . . . . <i>Olethreutes</i>	25	

	<i>Page</i>
16. Ductus bursae sclerotized with or without minor discontinuities from near genital opening to two-thirds or more its length . . . . .	
. . . . . <i>Hystricophora</i>	83
Not so . . . . .	17
17. Forewing $M_2$ and $M_3$ connate . . . . . <i>Rhyacionia, Retinia, Barbara</i>	37
Not so . . . . .	18
18. Hindwing $M_3$ and $Cu_1$ separate . . . . .	19
Not so . . . . .	20
19. Forewing $R_2$ originating nearer $R_3$ than $R_1$ . . . . . <i>Lobesia</i>	17
Not so . . . . . <i>Endopiza</i>	15
20. Signa of corpus bursae bladelike . . . . . <i>Ancylis</i>	78
Not so . . . . .	21
21. Abdominal sternum 7 with median bilobed projection over ostium bursae . . . . . <i>Spilonota</i>	41
Not so . . . . .	22
22. Lamella antevaginalis conical; lamella postvaginalis absent . . . . .	
. . . . . <i>Catastega</i>	77
Not so . . . . .	23
23. Sternum 7 deeply emarginate; lamella antevaginalis bordered by ridges of sternum . . . . . <i>Proteoteras</i>	64
Not so . . . . .	24
24. Sternum 7 inflected, overlapping ostium bursae; forewing with raised scale tufts . . . . . <i>Gretchena</i>	70
Not so . . . . . <i>Phaneta, Eucosma, Pelochrista, Epiblema, Notocelia, Zeiraphera, Pseudexentera, Epinotia</i>	41
25. Forewing $Cu_2$ originating before distal two-thirds of discal cell . . . . . <i>Pammene, Grapholita, Cydia</i>	85
Not so . . . . . <i>Eucosmomorpha</i>	86
26. Forewing $M_2$ , $M_3$ , and $Cu_1$ approximate at termen . . . . . <i>Rhopobota</i>	72
Not so . . . . .	27
27. Forewing termen concave . . . . . <i>Pseudogalleria</i>	95
Not so . . . . .	28
28. Hindwing $M_2$ and $M_3$ approximate, or $M_2$ appreciably bent at base . . . . . <i>Zomaria</i>	20
Not so . . . . . <i>Ecdytolopha</i>	95
29. Thorax with posterior tuft . . . . . <i>Endothenia</i>	17
Not so . . . . .	30
30. Forewing $R_4$ and $R_5$ united . . . . . <i>Suleima</i>	61
Not so . . . . .	31
31. Forewing upper internal vein of discal cell originating between $R_1$ and $R_2$ . . . . .	32
Not so . . . . . <i>Gypsonoma</i>	62
32. Hindwing $M_2$ and $M_3$ approximate, or $M_2$ appreciably bent at base . . . . . <i>Bactra</i>	14
Not so . . . . .	33
33. Forewing $R_1$ originating at or near middle of discal cell . . . . .	34
Not so . . . . . <i>Dichrorampha</i>	83
34. Forewing termen concave . . . . . <i>Talponia</i>	85
Not so . . . . .	35
35. Forewing $R_3$ and $R_4$ approximate . . . . . <i>Corticivora</i>	90
Not so . . . . . <i>Larisa</i>	87

## Species Diagnosis

Accounts of species in the Systematics section include illustrations of wing pattern, male valva, and female ostium bursae and associated parts. Also included are approved common names if any (Werner 1982); forewing length range in the Michigan-Wisconsin-Minnesota sample to the nearest 0.5 mm as a size index (Miller 1977b), excluding tegula and including fringe; color group (Kelly and Judd 1955) of one conspicuous forewing element, as estimated in fluorescent light at no greater magnification than 2 times; whether the species is polymorphic (Miller 1977a) or excessively variable, and what aspects of the variation are illustrated; supplementary diagnostic information when the illustrations alone may not suffice, or when a character is striking; range in sample capture dates; States in which the regional sample originated (MI for Michigan, WI for Wisconsin, and MN for Minnesota); voltinism if known; up to three larval host genera (Fernald 1970), or species where larval monophagy or stenophagy is known, or host monotypy occurs, together with feeding mode of the larva if known; most recent systematics literature (indicated by Syst); and most recent or comprehensive biological and larval host literature applicable in Midland North America if any (indicated by Biol).

The following partly coded information appears parenthetically for each species:

- Number of specimens in the Michigan-Wisconsin-Minnesota sample (N).
- Number of sample male genitalia preparations examined (Gm).
- Number of sample female genitalia preparations examined (Gf).
- The type was examined (T).
- Origin of specimens whose wing patterns are illustrated that are not from the Michigan-Wisconsin-Minnesota sample.

Types are extant for most of the species in this Guide. As indicated by the frequent appearance of T, nearly all such specimens were examined to confirm species identities. The modern species type concept began to take hold in North American olethreutine systematics in the early 1900's (Miller 1970), but holotypes were not consistently designated until after 1910. Two-thirds of the species treated here were described before 1910. For them, type specimen examination often involved lectotypes or provisional (unpublished) lectotype selections. Much relevant lectotype information can be found in the type studies of Klots (1942) and Miller (1970, 1973a). Some has appeared in scattered works dealing with identity problems, and these works are usually cited as systematics references for those species. More than one-fifth of the type examinations involved provisional lectotypes, particularly for species described by Walker, Zeller, and Walsingham. In such cases, provisional lectotype selections are believed to be sound, the specimens are well marked, and their publication is continuing.

## Systematics

The arrangement of taxa in this Guide follows the current Check List (Powell 1983) as closely as possible. No new taxa or nomenclatural changes are proposed. Synonyms are omitted; for these the user should consult the Check List or the cited systematics literature.

### Tribe Olethreutini

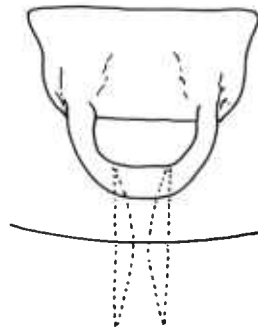
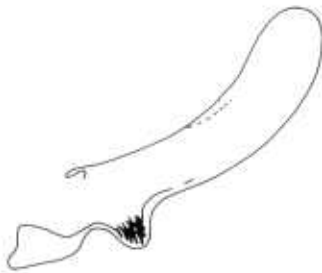
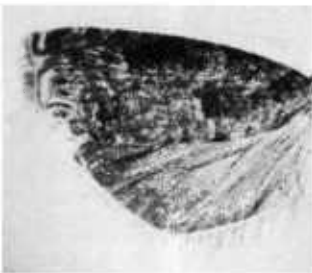
#### Genus *Episimus*

**Both sexes.** Thorax without posterior tuft. Forewing termen concave,  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  approximate at termen,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $Rs$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin unmodified. Hindmost tibia without dilated or tufted scaling, without basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Socius present; valval outer surface lacking spinelike setae,  $Sc_1$  on neck projection separate from cucullus; vesica with cornuti.

**Female.** Sternum 7 not emarginate posteriorly. Lamella antevaginalis developed, not wider than lamella postvaginalis in anteroposterior orientation. Ductus bursae sclerotized only at juncture with corpus bursae; corpus bursae with two thornlike signa.

**Comments.** Three Nearctic species of *Episimus* are known.



*Episimus argutanus* (Clemens)

Forewing 5.0 to 7.0 mm long, dark markings brown. Adults captured May 13–August 24. MI, MN. Bivoltine. Larva feeds in folded or rolled leaves of *Rhus*, *Hamamelis*, *Euphorbia*. Syst: Heinrich (1926). Biol: Jubb (1973). (36 N, 8 Gm, 2 Gf, T)

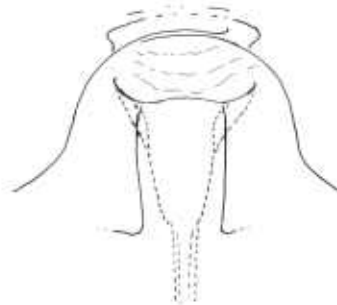
#### Genus *Bactra*

**Both sexes.** Thorax without posterior tuft. Forewing termen straight,  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  not approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $Rs$  and  $M_1$  stalked,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  neither united nor stalked.

**Male.** Hindwing inner margin unmodified. Hindmost tibia without dilated or tufted scaling, without basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Socius present; valval outer surface lacking spinelike setae,  $Sc_2$  not on a projecting digitus.

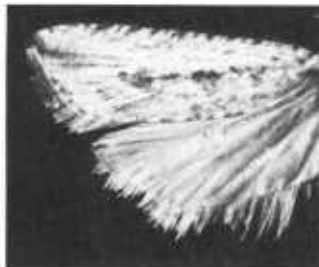
**Female.** Lamella antevaginalis developed, not wider than lamella postvaginalis in anteroposterior orientation. Ductus bursae sclerotized less than one-third its length from genital opening; corpus bursae with single scobinate signum.

**Comments.** Six Nearctic species of *Bactra* are known. Among the species treated here, *Bactra furfurana* also occurs in the Palearctic. Diakonoff (1964) gives a generic treatment.



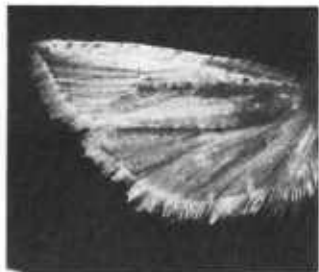
*Bactra furfurana* (Haworth)

Forewing 5.0 to 7.0 mm long, dark markings brown. Adults captured June 13–August 18. MI, WI, MN. Syst: Diakonoff (1964). (12 N, 5 Gm, 5 Gf)



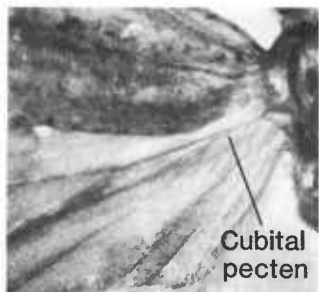
*Bactra verutana* Zeller

Forewing 5.5 to 8.0 mm long, light areas pale orange yellow. Adults captured July 30–October 15. MI, WI, MN. Multivoltine. Larva feeds on leaves, fascicles, and basal bulbs of *Cyperus esculentus*, *Scirpus*, *Juncus*. Syst: Diakonoff (1964). Biol: Frick and Garcia (1975). (17 N, 5 Gm, 5 Gf, T)



*Bactra maiorina* Heinrich

Forewing 8.0 to 9.0 mm long, dark markings brown. Adults captured June 26–July 11. MI, MN. Larva feeds on *Scirpus*. Syst: Diakonoff (1964). Biol: Heinrich (1923a). (2 N, 2 Gm, T, top photo specimen Clay Co., MO)



Cubital pecten

## Genus *Endopiza*

**Both sexes.** Thorax with posterior tuft. Forewing termen convex,  $R_2$  originating equidistant between  $R_1$  and  $R_3$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  separate.

**Male.** Hindwing inner margin unmodified. Hindmost tibia without dilated or tufted scaling, with basal hair pencil. Abdominal segment 1 with paired ven-

trolateral papilliform scale pockets. Socius and hamus absent; valval outer surface lacking spinelike setae, Sc<sub>1</sub> on neck projection separate from cucullus, Sc<sub>2</sub> not on a projecting digitus; vesica without cornuti.

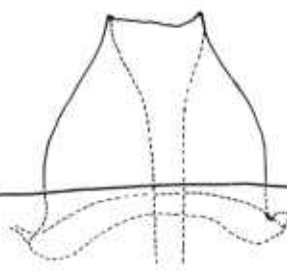
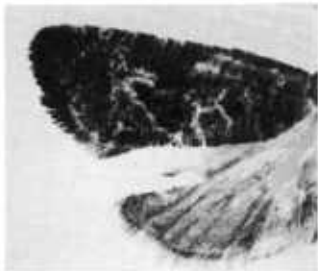
**Female.** Ductus bursae sclerotized less than one-third its length from genital opening; corpus bursae without signa.

**Comments.** Nearly 20 Nearctic species of *Endopiza* are known. *Endopiza* (formerly *Paralobesia*) was reviewed synoptically by Obraztsov (1953).



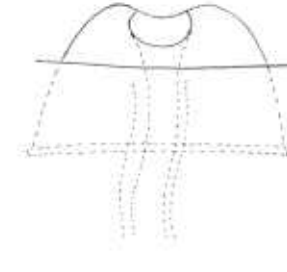
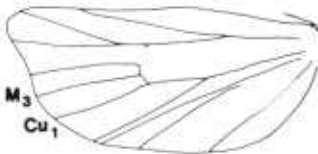
*Endopiza viteana* Clemens.  
Grape berry moth

Forewing 4.5 to 6.0 mm long, basal area purplish blue. Adults captured March 10–August 15. MI. Multivoltine. Larva feeds in developing *Vitis* fruit. Syst: Heinrich (1926). Biol: Pettit (1933). (64 N, 4 Gm, 5 Gf, T)



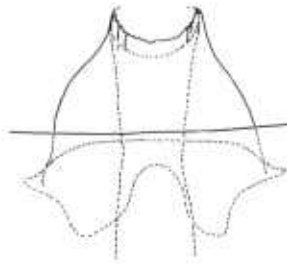
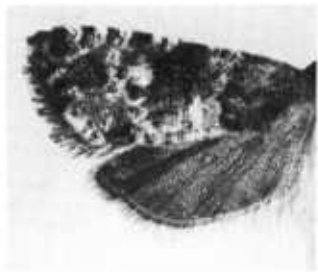
*Endopiza spiraeifolia*  
(Heinrich)

Forewing 4.5 to 5.0 mm long, dark markings brown. Adults captured July 15–August 9. MI, WI. Larva feeds on *Spiraea*. Syst: McDunnough (1938). Biol: Heinrich (1923a). (5 N, 3 Gm, 2 Gf, T)



*Endopiza palliolana*  
(McDunnough)

Forewing 4.5 to 5.0 mm long, dark markings brown. Adults captured or reared May 30–August 8. MI, MN. Bivoltine. Larva feeds within or on *Larix* needles. Syst: McDunnough (1938). Biol: Mosher and Wilson (1974). (15 N, 3 Gm, 4 Gf, T)



*Endopiza aemulana* (Heinrich)

Forewing 5.0 mm long, dark markings brown. Adult captured May 22. MI. Syst: McDunnough (1938). (1 N, 1 Gf, T)

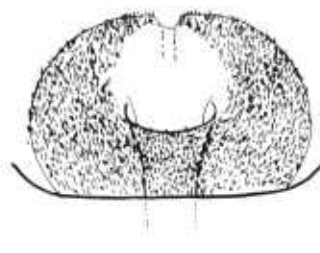
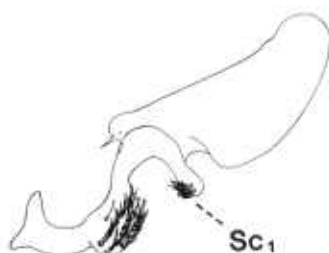
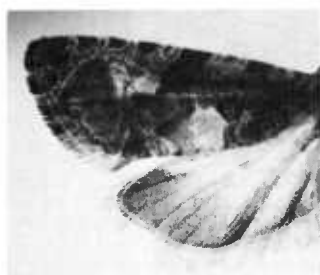
## Genus *Lobesia*

**Both sexes.** Thorax with posterior tuft. Forewing termen convex,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  separate.

**Male.** Hindwing inner margin unmodified. Hindmost tibia without dilated or tufted scaling, with basal hair pencil. Abdominal segment 1 with paired ventrolateral papilliform scale pockets. Socius and hamus absent; valval outer surface lacking spinelike setae,  $Sc_1$  at or near base of cucullus,  $Sc_2$  not on a projecting digitus; vesica without cornuti.

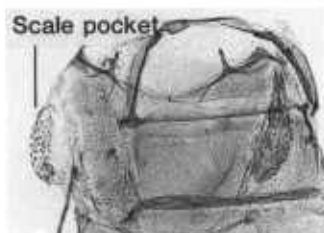
**Female.** Sternum 7 emarginate posteriorly. Ductus bursae sclerotized less than one-third its length from genital opening; corpus bursae without signa.

**Comments.** Two Nearctic species of *Lobesia* are known. Additional species are known from the Palearctic. The genus was reviewed synoptically by Obratsov (1953).



*Lobesia carduana* (Busck)

Forewing 5.0 mm long, dark markings brown or brownish black. Adult captured July 18. MI. Larva feeds in *Cirsium* terminals. Syst: Heinrich (1926). Biol: Marshall and Musgrave (1937). (1N, 1 Gf, T, photo specimen Wayne Co., OH)



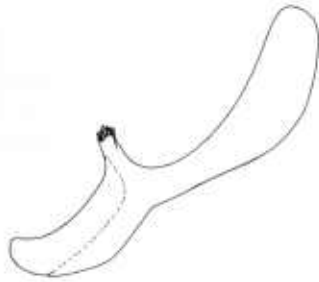
## Genus *Endothenia*

**Both sexes.** Thorax with posterior tuft. Forewing termen straight or convex,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  not approximate,  $M_2$  and  $M_3$  not approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindmost tibia without dilated or tufted scaling. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Uncus developed; socius present; valval outer surface lacking spinelike setae,  $Sc_1$  nearer sacculus than cucullus.

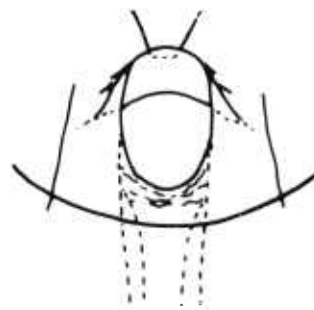
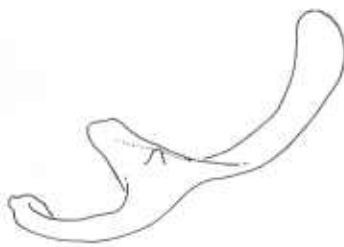
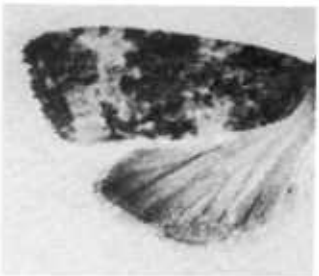
**Female.** Ductus bursae sclerotized less than one-third its length from genital opening; corpus bursae with single pocketlike signum.

**Comments.** Fifteen Nearctic species of *Endothenia* are known. Beebe's (1954) report of the Palearctic *E. gentianaeana* (Hübner) in Michigan is erroneous (Miller 1983b). Among the species treated here, *Endothenia hebesana* also occurs in the Palearctic.



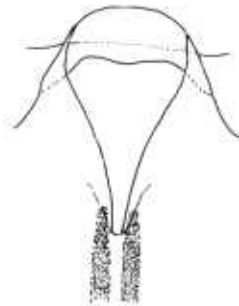
*Endothernia montanana*  
(Kearfott)

Forewing 7.0 to 8.5 mm long, dark areas grayish brown or brownish black. Male hind leg lacks tibial hair pencil. Adults captured May 22–August 7. MI, WI, MN. Larva feeds in *Stachys* stem bases. Syst: McDunnough (1929). Biol: Putman (1942). (7 N, 5 Gm, 2 Gf, T)



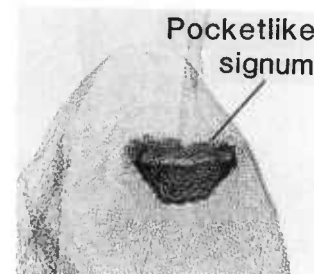
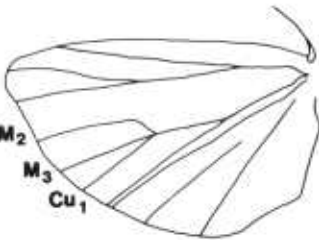
*Endothernia affiliana*  
McDunnough

Forewing 5.5 to 7.0 mm long, dark areas grayish brown or brownish black. Adults captured May 13–July 13. MI. Syst: McDunnough (1942). (5 N, 3 Gf, T)

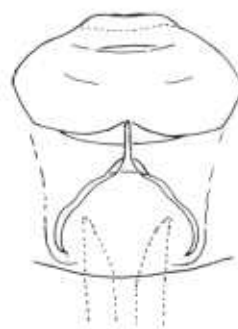
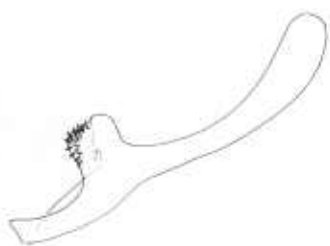


*Endothernia hebesana* (Walker).  
Verbena bud moth

Forewing 5.0 to 8.5 mm long, dark areas and markings brown or brownish black. Adults captured May 7–October 5. MI, WI, MN. Multivoltine. Larva feeds in flowers, stems, and seed capsules of *Verbascum*, *Verbena*, *Sarracenia*, others. Syst and Biol: Miller (1983b). (84 N, 20 Gm, 10 Gf, T)



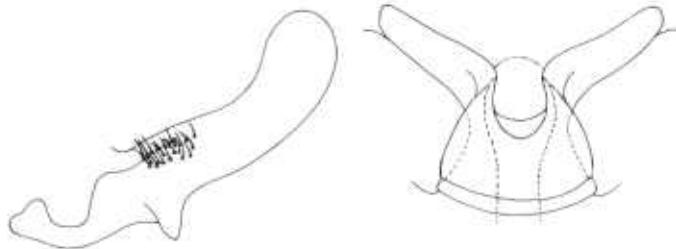
Pocketlike  
signum



*Endothernia nubilana* (Clemens)

Forewing 7.5 to 10.0 mm long, dark markings brown or brownish black. Adults captured June 16–September 3. MI, WI, MN. Larva feeds in *Stachys* stem bases. Syst: Miller (1983b). Biol: Putman (1942). (34 N, 11 Gm, 4 Gf, T)





*Endothernia albolineana* (Kearfott). Spruce needleminer

Forewing 5.0 to 6.0 mm long, dark areas grayish yellowish brown. Adults captured May 7–July 17. MI, WI, MN. Univoltine. Larva feeds within or on *Picea* needles. Syst: Heinrich (1926). Biol: Tashiro (1974). (67 N, 6 Gm, 3 Gf, T)



*Endothernia impudens* (Walsingham)

Forewing 5.5 to 7.0 mm long, dark areas and markings grayish yellowish brown or brownish black. Adults captured July 5–August 9. MI, WI. Syst: Heinrich (1926). (14 N, 5 Gm, 2 Gf, T)

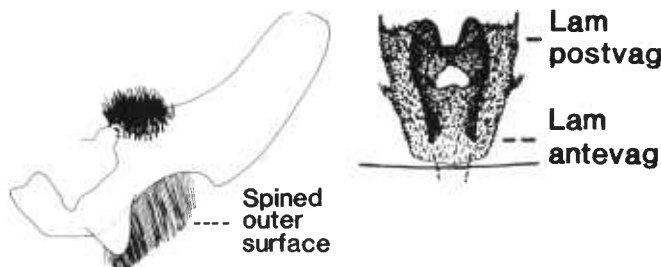
## Genus *Aterpia*

**Both sexes.** Thorax with posterior tuft. Forewing termen convex,  $R_1$  originating well before middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $Rs$  and  $M_1$  approximate,  $M_2$  and  $M_3$  not approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin unmodified. Hindmost tibia without dilated or tufted scaling, without basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Socius and hamus absent; valval outer surface with spinelike setae,  $Sc_1$  at or near base of cucullus; vesica without cornuti.

**Female.** Sternum 7 not emarginate posteriorly. Lamella antevaginalis developed, wider than lamella postvaginalis in anteroposterior orientation. Ductus bursae sclerotized near middle; corpus bursae with single scobinate signum.

**Comments.** Only one Nearctic species of *Aterpia* is known. Additional species are known from the Palearctic.



*Aterpia approximana* (Heinrich)

Forewing 6.0 to 7.5 mm long, dark areas brownish black. Adults captured June 14–September 6. MI, WI. Larva feeds in rolled *Lysimachia* leaves. Syst: Heinrich (1926). Biol: Godfrey *et al.* (1987). (12 N, 7 Gm, 2 Gf, T)

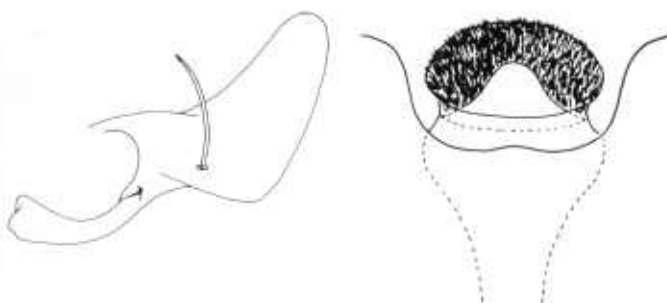
## Genus *Eumarozia*

**Both sexes.** Thorax with posterior tuft. Forewing termen convex, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $Rs$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin modified into a sclerotized ridge. Hindmost tibia without dilated or tufted scaling, without basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Uncus not developed; socius present; valval outer surface lacking spinelike setae,  $Sc_1$  consisting of one spinelike seta at or near base of cucullus,  $Sc_2$  not on a projecting digitus; vesica without cornuti.

**Female.** Sternum 7 emarginate posteriorly. Ductus bursae sclerotized entire length; corpus bursae with two thornlike signa.

**Comments.** Only one Nearctic species of *Eumarozia* is known. Additional species are known from the Neotropics. The genus was reviewed by Clarke (1973).



*Eumarozia malachitana* (Zeller)

Forewing 6.0 to 6.5 mm long, mid-area olive brown. Adults captured June 19–August 10. MI, MN. Larva feeds in rolled leaves of *Ostrya virginiana*, *Diospyros virginiana*. Syst: Clarke (1973). Biol: Marshall and Musgrave (1937). (9 N, 2 Gm, 2 Gf, T)

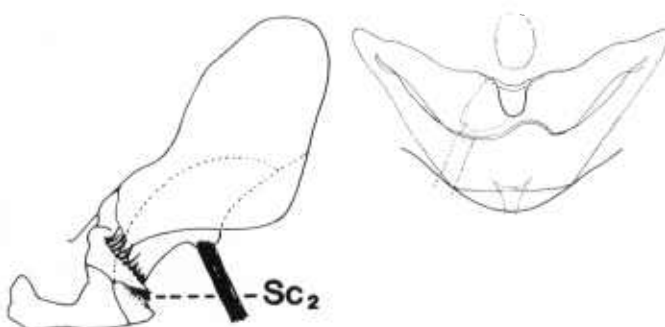
## Genus *Zomaria*

**Both sexes.** Thorax with posterior tuft. Forewing termen convex,  $R_1$  originating well before middle of discal cell,  $R_2$  originating equidistant between  $R_3$  and  $R_1$ , upper internal vein of discal cell originating between  $R_2$  and  $R_3$ ,  $R_3$  and  $R_4$  not approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $Rs$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin unmodified. Hindmost tibia without dilated or tufted scaling, without basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Uncus developed; socius present on underside of uncus; valval outer surface lacking spinelike setae,  $Sc_1$  at or near base of cucullus,  $Sc_2$  on a small projecting digitus; vesica without cornuti.

**Female.** Sternum 7 emarginate posteriorly. Ductus bursae sclerotized only near genital opening; corpus bursae with single scobinate signum that has earlike projections.

**Comments.** Three Nearctic species of *Zomaria* are known.



*Zomaria interruptolineana* (Fernald)

Forewing 5.0 to 7.0 mm long, dark markings brown or brownish black. Adults captured May 22–August 7. MI, WI, MN. Larva feeds in tied leaves of *Gaylussacia*, *Vaccinium*. Syst and Biol: Heinrich (1926). (21 N, 3 Gm, 2 Gf, T)

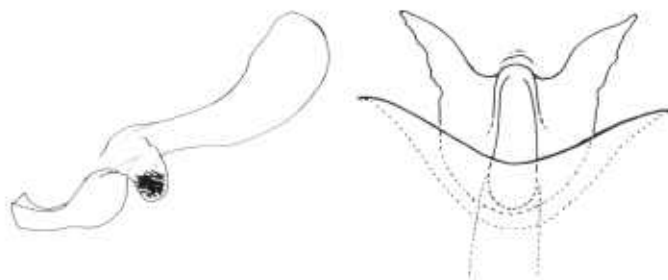
## Genus *Apotomis*

**Both sexes.** Thorax with posterior tuft. Forewing termen straight or slightly convex,  $R_1$  originating well before middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  not approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin modified into a sclerotized ridge. Hindmost tibia without dilated or tufted scaling, with basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Socius present; valval outer surface lacking spinelike setae,  $Sc_1$  on neck projection separate from cucullus; vesica with cornuti.

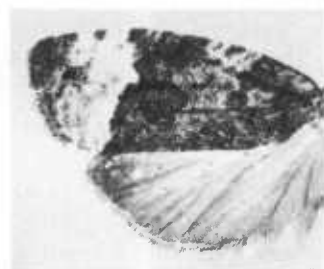
**Female.** Sternum 7 emarginate posteriorly. Ductus bursae sclerotized near middle; corpus bursae with two scobinate signa.

**Comments.** Nearly 20 Nearctic species of *Apotomis* are known. Among the species treated here, *A. capreana* and *A. infida* also occur in the Palearctic. The genus was reviewed by Adamski and Peters (1986).



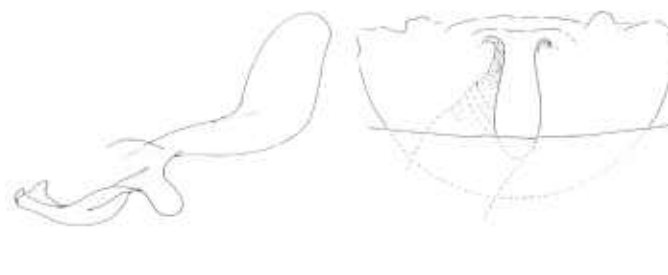
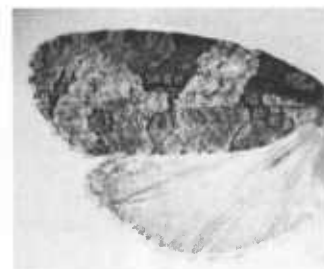
*Apotomis capreana* (Hübner)

Forewing 6.5 to 9.0 mm long, dark areas and markings grayish brown or brownish black. Adults captured June 4–September 7. MI, WI, MN. Larva feeds on *Salix*, *Betula* leaves. Syst: Adamski and Peters (1986). Biol: Prentice (1966). (33 N, 21 Gm, 8 Gf)



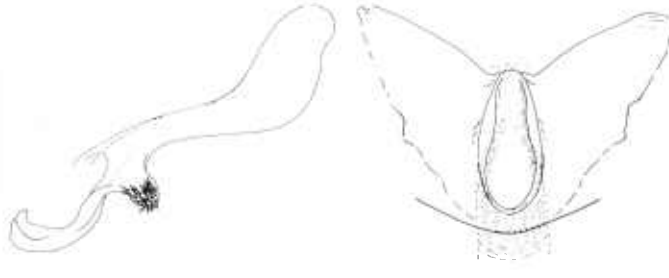
*Apotomis funerea* (Meyrick)

Forewing 8.5 to 10.0 mm long, dark areas grayish brown or brownish black. Adults captured July 4–September 2. MI, WI, MN. Larva feeds in rolled leaves of *Betula*, *Corylus*, others. Syst: Adamski and Peters (1986). Biol: Prentice (1966). (76 N, 13 Gm, 19 Gf, T)



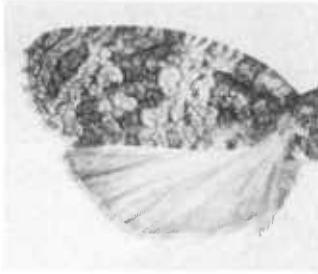
*Apotomis deceptana* (Kearfott)

Forewing 8.5 to 9.0 mm long, dark areas and markings grayish brown. Adults captured July 31–August 25. MI, MN. Larva feeds on *Salix*, *Populus*. Syst: Miller (1979a). Biol: Freeman (1957). (4 N, 3 Gm, 1 Gf, T)



*Apotomis removana* (Kearfott)

Forewing 7.5 to 10.5 mm long, variable pattern; dark areas and markings brownish gray or brownish black. Common variants shown. Adults captured May 29–September 24. MI, WI, MN. Larva feeds in rolled leaves of *Populus*, *Salix*. Syst: Adamski and Peters (1986). Biol: Prentice (1966). (86 N, 30 Gm, 16 Gf, T)



*Apotomis infida* (Heinrich)

Forewing 8.0 to 9.0 mm long, dark areas and markings grayish brown or brownish black. Adults captured June 20–July 14. MI, WI, MN. Larva feeds in rolled leaves of *Salix*, *Populus*. Syst: Adamski and Peters (1986). Biol: Prentice (1966). (7 N, 7 Gm, T)

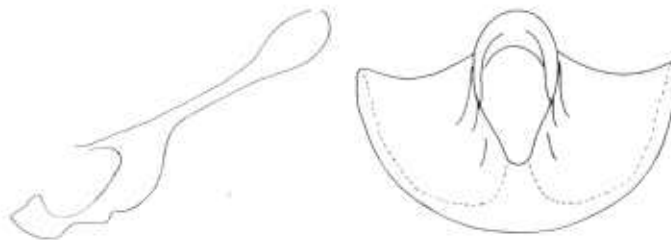
## Genus *Pseudosciaphila*

**Both sexes.** Thorax with posterior tuft. Forewing termen convex,  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin unmodified. Hindmost tibia without dilated or tufted scaling, without basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Uncus developed; valval outer surface with spinelike setae,  $Sc_2$  not on a projecting digitus; vesica without cornuti.

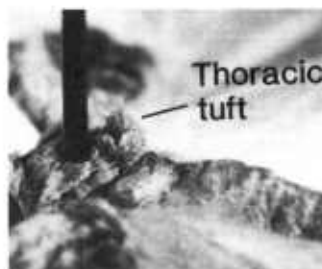
**Female.** Sternum 7 emarginate posteriorly. Lamella antevaginalis developed, wider than lamella postvaginalis in anteroposterior orientation. Ductus bursae sclerotized less than one-third its length from genital opening; corpus bursae with two bladellike signa.

**Comments.** Only one Nearctic species of *Pseudosciaphila* is known. Additional species are known from the Palearctic.



*Pseudosciaphila duplex*  
(Walsingham)

Forewing 9.5 to 11.5 mm long, variable pattern; dark markings or areas yellowish brown, grayish yellowish brown, or brownish black. Nonmelanic and melanic variants shown. Adults captured May 21–July 24. MI, WI, MN. Univoltine. Larva feeds in rolled leaves of *Populus*, *Betula*, *Salix*. Syst: Heinrich (1926). Biol: McGregor (1967). (122 N, 9 Gm, 5 Gf, T)



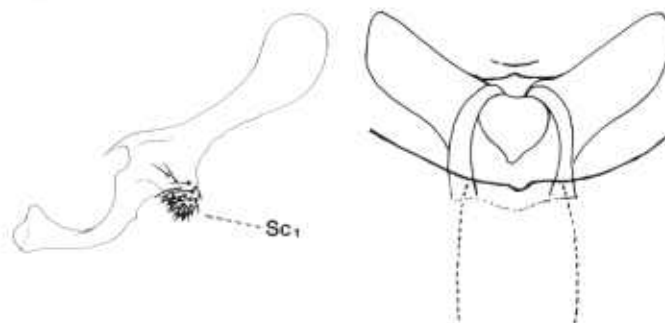
## Genus *Orthotaenia*

**Both sexes.** Thorax with posterior tuft. Forewing termen straight or convex,  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  not approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin modified into a sclerotized thickening. Hindmost tibia without dilated or tufted scaling, with basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Socius present; valval outer surface lacking spinelike setae,  $Sc_1$  on neck projection separate from cucullus; vesica with cornuti.

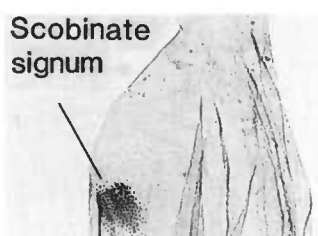
**Female.** Sternum 7 emarginate posteriorly. Ductus bursae sclerotized less than one-half its length from genital opening; corpus bursae with single scobinate signum.

**Comments.** *Orthotaenia* appears to be monotypic; *O. undulana* also occurs in the Palearctic.



*Orthotaenia undulana* (Denis and Schifferrmüller)

Forewing 5.5 to 9.0 mm long, dark areas yellowish brown. Adults captured May 29–August 6. MI, WI, MN. Larva feeds in rolled leaves or terminals of *Populus*, *Salix*, *Betula*, others. Syst: Heinrich (1926). Biol: Prentice (1966). (115 N, 23 Gm, 12 Gf)



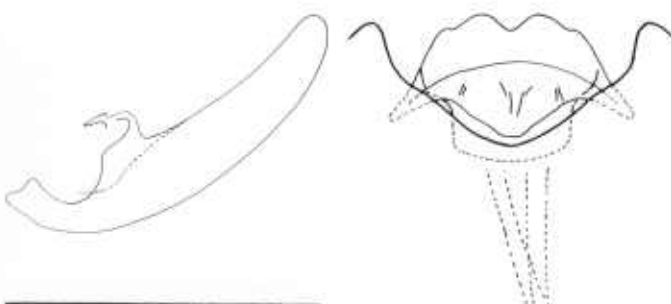
Genus *Phaecasiophora*

**Both sexes.** Thorax with posterior tuft. Forewing termen convex,  $R_1$  originating well before middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  not approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin modified into a sclerotized ridge. Hindmost tibia with dilated and tufted scaling, with basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Uncus not developed; socius present; valval outer surface lacking spinelike setae, vesica with cornuti.

**Female.** Lamella antevaginalis undeveloped. Ductus bursae sclerotized two-thirds its length from genital opening; corpus bursae without signa.

**Comments.** Three Nearctic species of *Phaecasiophora* are known.

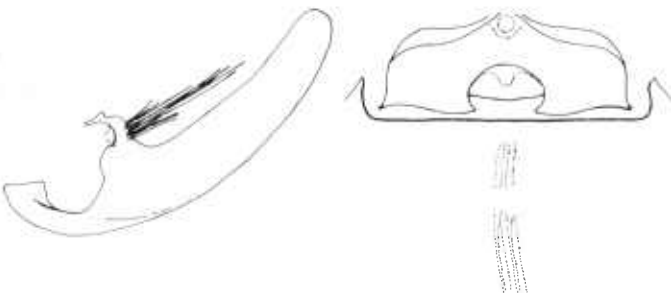
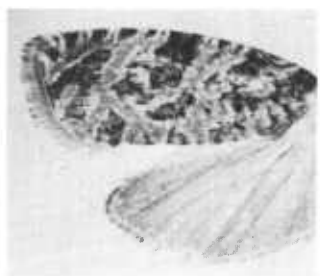


*Phaecasiophora confixana*  
(Walker)

Forewing 8.0 to 9.5 mm long, variable pattern; dark markings or areas yellowish brown or brownish black. Nonmelanic and melanic variants shown. Adults captured May 13–July 13. MI. Syst: Heinrich (1926). (6 N, 2 Gm, 3 Gf, T)

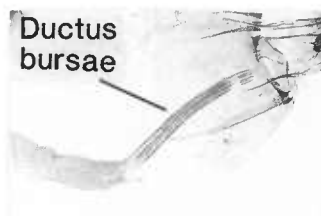


Dilated tibial  
scaling



*Phaecasiophora niveiguttana*  
(Grote)

Forewing 6.5 to 9.0 mm long, dark markings yellowish brown. Adults captured May 14–August 15. MI, WI, MN. Larva feeds in rolled leaves of *Sassafras albidum*. Syst: Heinrich (1926). Biol: MacKay (1959). (34 N, 3 Gm, 3 Gf, T)



Ductus  
bursae

## Genus *Olethreutes*

**Both sexes.** Thorax with posterior tuft. Forewing termen convex or straight, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating before distal three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindmost tibia without dilated or tufted scaling, with basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Uncus developed; socius present.

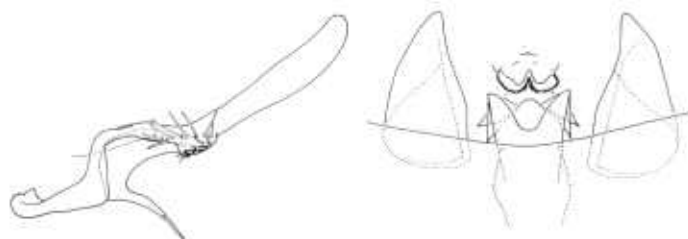
**Female.** Ductus bursae sclerotized only near genital opening; corpus bursae with or without single scobinate or thornlike signum.

**Comments.** Males of the former *Exartema* species are recognizable by the characteristic extended lobe on the hindwing anal margin. Such species constitute more than half the genus. Diakonoff (1973) cursorily treated them in an appendix to his treatment of South Asiatic *Olethreutes*.

More than 75 Nearctic species of *Olethreutes* are known. Among the species treated here, *O. cespitana* and *O. metallicana* occur in the Palearctic.

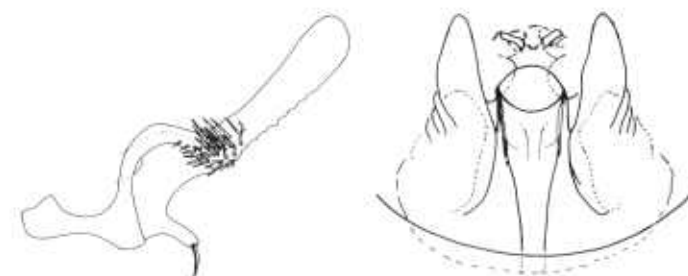
I was unable to confirm Fernekes' (1906) report of *O. exoletus* (Zeller) in the region.

In some species groups, intraspecific variability is high and species limits are ill defined at present. One example is the *O. permundana* group (*clavana*, *melanomesa*, *nigrana*, *permundana*, *tiliana*, *viburnana*). Some specimens of such groups are not always easily identified to species.



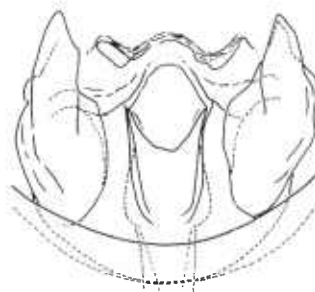
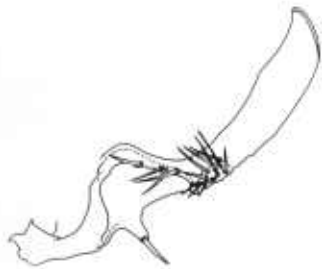
*Olethreutes nitidana* (Clemens)

Forewing 8.0 mm long, pale areas orange yellow. Adult captured July 18. WI. Syst: Heinrich (1926). (1 N, 1 Gm, T, photo specimen lectotype)



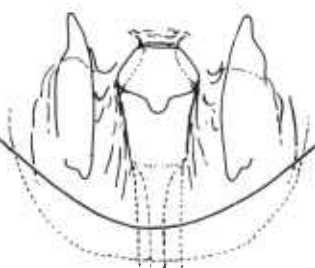
*Olethreutes furfurana* (McDunnough)

Forewing 6.0 to 7.5 mm long, dark areas grayish yellowish brown or brownish black. Adults captured May 30–September 2. MI, WI, MN. Larva feeds on *Rubus*. Syst and Biol: Miller (1979b). (12 N, 8 Gm, 1 Gf, T)



*Olethreutes comandrana*  
(Clarke)

Forewing 7.0 mm long, pale areas of apical third orange. Adult captured July 1. MI. Larva feeds on *Comandra*. Syst and Biol: Clarke (1953). (1 N, 1 Gf, T, photo specimen Putnam Co., IL)



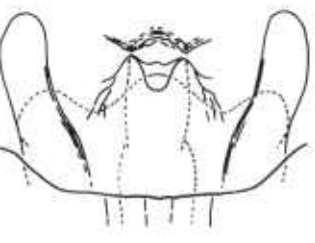
*Olethreutes olivaceana* (Fernald)

Forewing 5.0 to 7.0 mm long, dark areas yellowish brown or olive brown. Adults captured June 11–August 24. MI. Larva feeds on *Fragaria*. Syst: Heinrich (1926). Biol: Vincent *et al.* (1985). (7 N, 6 Gm, 1 Gf, T)



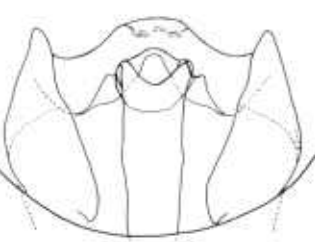
*Olethreutes electrofuscus*  
(Heinrich)

Forewing 6.5 to 8.0 mm long, dark areas yellowish brown. Adults captured July 12–August 2. MI. Larva feeds on *Myrica*, *Comptonia*. Syst: Heinrich (1926). Biol: Ferguson (1975). (7 N, 4 Gm, 1 Gf, T)



*Olethreutes footiana* (Fernald)

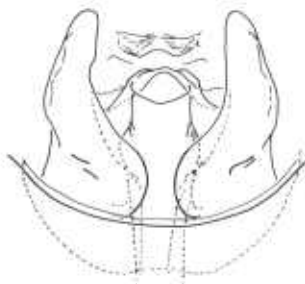
Forewing 8.5 mm long, dark areas yellowish brown or brownish black. Adults captured August 8–9. MI. Larva feeds on *Hamamelis*, *Quercus*. Syst: Heinrich (1926). Biol: Putman (1935). (2 N, 2 Gf, T, photo specimen Essex Co., NJ)



*Olethreutes atrodentana*  
(Fernald)

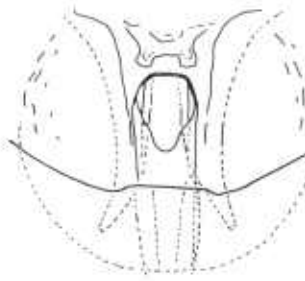
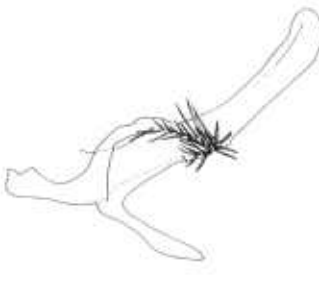
Forewing 7.0 to 9.5 mm long, dark areas yellowish brown or brownish black. Adults captured July 4–August 15. MI, WI, MN. Larva feeds in rolled *Quercus* leaves. Syst: Heinrich (1926). Biol: Prentice (1966). (23 N, 6 Gm, 4 Gf, T)





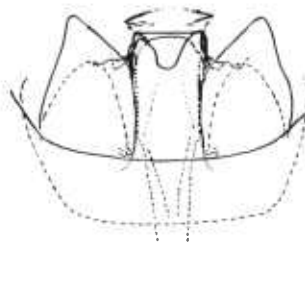
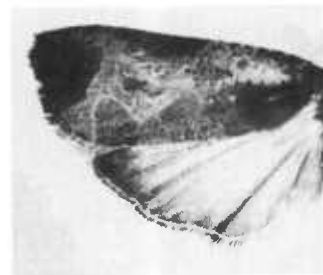
*Olethreutes punctana*  
(Walsingham)

Forewing 7.5 to 9.0 mm long, dark markings brownish black. Adults captured June 18–August 1. MI, WI. Larva feeds in tied *Cornus* leaves. Syst: McDunnough (1935). Biol: McDunnough (1933). (25 N, 6 Gm, 2 Gf, T)



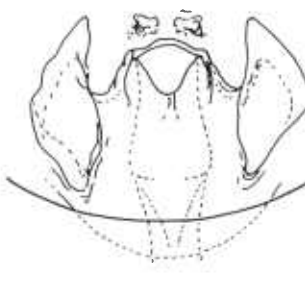
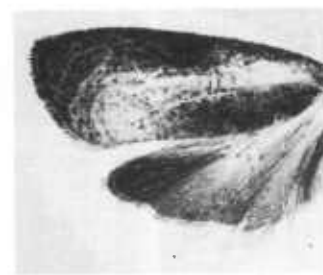
*Olethreutes connectus*  
(McDunnough)

Forewing 7.0 to 9.0 mm long, dark markings brownish black. Adults captured June 13–August 30. MI. Larva feeds in folded *Cornus* leaves. Syst: McDunnough (1935). Biol: McDunnough (1933). (26 N, 7 Gm, 4 Gf, T)



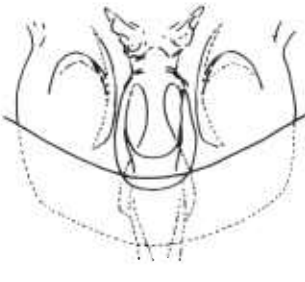
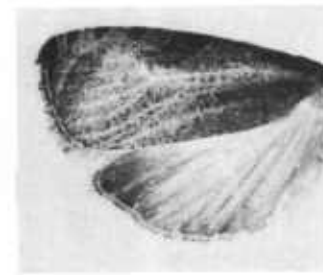
*Olethreutes inornatana*  
(Clemens)

Forewing 8.5 to 10.5 mm long, dark areas brownish black. Adults captured June 26–August 23. MI, WI, MN. Larva feeds on *Prunus*, *Cornus*, others. Syst: Heinrich (1926). Biol: Putman (1935). (38 N, 6 Gm, 2 Gf, T)



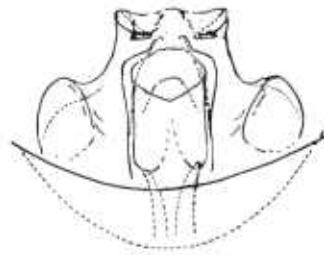
*Olethreutes mysteriana* Miller

Forewing 7.5 to 8.5 mm long, dark areas brownish black. Adults captured July 7–30. MI, WI. Larva feeds on tied *Ulmus*, *Celtis* leaves. Syst and Biol: Miller (1979b). (6 N, 6 Gm, T, photo specimen holotype)



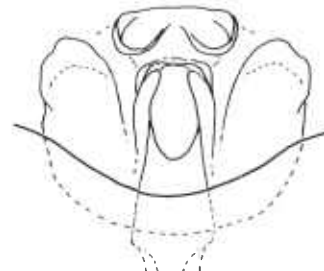
*Olethreutes quadrifidus* (Zeller)

Forewing 8.0 to 9.0 mm long, dark areas grayish yellowish brown or brownish black. Adults captured June 7–July 21. MI, WI. Larva feeds in rolled *Prunus*, *Cornus* leaves. Syst: Heinrich (1926). Biol: Prentice (1966). (18 N, 6 Gm, 3 Gf, T)



*Olethreutes tiliana* (Heinrich)

Forewing 8.0 to 10.0 mm long, pale areas grayish brown. Adults captured July 9–August 12. MI. Larva feeds in tied *Tilia* leaves. Syst and Biol: McDunnough (1942). (13 N, 8 Gm, 1 Gf, T)



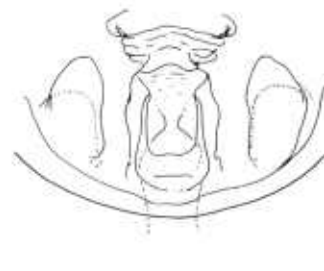
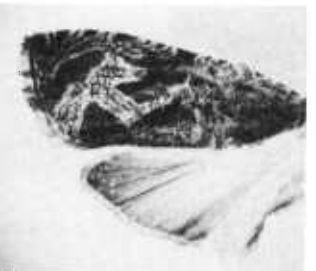
*Olethreutes clavana* (Walker)

Forewing 7.0 to 10.0 mm long, variable pattern; dark markings brown or yellowish brown. Common variants shown. Adults captured July 1–29. MI, WI. Larva feeds in tied *Corylus* leaves. Syst and Biol: Miller (1979b). (23 N, 12 Gm, 5 Gf, T, bottom photo specimen Papineau Co., PQ)



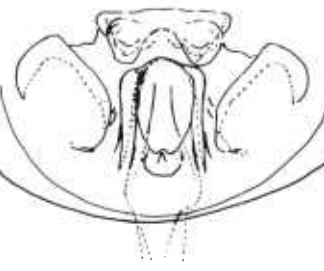
*Olethreutes nigrana* (Heinrich)

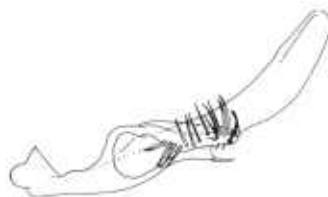
Forewing 9.5 to 10.0 mm long, variable pattern; dark markings brown or yellowish brown. Common variants shown. Adults captured July 15–August 4. MI, WI. Larva feeds on *Acer*, *Carya*. Syst and Biol: Miller (1979b). (4 N, 4 Gm, T, bottom photo specimen Putnam Co., IL)



*Olethreutes viburnana* (McDunnough)

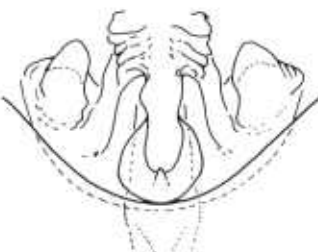
Forewing 8.0 to 9.5 mm long, dark markings grayish brown or brownish black. Adults captured July 10–19. MI, WI. Larva feeds on *Viburnum*. Syst: McDunnough (1935). Biol: Putman (1935). (3 N, 1 Gm, 2 Gf, T)





*Olethreutes merrickana*  
(Kearfott)

Forewing 8.0 to 9.5 mm long, dark markings brown. Adults captured June 26–August 10. MI, WI. Larva feeds on *Ostrya virginiana*, *Carya*. Syst: McDunnough (1944). Biol: Prentice (1966). (14 N, 8 Gm, 2 Gf, T)



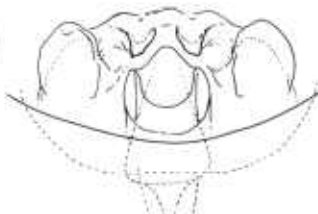
*Olethreutes hamameliana*  
(McDunnough)

Forewing 8.0 to 9.0 mm long, pale areas orange yellow or yellowish brown. Adults captured June 6–July 15. MI, WI. Larva feeds on *Hamamelis*. Syst: McDunnough (1944). Biol: Ferguson (1975). (22 N, 4 Gm, 5 Gf, T)



*Olethreutes corylana* (Fernald)

Forewing 7.0 to 8.5 mm long, dark markings yellowish brown. Adults captured June 9–August 19. MI, WI. Larva feeds on *Corylus*. Syst: Heinrich (1926). Biol: Prentice (1966). (10 N, 6 Gm, 2 Gf, T)



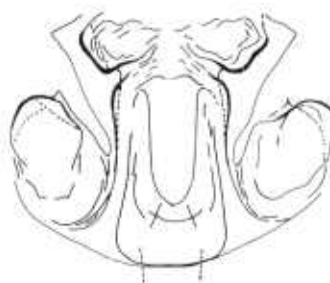
*Olethreutes ochrosuffusana*  
(Heinrich)

Forewing 10.5 mm long, pale areas yellowish brown. Adults captured July 9–15. MI. Larva feeds in folded *Aesculus* leaves. Syst and Biol: Braun (1951). (2 N, 1 Gm, 1 Gf, T, photo specimen Boone Co., MO)



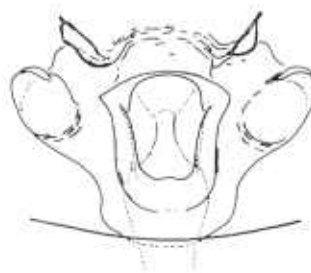
*Olethreutes fagigemmeana*  
(Chambers)

Forewing 7.5 mm long, dark areas brown. Adult captured August 5. MI. Larva feeds on *Fagus grandifolia*, *Ostrya virginiana*. Syst and Biol: Heinrich (1926). (1 N, 1 Gf, T, photo specimen Hamilton Co., OH)



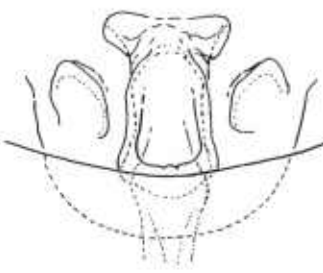
*Olethreutes sericorana*  
(Walsingham)

Forewing 8.5 mm long, pale areas orange yellow. Adult captured July 19. WI, MN. Larva feeds on *Myrica*. Syst: Heinrich (1926). Biol: Schaffner (1959). (2 N, 1 Gm, T)



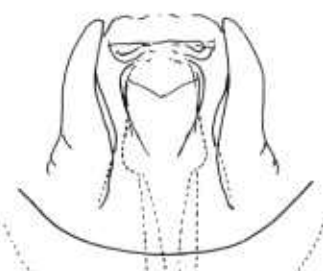
*Olethreutes melanomesa*  
(Heinrich)

Forewing 7.5 to 8.5 mm long, dark areas brown. Adults captured July 7–August 8. MI. Larva feeds in *Kalmia* terminals. Syst: Heinrich (1926). Biol: McDunnough (1954). (11 N, 5 Gm, T)



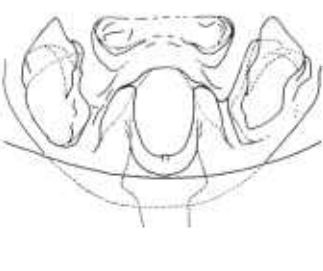
*Olethreutes valdana*  
(McDunnough)

Forewing 6.5 to 8.5 mm long, dark areas yellowish brown. Adults captured June 7–August 17. MI, WI. Larva feeds in tied terminal leaves of *Spiraea*, *Myrica*. Syst and Biol: McDunnough (1956). (31 N, 7 Gm, 11 Gf, T)



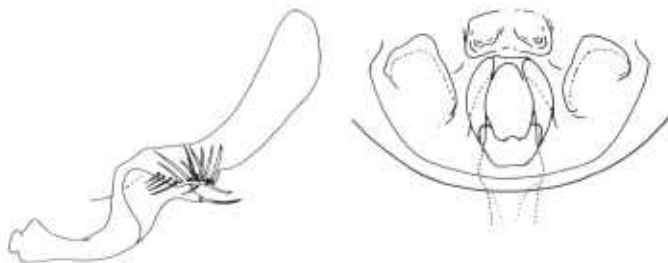
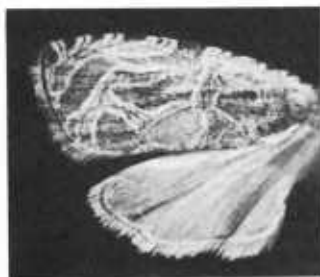
*Olethreutes versicolorana*  
(Clemens)

Forewing 7.0 to 8.5 mm long, dark areas yellowish brown. Adults captured July 1–August 3. MI, WI. Larva feeds on *Cornus*. Syst: Heinrich (1926). Biol: MacKay (1959). (25 N, 10 Gm, 4 Gf, T)



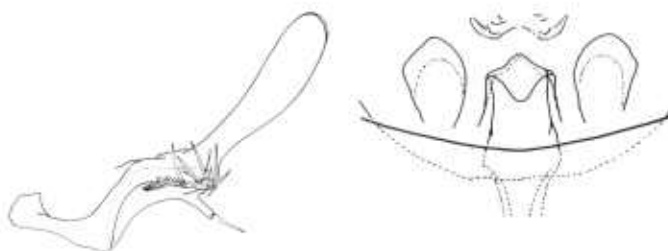
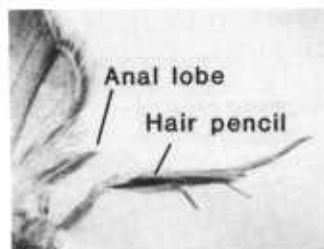
*Olethreutes galevora*  
(McDunnough)

Forewing 7.0 to 8.5 mm long, dark areas brown or brownish black. Adults captured July 6–August 4. MI, WI. Larva feeds in tied terminal leaves of *Myrica*. Syst and Biol: McDunnough (1956). (10 N, 7 Gm, 3 Gf, T)



*Olethreutes permundana*  
(Clemens). Raspberry leafroller

Forewing 6.5 to 9.5 mm long, dark areas yellowish brown. Adults captured June 25–August 15. MI, WI. Larva feeds in leaf and flower buds, rolled leaves of *Rubus*, *Physocarpus opulifolius*. Syst: McDunnough (1956). Biol: Wheeler and Hoebeke (1985). (66 N, 8 Gm, 2 Gf, T)



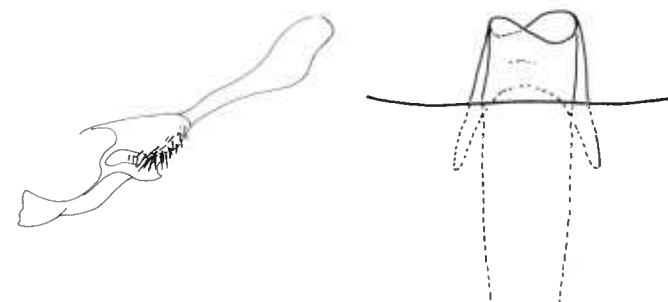
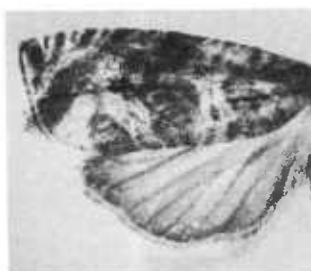
*Olethreutes submissana*  
(McDunnough)

Forewing 7.0 to 8.0 mm long, dark markings brown. Adults captured July 17–August 16. MI, WI, MN. Larva feeds on *Alnus*. Syst: Heinrich (1926). Biol: Ferguson (1975). (19 N, 2 Gm, 7 Gf, T)



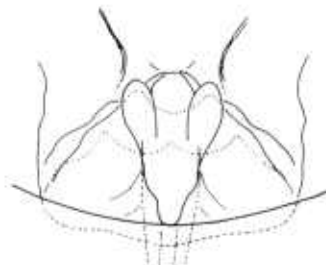
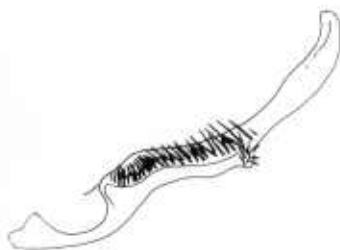
*Olethreutes malana* (Fernald)

Forewing 7.0 mm long, dark markings grayish brown or brownish black. Adult captured August 11. MI, WI. Univoltine. Larva feeds in buds and rolled leaves of *Pyrus*. Syst: Heinrich (1926). Biol: Chapman and Lienk (1971). (2 N, 1 Gf, T, photo specimen McHenry Co., IL)



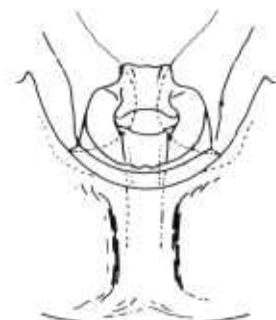
*Olethreutes appendicea* (Zeller)

Forewing 6.0 to 8.0 mm long, dark areas grayish yellowish brown. Adults captured June 11–August 31. MI, WI, MN. Larva feeds in rolled leaves of *Populus*, *Prunus*, *Amelanchier*, others. Syst: Heinrich (1926). Biol: Prentice (1966). (48 N, 21 Gm, 6 Gf, T)



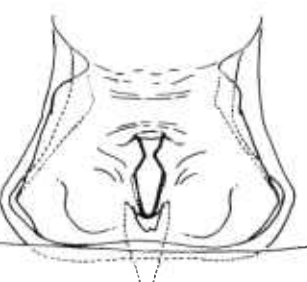
*Olethreutes concinnana*  
(Clemens)

Forewing 6.0 to 7.0 mm long, variable pattern; dark markings and areas grayish brown or brownish black. Melanic and nonmelanic variants shown. Adults captured July 4–13. MI. Syst: Miller (1979b). (4 N, 1 Gm, 2 Gf, T, top photo specimen Jefferson Co., KY)



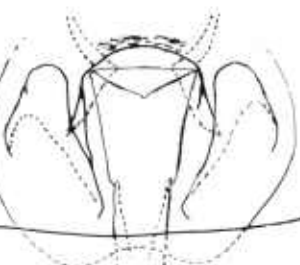
*Olethreutes fasciata*  
(Clemens)

Forewing 5.0 to 7.5 mm long, dark areas brownish black. Adults captured May 26–August 9. MI, WI, MN. Larva feeds in rolled leaves of *Populus*, *Salix*. Syst: Heinrich (1926). Biol: Prentice (1966). (67 N, 12 Gm, 8 Gf, T)



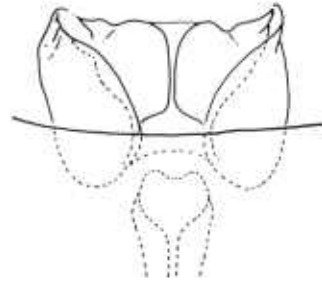
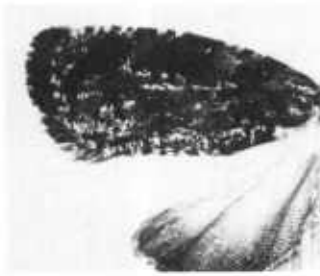
*Olethreutes troglodana*  
(McDunnough)

Forewing 6.5 to 7.0 mm long, dark markings yellowish brown. Adults captured June 21–23. WI. Syst: Miller (1985d). (2 N, 2 Gm, T)



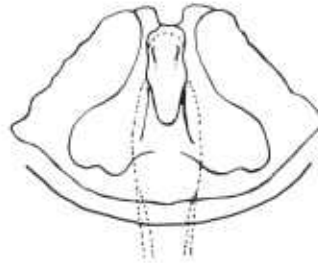
*Olethreutes exaeresima*  
(Heinrich)

Forewing 9.0 mm long, dark markings brown. Adult captured July 23. MI, WI. Larva feeds on *Cornus*. Syst and Biol: Heinrich (1926). (2 N, 2 Gm, T)



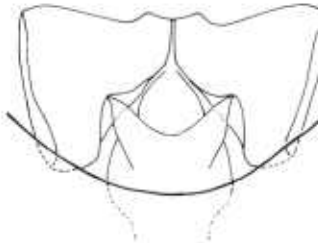
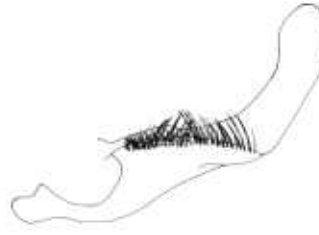
*Olethreutes auricapitana*  
(Walsingham)

Forewing 5.5 mm long, dark areas brownish black. Adult captured between July 17–27. MI. Larva feeds on *Betula*. Syst and Biol: Heinrich (1926). (1 N, 1 Gm, T, photo specimen Boone Co., MO)



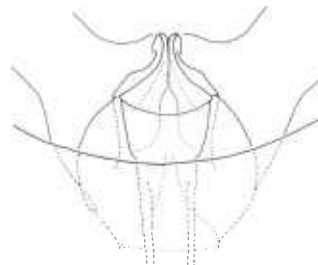
*Olethreutes agilana* (Clemens)

Forewing 4.5 to 7.0 mm long, dark areas brownish black. Adults captured May 23–July 9. MI, MN. Larva feeds in *Impatiens* stems. Syst: Miller (1985d). Biol: MacKay (1959). (9 N, 5 Gm, 4 Gf, T, photo specimen Boone Co., MO)



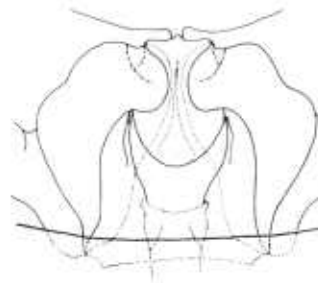
*Olethreutes albiciliana* (Fernald)

Forewing 5.0 to 6.5 mm long, dark markings brownish black. Adults captured June 12–July 9. MI, WI. Larva feeds in rolled leaves of *Spiraea*, *Alnus*. Syst: Heinrich (1926). Biol: Prentice (1966). (9 N, 7 Gm, 2 Gf, T)



*Olethreutes galaxana* Kearfott

Forewing 7.5 mm long, pale areas orange yellow. Adult captured June 25. MN. Syst: Heinrich (1926). (1 N, 1 Gf, T, photo specimen Slope Co., ND)

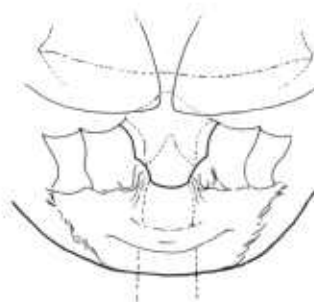
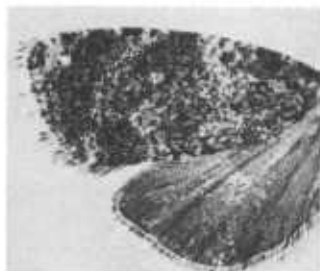


*Olethreutes coruscana*  
(Clemens)

Forewing 7.0 to 10.5 mm long, pale areas orange. Adults captured May 30–August 22. MI, WI, MN. Syst: Miller (1985d). (45 N, 20 Gm, 9 Gf, T)

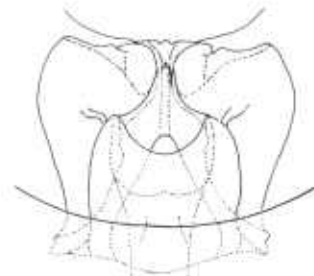
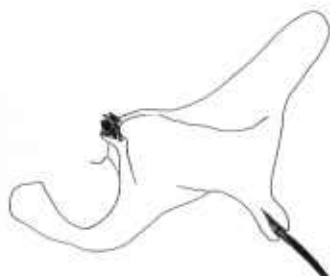
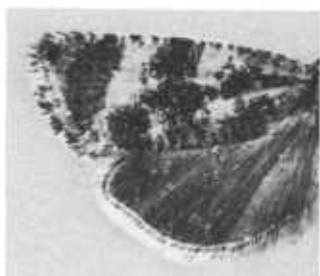






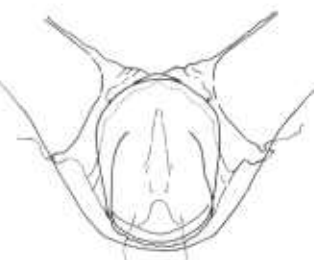
*Olethreutes astrologana* (Zeller)

Forewing 6.5 to 8.5 mm long, pale areas orange yellow. Adults captured May 30–August 15. MI, WI, MN. Syst: Heinrich (1926). (56 N, 28 Gm, 9 Gf, T)



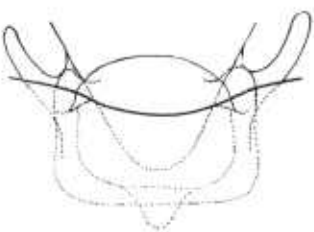
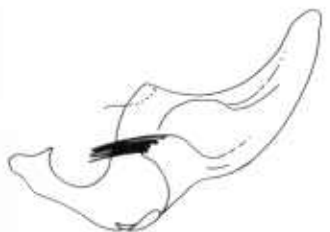
*Olethreutes ferrolinaea* (Walker)

Forewing 7.0 to 9.5 mm long, pale areas orange. Adults captured May 24–July 18. MI, MN. Syst: Miller (1985d). (15 N, 7 Gm, 5 Gf, T)



*Olethreutes metallica* (Hübner)

Forewing 8.0 to 8.5 mm long, variable pattern; pale areas orange yellow. Limits of variation shown. Adults captured July 5–13. MI, WI, MN. Syst: Miller (1985d). (3 N, 3 Gf, top photo specimen *O. murina* lectotype, bottom, *O. major* lectotype)

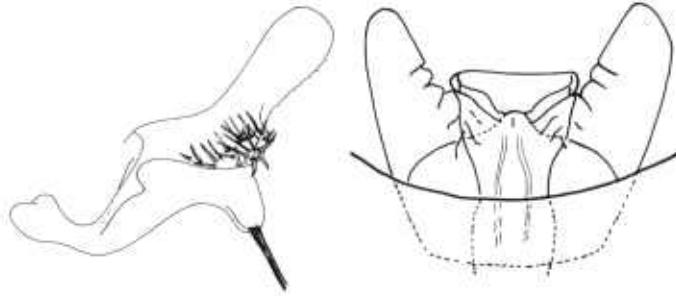


*Olethreutes carolana* (McDunnough)

Forewing 6.0 to 7.0 mm long, dark markings yellowish brown or brownish black. Adults captured June 6–27. MI, MN. Syst: Heinrich (1926). (5 N, 4 Gf, T)

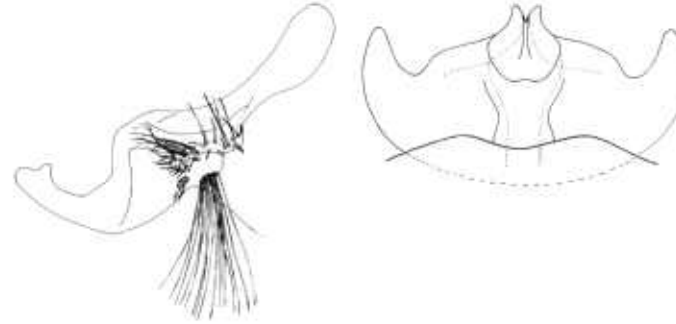
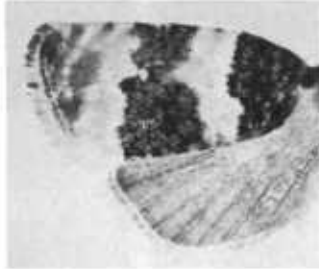






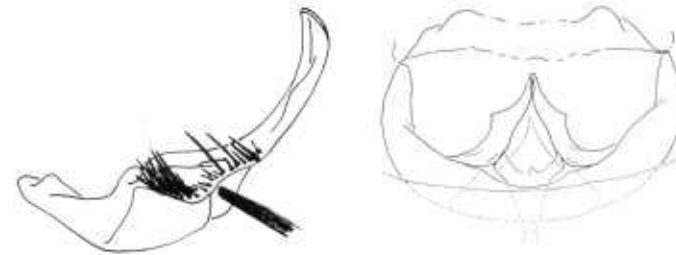
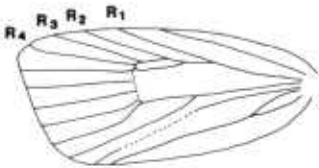
*Olethreutes glaciana* (Möschler)

Forewing 5.0 to 8.0 mm long, dark markings yellowish brown or brownish black. Adults captured June 2–October 17. MI, WI, MN. Larva feeds in rolled leaves of *Betula*, *Populus*, *Acer*, others. Syst: Miller (1985d). Biol: Prentice (1966). (41 N, 22 Gm, 6 Gf, T)



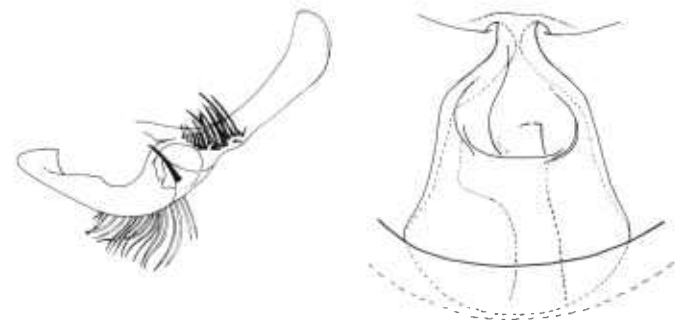
*Olethreutes bipartitana* (Clemens)

Forewing 6.0 to 9.0 mm long, dark markings grayish yellowish brown or brownish black. Adults captured May 23–September 6. MI, WI, MN. Larva feeds on *Spermolepis*. Syst and Biol: Heinrich (1926). (73 N, 13 Gm, 11 Gf, T)



*Olethreutes trinitana* (McDunnough)

Forewing 7.0 to 7.5 mm long, dark areas and markings yellowish brown or brownish black. Adults captured July 2–28. MI, WI. Syst: McDunnough (1931). (5 N, 1 Gm, 4 Gf, T)



*Olethreutes cespitana* (Hübner)

Forewing 5.5 to 8.0 mm long, dark markings yellowish brown. Adults captured May 26–September 16. MI, WI, MN. Bivoltine. Larva feeds in tied leaves of *Trifolium*, *Fragaria*, *Populus*, others. Syst: Heinrich (1926). Biol: Bennett (1961). (331 N, 29 Gm, 9 Gf)

## Genus *Hedya*

**Both sexes.** Thorax with posterior tuft. Forewing termen convex or straight, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $Rs$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindmost tibia without dilated or tufted scaling, with basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Uncus developed; socius present; valval  $Sc_2$  not on a projecting digitus; vesica without cornuti.

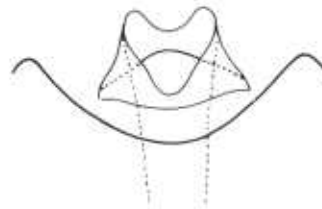
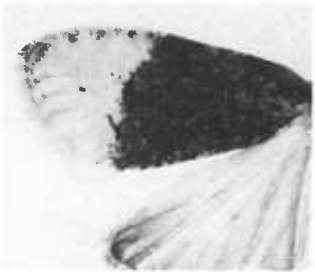
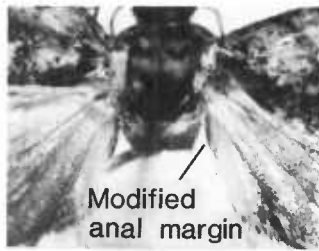
**Female.** Ductus bursae sclerotized only near genital opening; corpus bursae with two thornlike or scobinate signa.

**Comments.** Five Nearctic species of *Hedya* are known. Among the species treated here, *H. ochroleucana* also occurs in the Palearctic. My attempts to confirm the presence of *H. nubiferana* (Haworth) in the region (Strickler and Whalon 1985) were unsuccessful.



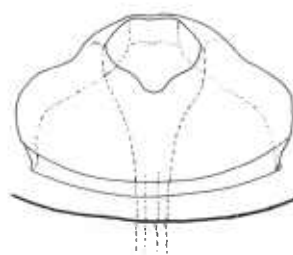
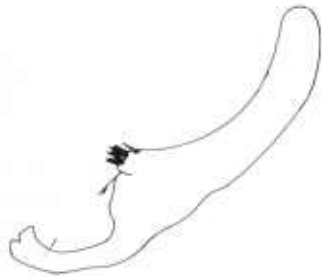
*Hedya separatana* (Kearfott)

Forewing 5.0 to 7.0 mm long, dark areas brownish black or grayish brown. Adults captured May 23–August 22. MI. Bivoltine. Larva feeds in rolled leaves of *Rosa*, *Rubus*, *Prunus*, others. Syst: Heinrich (1926). Biol: Chapman and Lienk (1971). (16 N, 7 Gm, 3 Gf, T)



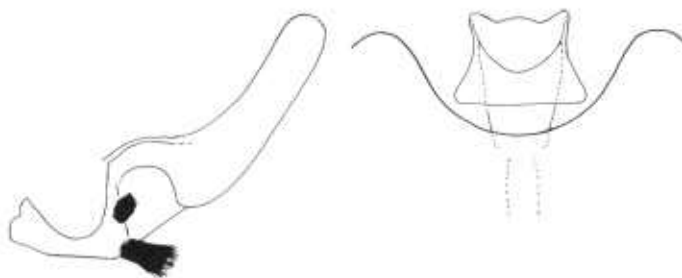
*Hedya ochroleucana* (Frölich)

Forewing 7.5 to 9.5 mm long, dark areas brownish black or grayish brown. Adults captured June 2–September 6. MI, WI, MN. Larva feeds in rolled leaves of *Rosa*, *Pyrus*. Syst: Heinrich (1926). Biol: Prentice (1966). (48 N, 25 Gm, 2 Gf)



*Hedya chionosema* (Zeller)

Forewing 6.5 to 8.0 mm long, dark markings grayish brown or brownish black. Adults captured June 14–August 14. MI, WI. Univoltine. Larva feeds on buds and leaves of *Pyrus*, *Crataegus*, *Amelanchier*, others. Syst: Heinrich (1926). Biol: Chapman and Lienk (1971). (22 N, 3 Gm, 3 Gf)



*Hedya cyanana* (Murtfeldt)

Forewing 6.5 to 7.0 mm long, dark areas brownish black. Adults captured June 16-30. MI, MN. Larva feeds in rolled leaves of *Cirsium*, *Rosa*. Syst: Heinrich (1926). Biol: Marshall and Musgrave (1937). (6 N, 4 Gm, 1 Gf, photo specimen Boone Co., MO)

## Genus *Evora*

**Both sexes.** Thorax with posterior tuft. Forewing termen straight,  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  not approximate,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating at or beyond distal three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin modified into a sclerotized ridge. Hindmost tibia without dilated or tufted scaling, with basal hair pencil. Abdominal segment 1 without paired ventrolateral papilliform scale pockets. Uncus developed; socius present; valval outer surface with spinelike setae,  $Sc_1$  at or near base of cucullus,  $Sc_2$  not on a projecting digitus; vesica without cornuti.

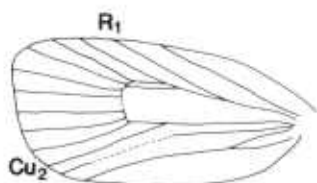
**Female.** Sternum 7 emarginate posteriorly. Lamella antevaginalis developed, not wider than lamella postvaginalis in anteroposterior orientation. Ductus bursae sclerotized only near genital opening; corpus bursae without signa.

**Comments.** *Evora* appears to be monotypic.



*Evora hemidesma* (Zeller)

Forewing 6.5 to 9.0 mm long, pale and dark areas brown. Adults captured June 10-September 2. MI, WI, MN. Multivoltine. Larva feeds in rolled *Spiraea* leaves. Syst: Heinrich (1926). Biol: Roberts (1966). (62 N, 5 Gm, 8 Gf, T, photo specimen Page Co., IA)



## Tribe Eucosmini

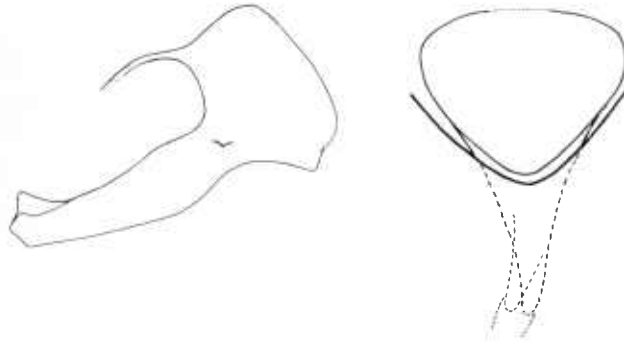
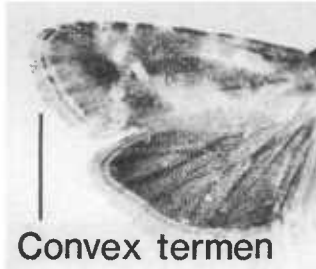
### Genus *Rhyacionia*

**Both sexes.** Forewing without raised scale tufts, termen straight or slightly convex, apex not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate,  $M_2$  and  $M_3$  connate,  $M_2$ ,  $M_3$ , and  $Cu_1$  remote at termen. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; valva not divided, without rudimentary clasper, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin.

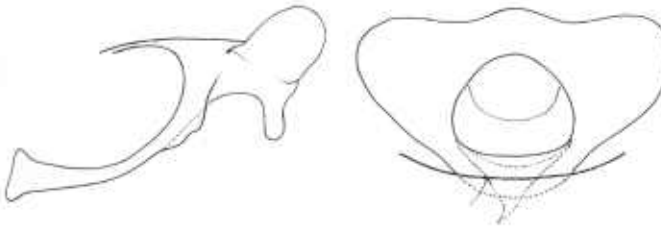
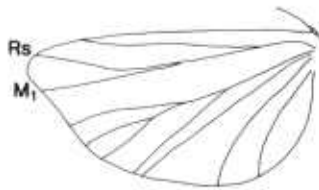
**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Lamella antevaginalis not conical, lamella postvaginalis present. Ductus bursae sclerotized one-third or less its length from genital opening; corpus bursae without sclerotized sides, with two thornlike signa or without signa.

**Comments.** More than 20 Nearctic species of *Rhyacionia* are known. Among the species treated here, *R. buoliana* also occurs in the Palearctic. It was introduced to the Nearctic before 1914. Powell and Miller (1978) reviewed the species of *Rhyacionia*.



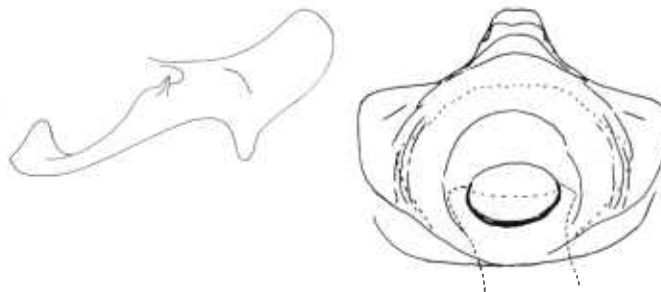
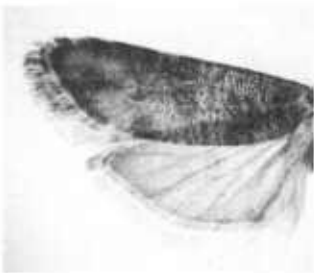
*Rhyacionia buoliana* (Denis and Schiffermüller). European pine shoot moth

Forewing 7.5 to 11.0 mm long, dark areas orange. Adults captured June 3–July 23. MI, WI. Univoltine. Larva feeds in *Pinus* terminals. Syst: Powell and Miller (1978). Biol: Miller (1967). (158 N, 3 Gm, 4 Gf)



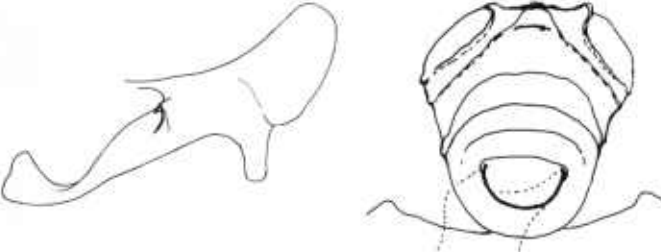
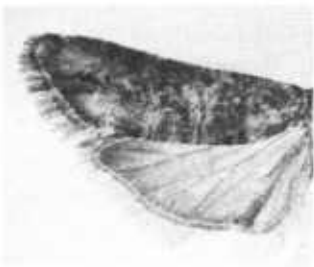
*Rhyacionia adana* Heinrich

Forewing 5.5 to 7.0 mm long, pale apical areas orange. No capture dates; all adults reared. MI. Univoltine. Larva feeds in terminals of *Pinus resinosa*, *P. banksiana*, *P. sylvestris*. Syst: Powell and Miller (1978). Biol: Martin (1960). (12 N, 4 Gm, 3 Gf, T)



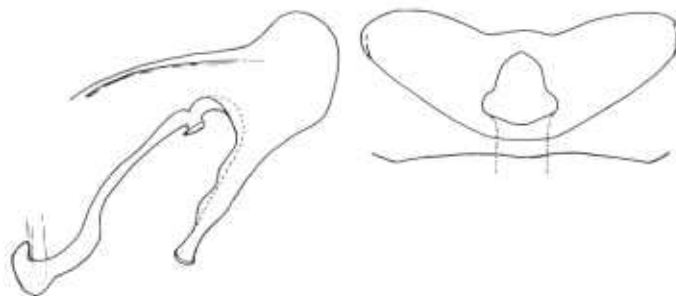
*Rhyacionia busckana* Heinrich

Forewing 8.0 mm long, pale apical areas orange. Male antennal pecten subequal in length to antennal segments. Adult captured May 25. MI. Larva feeds in terminals of *Pinus resinosa*, *P. sylvestris*. Syst and Biol: Miller (1985b). (1 N, 1 Gf, T, photo specimen Simcoe Co., ON)



*Rhyacionia granti* Miller

Forewing 6.0 to 8.0 mm long, pale apical areas orange. Male antennal pecten twice or more as long as antennal segments. Adults captured April 28–May 25. MI, WI. Larva feeds in *Pinus banksiana* terminals. Syst and Biol: Miller (1985b). (13N, 3 Gm, 6 Gf, T, photo specimen Dauphin Co., PA)



*Rhyacionia sonia* Miller

Forewing 6.5 mm long, pale areas orange. Adult captured May 25. MI. Larva feeds in *Pinus banksiana* terminals. Syst: Powell and Miller (1978). Biol: McDowell and Wong (1962). (1 N, 1 Gm, T, photo specimen holotype)

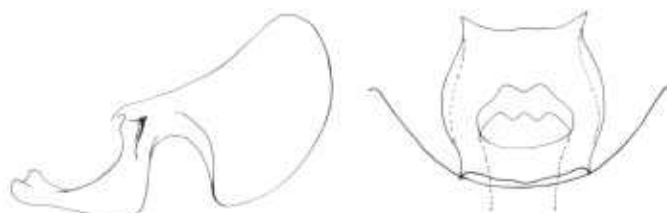
## Genus *Retinia*

**Both sexes.** Forewing without raised scale tufts, termen straight or slightly concave, apex not falcate, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate or approximate,  $M_2$  and  $M_3$  connate,  $M_2$ ,  $M_3$ , and  $Cu_1$  remote at termen. Hindwing Rs and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius present; valva not divided, rudimentary clasper present, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Ductus bursae sclerotized less than three-fourths its length from genital opening; corpus bursae without sclerotized sides, with two thorn- or finlike signa.

**Comments.** Fifteen Nearctic species of *Retinia* are known. The validity of the name *Retinia* for this genus (formerly *Petrova*) was documented by Leraut (1978). The pitch-blister-making group of *Retinia* was reviewed by Miller (1978b).

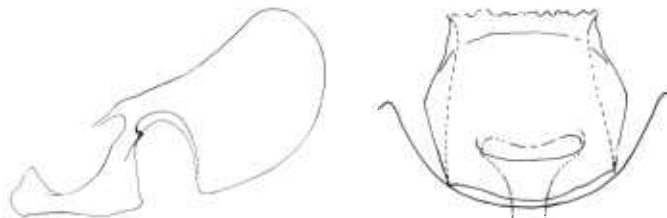
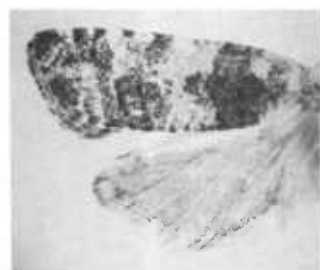


*Retinia albicapitana* (Busck).  
Northern pitch twig moth

Forewing 5.0 to 10.5 mm long, dark areas orange or brown. Adults captured April 28–July 27. MI, WI, MN. Semivoltine. Larva feeds on *Pinus* terminals and twigs beneath pitch blister. Syst: Miller (1978b). Biol: Turnock (1953). (154 N, 5 Gm, 7 Gf, T)

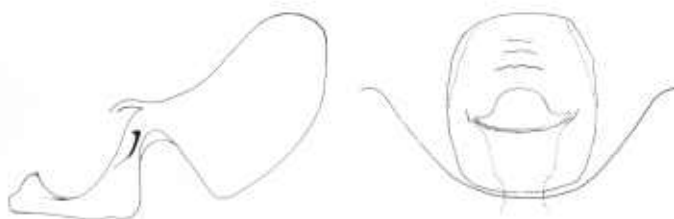
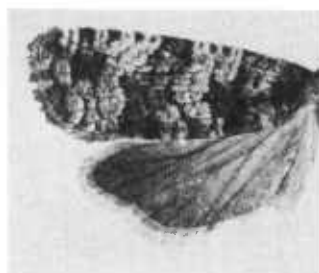


Finger- or ribbonlike socius



*Retinia metallica* (Busck)

Forewing 7.0 to 10.0 mm long, dark areas brown or purplish blue. No capture dates; all adults reared. MN. Univoltine. Larva feeds on *Pinus* terminals beneath pitch blister. Syst and Biol: Miller (1978b). (30 N, 2 Gm, 2 Gf, T)



*Retinia mafica* (Miller)

Forewing 6.5 to 8.0 mm long, dark areas grayish brown or brownish black. Adults captured May 7–June 13. MI, WI. Larva feeds on *Pinus*. Syst and Biol: Miller (1978b). (10 N, 6 Gm, 1 Gf, T, photo specimen holotype)



*Retinia gemistrigulana* (Kearfott)

Forewing 7.5 to 10.5 mm long, dark markings grayish yellowish brown. Adults captured July 1–August 7. MI, WI. Syst: Heinrich (1923b). (24 N, 14 Gm, 2 Gf, T)



*Retinia pallipennis*  
(McDunnough)

Forewing 6.5 to 8.5 mm long, dark markings brown or brownish orange. Adults captured May 10–30. MI, WI, MN. Larva feeds in *Pinus* buds. Syst: McDunnough (1938). Biol: Butcher and Hodson (1949). (10 N, 6 Gm, 2 Gf, T)

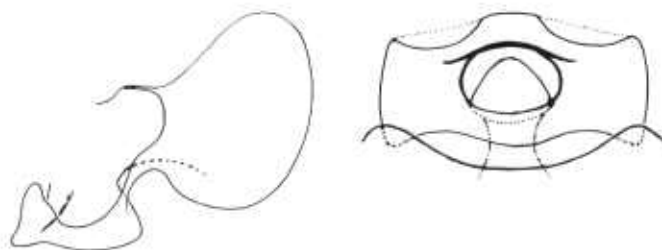
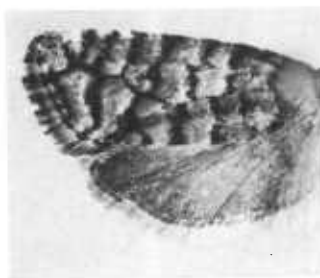
## Genus *Barbara*

**Both sexes.** Forewing without raised scale tufts, termen straight or slightly concave, apex not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate or approximate,  $M_2$  and  $M_3$  connate,  $M_2$ ,  $M_3$ , and  $Cu_1$  remote at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius present; valva not divided, without rudimentary clasper, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 not deeply emarginate, neither inflected nor overlapping ostium bursae. Ductus bursae sclerotized one-half its length from genital opening; corpus bursae without sclerotized sides, with two thorn- or finlike signa.

**Comments.** Three Nearctic species of *Barbara* are known.



*Barbara mappana* Freeman

Forewing 7.0 mm long, pale areas brownish gray. Adults captured June 25. WI. Larva feeds in cones of *Abies balsamea*, *Picea*. Syst: Freeman (1941). Biol: Prentice (1966). (4 N, 1 Gm, 1 Gf, T)

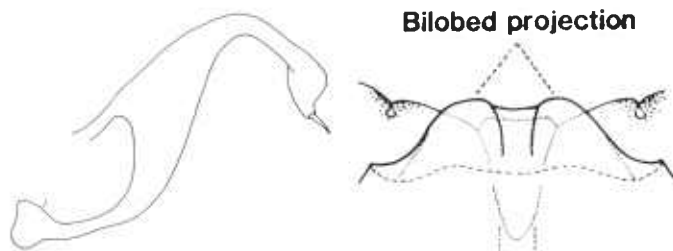
## Genus *Spilonota*

**Both sexes.** Forewing without raised scale tufts, termen straight or slightly concave, apex not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  connate,  $M_2$  and  $M_3$  approximate at base,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius present; valva not divided, without rudimentary clasper, outer surface lacking spinelike setae, with a thin spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 inflected and overlapping ostium bursae, with median bilobed projection. Ductus bursae sclerotized near middle; corpus bursae without sclerotized sides, with single thornlike signum.

**Comments.** Two Nearctic species of *Spilonota* are known; both also occur in the Palearctic. *Spilonota ocellana* was introduced to the Nearctic before 1840.



**Bilobed projection**

*Spilonota ocellana* (Denis and Schiffermüller). Eyespotted bud moth

Forewing 6.0 to 7.5 mm long, dark areas and markings grayish yellowish brown or brownish black. Adults captured June 8–September 1. MI, WI, MN. Univoltine. Larva feeds on buds, flowers, leaves of *Pyrus malus*, *Prunus*, *Rubus*, others. Syst: Heinrich (1923b). Biol: Oatman *et al.* (1962). (98 N, 4 Gm, 5 Gf)



## Genus *Phaneta*

**Both sexes.** Forewing without raised scale tufts, termen straight or slightly concave, apex not falcate, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate,  $M_2$  and  $M_3$  separate,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked or united.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius finger- or

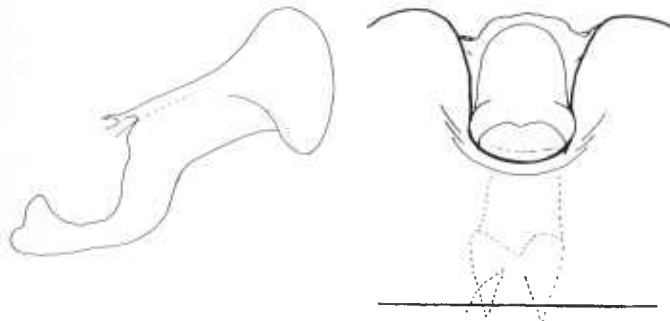
ribbonlike, not heavily sclerotized; valva not divided, without rudimentary clasper, sacculus not densely clothed with spinelike setae, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 deeply emarginate around ostium bursae, neither inflected nor overlapping ostium bursae. Lamella antevaginalis not conical, lamella postvaginalis not recessed, ostium bursae not elongate. Ductus bursae sclerotized near middle; corpus bursae without sclerotized sides, with dual thorn- or finlike signa.

**Comments.** Among the species treated here, all except *Phaneta ornatula* have forewing  $R_2$  originating nearer  $R_3$  than  $R_1$ .

Nearly 100 Nearctic species of *Phaneta* are known.

In the *P. radiatana* species group (*radiatana*, *essexana*, *awemeana*, *umbrastriana*, *formosana*), intraspecific variability is high and species limits are ill defined at present. Some specimens of this group are not easily identified to species. Members of the group are treated here according to the synopsis given by Miller (1983a).



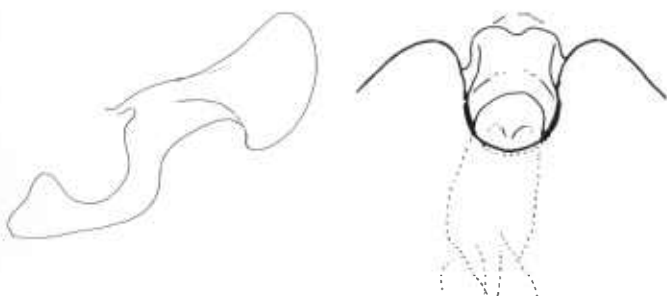
*Phaneta radiatana*  
(Walsingham)

Forewing 9.0 to 11.5 mm long, pale areas in male pale orange yellow, in female yellowish brown. Adults captured May 20–June 24. MI. Larva feeds in *Solidago* stems. Syst and Biol: Heinrich (1923b). (22 N, 4 Gm, 1 Gf, T)



*Phaneta essexana* (Kearfott)

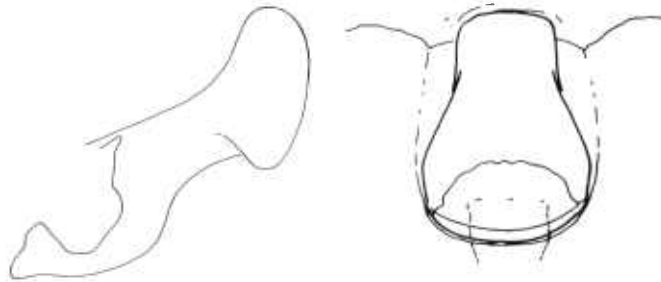
Forewing 9.5 to 11.5 mm long, pale areas in male pale orange yellow, in female yellowish brown. Adults captured May 13–June 3. MI. Larva feeds in *Aster* stems. Syst: Heinrich (1923b). Biol: MacKay (1962). (8 N, 3 Gm, T)



*Phaneta awemeana* (Kearfott)

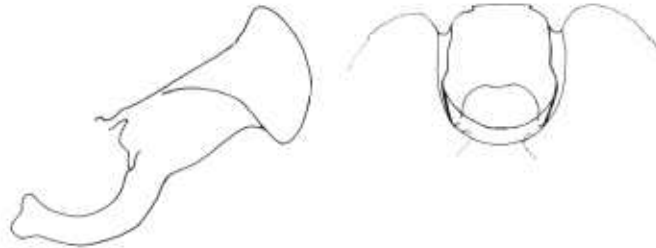
Forewing 8.0 to 10.0 mm long, pale areas in male pale orange yellow, in female yellowish brown. Adults captured May 5–June 29. MI, WI, MN. Syst: Heinrich (1923b). (33 N, 5 Gm, 1 Gf, T)





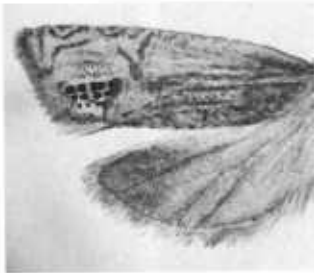
*Phaneta umbrastriana* (Kearfott)

Forewing 7.5 to 10.0 mm long, pale areas in male pale orange yellow, in female yellowish brown. Adults captured May 11–June 23. MI, WI, MN. Syst: Miller (1983a). (49 N, 14 Gm, 1 Gf, T)



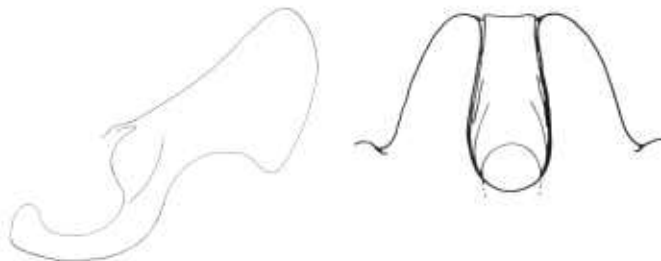
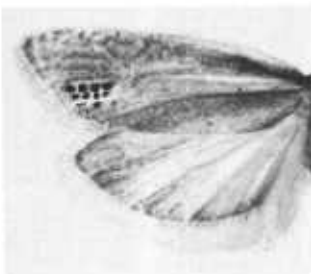
*Phaneta formosana* (Clemens)

Forewing 8.0 to 9.5 mm long, dark areas brown. Adults captured May 30–July 5. MI, WI. Larva feeds in *Solidago* terminals. Syst: Heinrich (1923b). Biol: Putman (1942). (26 N, 3 Gm, 6 Gf, T)



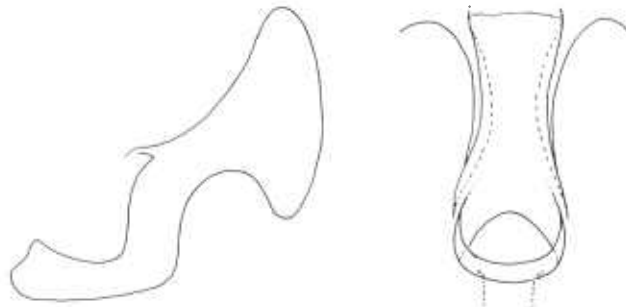
*Phaneta autumnana* (McDunnough)

Forewing 7.0 to 8.5 mm long, medium pale areas orange yellow. Adults captured August 28–October 13. MI, WI. Syst: Miller (1971). (23 N, 19 Gm, 1 Gf, T)



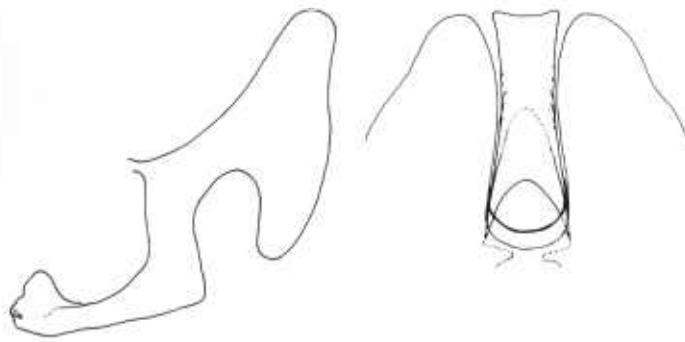
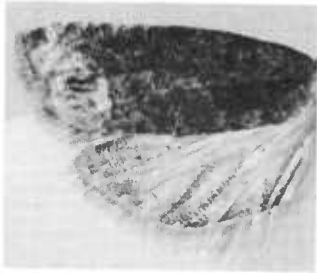
*Phaneta verna* Miller

Forewing 7.0 to 8.0 mm long, medium pale areas orange yellow. Adults captured May 15–29. MI. Syst: Miller (1971). (8 N, 5 Gm, 1 Gf, T)



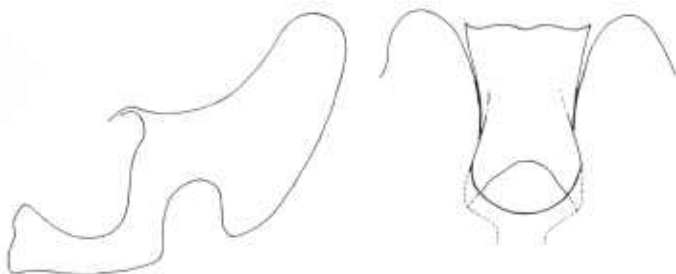
*Phaneta raracana* (Kearfott)

Forewing 5.5 to 6.5 mm long, dark areas brown. Adults captured July 22–August 11. MI. Larva feeds on *Solidago*. Syst and Biol: Heinrich (1923b). (10 N, 3 Gm, 1 Gf, T)



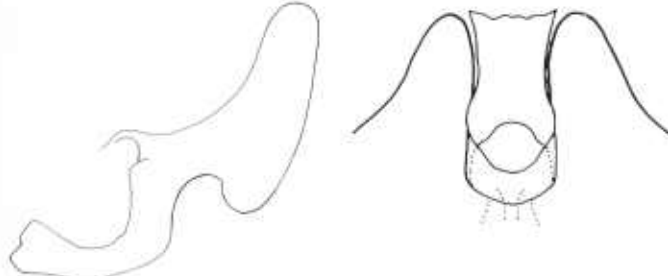
*Phaneta ochroterminana*  
(Kearfott)

Forewing 5.0 to 6.5 mm long, dark areas bluish black. Adults captured July 12–August 29. MI, WI, MN. Larva feeds on *Solidago* flower heads. Syst: Heinrich (1923b). Biol: Putman (1942). (24 N, 8 Gm, 2 Gf, T)



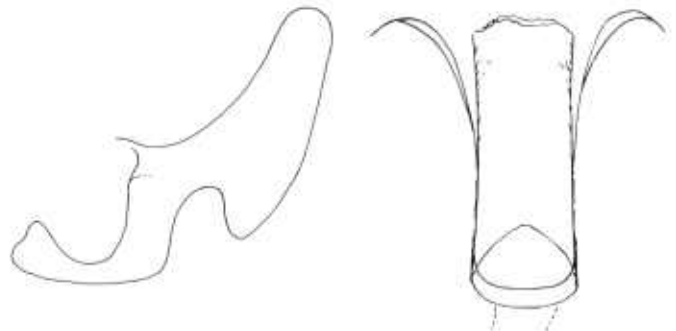
*Phaneta marmontana* (Kearfott)

Forewing 6.0 to 7.5 mm long, dark areas of basal three-quarters grayish brown. Adults captured June 6–July 23. MI, WI. Syst: Heinrich (1923b). (9 N, 5 Gm, 3 Gf, T)



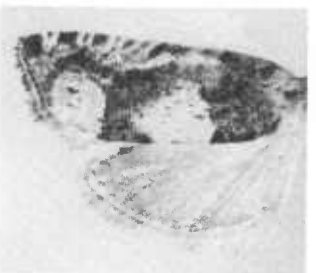
*Phaneta tomonana* (Kearfott)

Forewing 5.5 to 7.5 mm long, dark markings brownish black. Adults captured August 12–September 7. MI, WI. Larva feeds on *Aster* flower heads. Syst: Heinrich (1923b). Biol: Putman (1942). (19 N, 5 Gm, 2 Gf, T)



*Phaneta parmatana* (Clemens)

Forewing 5.0 to 7.0 mm long, size of pale dorsal spot variable; dark areas grayish brown or brownish black. Limits of variation shown. Adults captured May 20–September 8. MI, WI, MN. Larva feeds on *Aster* flower heads. Syst and Biol: Miller (1983a). (85 N, 38 Gm, 8 Gf, T)





**Female  
unknown**

*Phaneta modernana*  
(McDunnough)

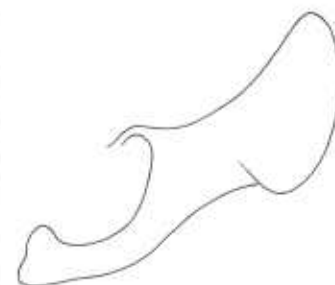
Forewing 9.5 mm long, pale areas grayish yellowish brown. Adult captured June 8. MI. Syst: McDunnough (1925a). (1 N, 1 Gm, T)



**Female  
unknown**

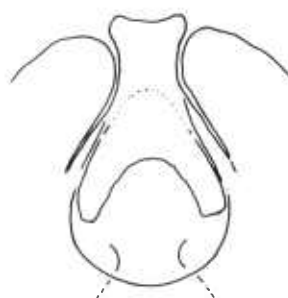
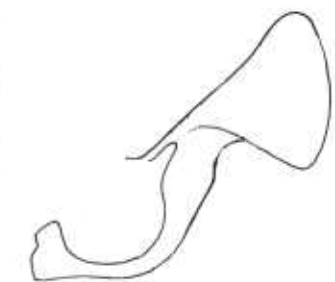
*Phaneta convergana*  
(McDunnough)

Forewing 7.5 mm long, pale areas light grayish yellowish brown. Adult captured May 21. MI. Syst: McDunnough (1925a). (1 N, 1 Gm, T)



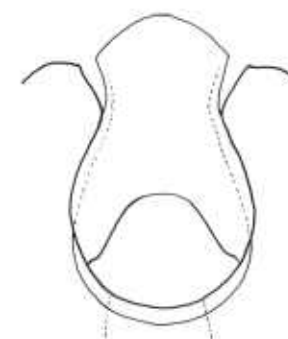
*Phaneta ornatula* (Heinrich)

Forewing 6.0 to 7.5 mm long, dark markings brownish black. Adults captured June 26–August 27. MI, WI, MN. Larva feeds in *Lactuca* flower heads. Syst: Heinrich (1924). Biol: Putman (1942). (19 N, 5 Gm, 2 Gf, T)



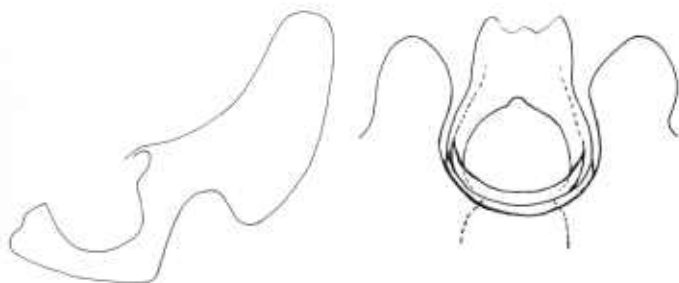
*Phaneta vernalana*  
(McDunnough)

Forewing 8.5 mm long, dark markings grayish yellowish brown. Adult captured May 24. MI. Syst: McDunnough (1942). (1 N, 1 Gm, T)



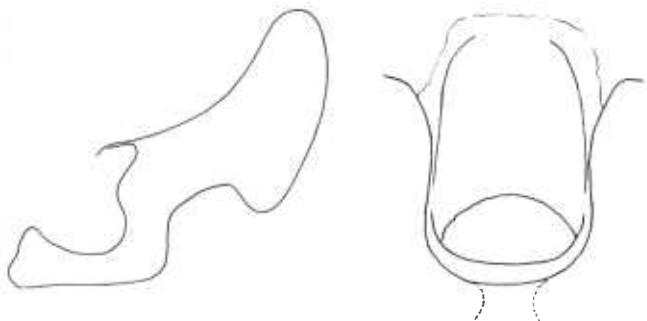
*Phaneta tarandana* (Möschler)

Forewing 10.5 to 11.5 mm long, pale areas pale yellow. Adults captured July 27–September 20. MI. Syst: Heinrich (1923b). (3 N, 2 Gm, 1 Gf)



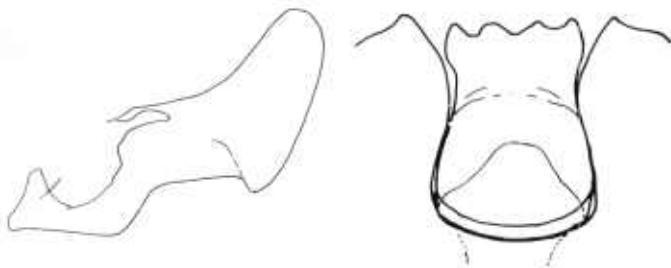
*Phaneta clavana* (Fernald)

Forewing 6.5 to 8.0 mm long, dark markings yellowish brown or brownish black. Adults captured July 25–August 16. MI, MN. Syst: Heinrich (1923b). (13 N, 3 Gm, 1 Gf, T)



*Phaneta argenticostana* (Walsingham)

Forewing 8.0 to 10.5 mm long, dark areas and markings yellowish brown. Adults captured May 13–June 25. MI, MN. Larva feeds on *Artemisia*. Syst: Heinrich (1923b). Biol: Brown et al. (1983). (28 N, 3 Gm, 2 Gf, T)



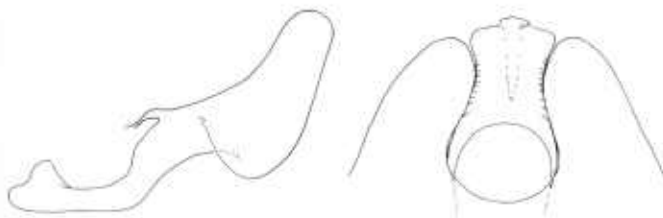
*Phaneta dorsiatomana* (Kearfott)

Forewing 9.0, medium light areas yellowish brown. No capture date. MN. Larva feeds on *Artemisia*. Syst: Heinrich (1923b). Biol: Brown et al. (1983). (1 N, 1 Gm, T, photo specimen Emmons Co., ND)



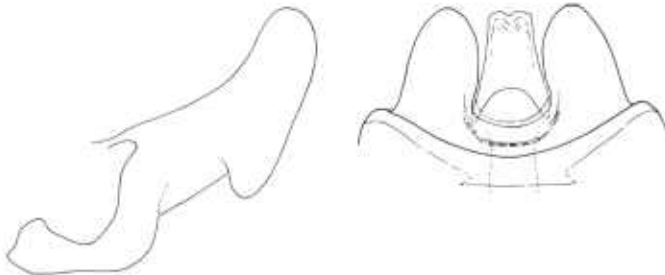
*Phaneta striatana* (Clemens)

Forewing 6.0 to 8.0 mm long, dark areas and markings grayish brown. Adults captured May 20–August 8. MI, WI, MN. Syst: Heinrich (1923b). (73 N, 5 Gm, 5 Gf, T)



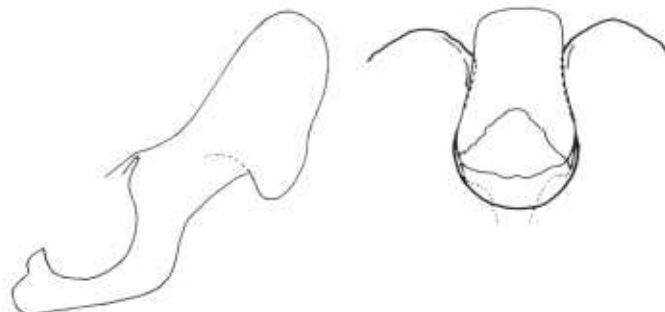
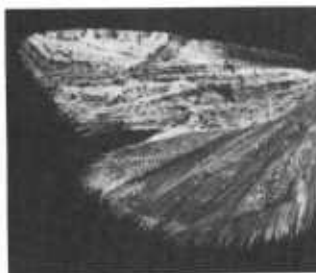
*Phaneta pallidicostana* (Walsingham)

Forewing 7.0 to 8.5 mm long, white with yellowish white or yellowish gray markings. Adults captured June 22–July 24. MN. Syst: Heinrich (1923b). (5 N, 2 Gm, 2 Gf, T)



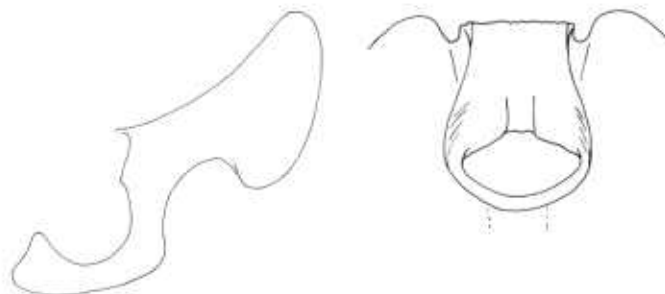
*Phaneta kiscana* (Kearfott)

Forewing 6.0 mm long, dark areas and markings yellowish brown or brownish black. Adult captured June 29. MI. Syst: Heinrich (1923b). (1 N, 1 Gm, T)



*Phaneta montanana* (Walsingham)

Forewing 7.5 to 10.5 mm long, pale areas pale orange yellow. Adults captured May 23–August 25. MI, MN. Larva feeds in *Artemisia* roots. Syst and Biol: Heinrich (1923b). (8 N, 5 Gm, T)



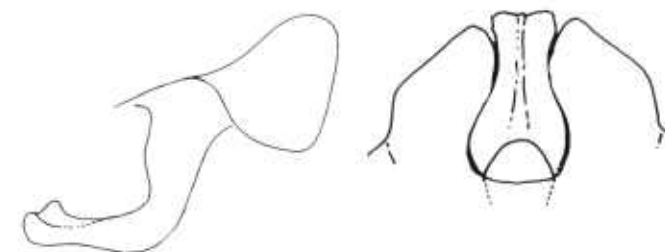
*Phaneta olivaceana* (Riley)

Forewing 7.5 to 9.0 mm long, pale areas pale orange yellow. Adults captured June 24–August 3. MI. Larva feeds on *Solidago*. Syst: Heinrich (1923b). (9 N, 2 Gm, 3 Gf, T)



*Phaneta ochrocephala* (Walsingham)

Forewing 6.0 to 8.0 mm long, dark areas purplish gray or grayish purple. Adults captured July 31–August 22. MI, MN. Univoltine. Larva feeds in *Xanthium* seeds. Syst: Miller (1983a). Biol: Hare (1977). (10 N, 4 Gm, 2 Gf, T)



*Phaneta ambodaidaleia* Miller

Forewing 9.5 to 10.0 mm long, grayish yellowish brown streaked with light yellowish brown. Adults captured April 26. MI. Syst: Miller (1983a). (2 N, 2 Gm, T)

## Genus *Eucosma*

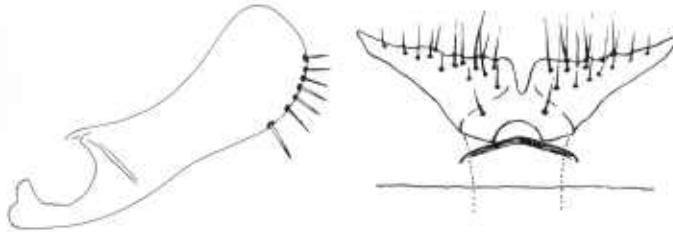
**Both sexes.** Forewing without raised scale tufts, termen straight or concave,  $R_1$  originating well before middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate,  $M_2$  and  $M_3$  separate. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked or united.

**Male.** Antenna not notched near base. Forewing with costal fold. Hindwing without upper-surface melanic sex scaling. Socius finger- or ribbonlike, not heavily sclerotized; valva not divided, sacculus not densely clothed with spinelike setae, without rudimentary clasper, outer surface lacking spinelike setae, with or without thin spinelike seta on cucullus at lower margin, midpoint of neck constriction at or beyond midpoint between valval base and apex.

**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Lamella antevaginalis not conical, lamella postvaginalis present. Ductus bursae sclerotized less than two-thirds its length from genital opening; corpus bursae with one or two thorn- or finlike signa.

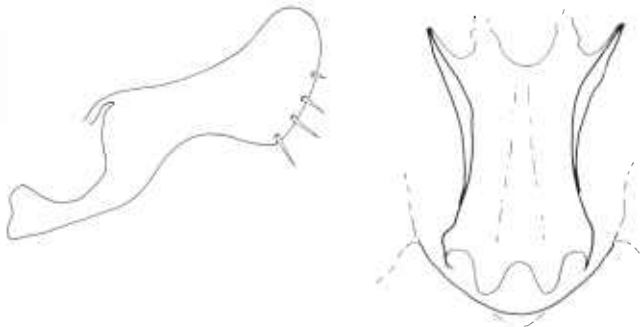
**Comments.** Nearly 150 Nearctic species of *Eucosma* are known.

I was unable to confirm Fernekes' (1906) report of *E. canana* (Walsingham) in the region, or Prentice's (1966) report of *E. ridingsana* (Robinson) boring in *Pinus* branchlets.



*Eucosma robinsonana* (Grote)

Forewing 5.0 to 8.5 mm long, dark markings yellowish brown. Ovipositor with ventral extensions of papillae anales. Adults captured June 2–August 7. MI, MN. Syst: Heinrich (1923b). (43 N, 4 Gm, 5 Gf, T)



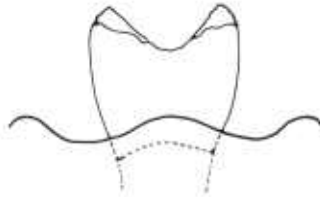
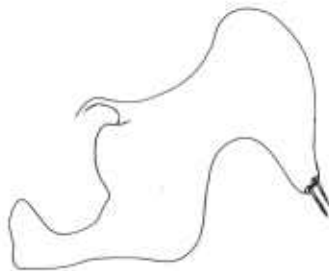
*Eucosma ridingsana* (Robinson)

Forewing 8.0 mm long, dark markings yellowish brown. Adult captured Aug 13. MN. Larva feeds in *Gutierrezia* roots. Syst: Heinrich (1923b). Biol: Hetz and Werner (1979). (1 N, T, photo specimen lectotype)



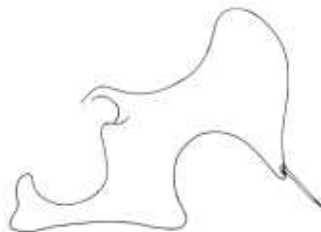
*Eucosma heathiana* Kearfott

Forewing 8.5 mm long, dark areas and markings yellowish brown. Ovipositor with ventral extensions of papillae anales. Adults captured July 3–8. MI, MN. Syst: Heinrich (1923b). (2 N, 2 Gm, T)



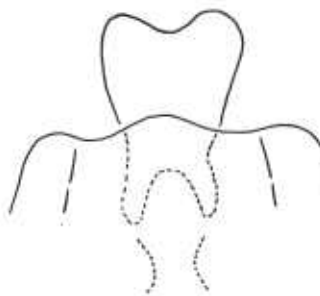
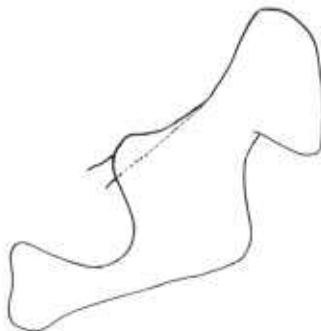
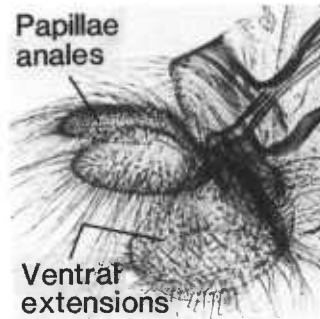
*Eucosma morrisoni*  
(Walsingham)

Forewing 7.0 to 9.5 mm long, dark markings yellowish brown or grayish yellowish brown. Ovipositor with ventral extensions of papillae anales. Adults captured July 14–August 18. MI, MN. Syst: Heinrich (1923b). (28 N, 6 Gm, 3 Gf, T)



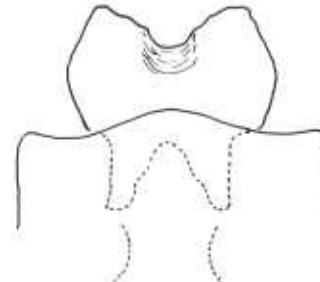
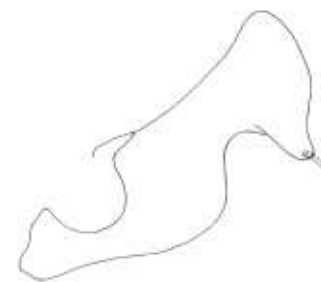
*Eucosma agricola*  
(Walsingham)

Forewing 6.5 to 9.5 mm long, dark markings grayish yellowish brown or brownish black. Ovipositor with ventral extensions of papillae anales. Adults captured May 23–July 17. MI, WI, MN. Larva feeds on *Artemisia*. Syst: Miller (1974). Biol: Brown *et al.* (1983). (103 N, 19 Gm, 5 Gf, T)



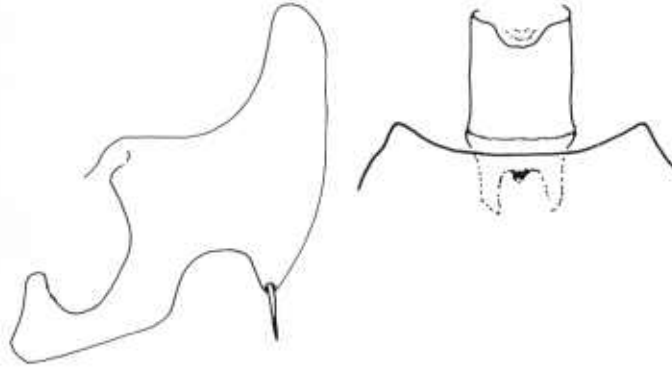
*Eucosma smithiana*  
(Walsingham)

Forewing 7.0 to 8.5 mm long, pale areas yellowish white. Male uncus unevenly rounded in outline. Ovipositor with ventral extensions of papillae anales. Adults captured June 14–July 16. MN. Larva feeds in *Chrysanthemum* roots. Syst and Biol: Miller (1974). (10 N, 3 Gm, 6 Gf, T)



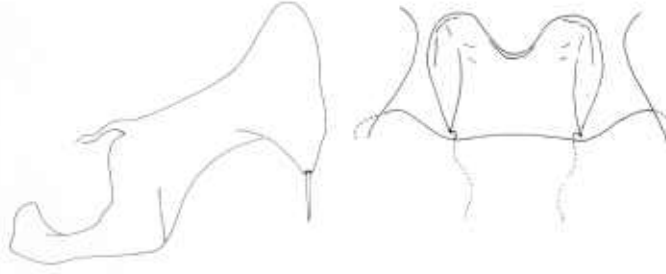
*Eucosma barbara* Miller

Forewing 7.0 to 8.5 mm long, pale areas yellowish white. Male uncus evenly rounded in outline. Ovipositor with ventral extensions of papillae anales. Adults captured July 3–August 8. MN. Syst: Miller (1974). (12 N, 5 Gm, 7 Gf, T)



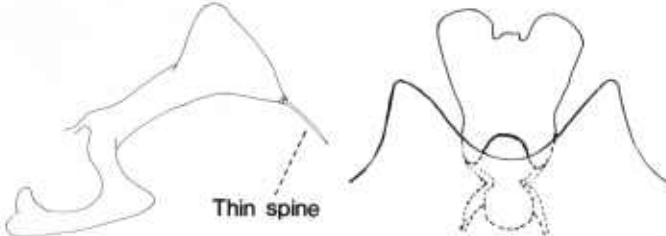
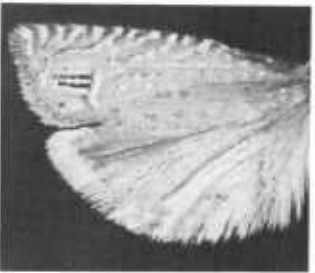
*Eucosma rindgei* Miller

Forewing 7.0 to 8.0 mm long, dark markings blackish brown. Ovipositor with ventral extensions of papillae anales. Adults captured June 26–August 22. MN. Syst: Miller (1985c). (17 N, 4 Gm, 4 Gf, T)



*Eucosma vagana* McDunnough

Forewing 7.0 to 9.0 mm long, dark areas and markings brown. Ovipositor with ventral extensions of papillae anales. Adults captured June 23–August 10. MI, WI, MN. Larva feeds in *Solidago* roots. Syst: Miller (1985c). Biol: Brown *et al.* (1983). (25 N, 10 Gm, 5 Gf, T)



*Eucosma albiguttana* (Zeller)

Forewing 6.5 to 8.5 mm long, medium pale areas orange yellow. Ovipositor with ventral extensions of papillae anales. Adults captured July 11–15. MI, MN. Syst: Heinrich (1923b). (12 N, 7 Gm, T)



*Eucosma graciliana* Kearfott

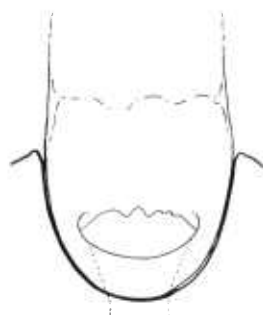
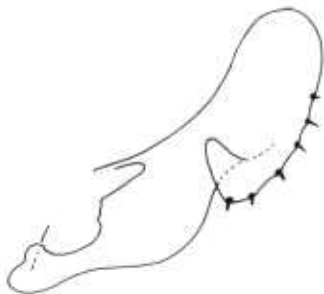
Forewing 5.0 to 6.5 mm long, medium pale areas orange yellow. Ovipositor with ventral extensions of papillae anales. Adults captured June 19–July 29. MI, WI, MN. Syst: Heinrich (1923b). (7 N, 4 Gm, T)



*Eucosma maculatana* (Walsingham)

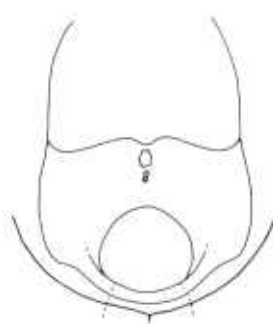
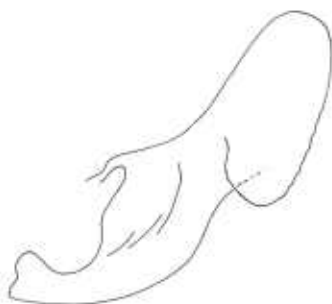
Forewing 8.0 to 8.5 mm long, dark markings brown. No capture dates. MN. Syst: Heinrich (1923b). (2 N, 1 Gm, 1 Gf, T)





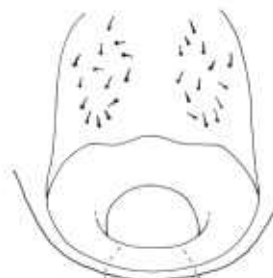
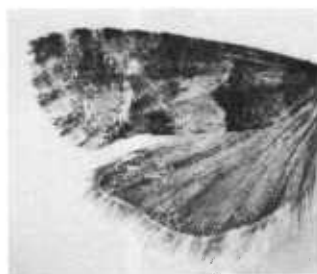
*Eucosma gloriola* Heinrich.  
Eastern pine shoot borer

Forewing 6.5 to 8.0 mm long, dark areas orange. Adults captured May 6–June 2. MI, WI, MN. Univoltine. Larva feeds in shoots of *Pinus*, *Picea*, *Pseudotsuga*. Syst and Biol: DeBoo et al. (1971). (23 N, 6 Gm, 5 Gf, T)



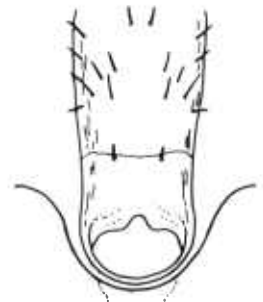
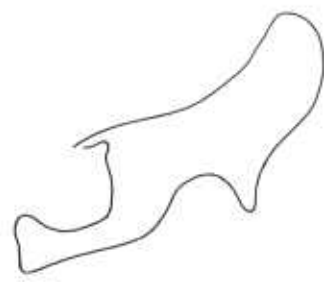
*Eucosma monitorana* Heinrich

Forewing 7.0 to 8.5 mm long, dark areas brown. Adults captured June 19–July 7. WI, MN. Univoltine. Larva feeds in cones of *Pinus resinosa*, *P. virginiana*. Syst: Powell (1968). Biol: Barras and Norris (1969). (15 N, 6 Gm, 5 Gf, T)



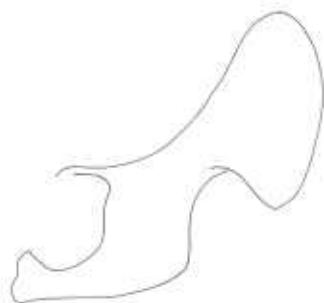
*Eucosma tocullionana* Heinrich. White pine cone borer

Forewing 5.5 to 9.0 mm long, dark areas and markings brown or brownish black. Adults captured May 31–July 14. MI, WI. Larva feeds in cones of *Pinus*, *Picea*, *Abies*. Syst and Biol: Powell (1968). (34 N, 3 Gm, 8 Gf, T)



*Eucosma matutina* (Grote)

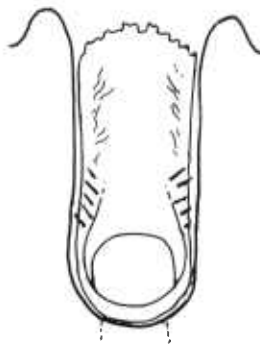
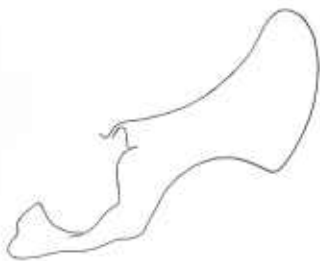
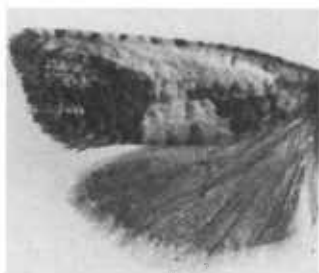
Forewing 5.0 to 8.0 mm long, dark areas and markings brownish black or grayish brown. Adults captured June 21–August 30. MI, WI, MN. Syst: Miller (1985c). (33 N, 5 Gm, 5 Gf, T)



**Female  
unknown**

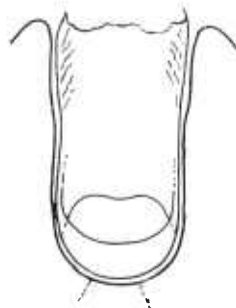
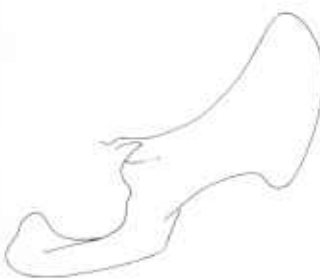
*Eucosma palabundana* Heinrich

Forewing 7.0 to 8.5 mm long, dark areas and markings grayish brown or grayish yellowish brown. Adults captured July 17–August 11. MI. Syst: Heinrich (1923b). (11 N, 5 Gm, T)



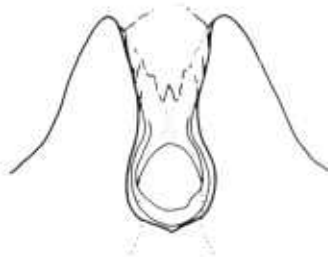
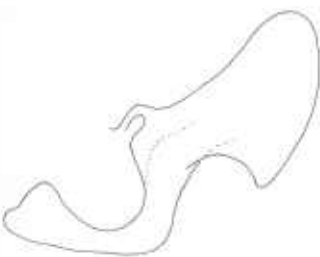
*Eucosma giganteana* (Riley)

Forewing 12.0 to 17.0 mm long, dark areas and markings yellowish brown or brownish black. Adults captured July 9–August 10. MI, WI, MN. Larva feeds in *Silphium* roots. Syst: Heinrich (1923b). Biol: Godfrey *et al.* (1987). (5 N, 1 Gm, 3 Gf, T)



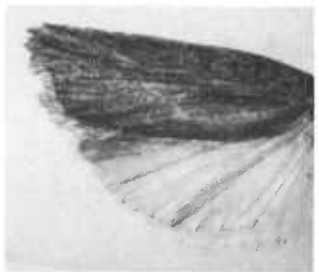
*Eucosma bipunctella* (Walker)

Forewing 16.5 to 18.0 mm long, pale areas yellow. Adults captured June 6–22. MI, WI. Larva feeds in *Silphium* roots. Syst: Heinrich (1923b). Biol: MacKay (1959). (4 N, 2 Gm, 1 Gf, T)



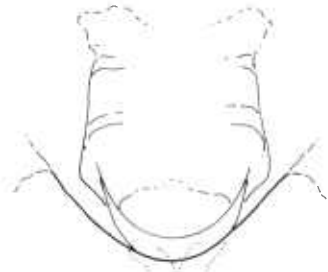
*Eucosma bilineana* Kearfott

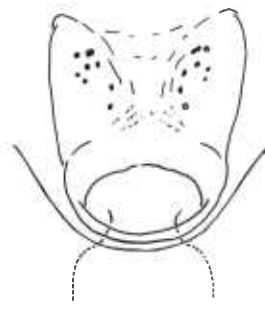
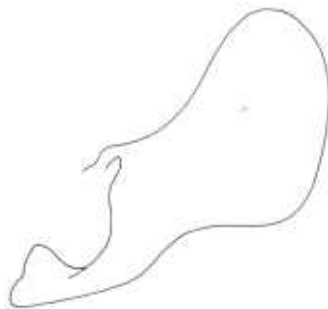
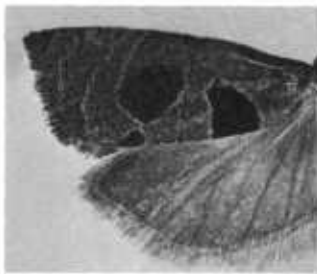
Forewing 12.5 mm long, variable pattern; dark areas and markings yellowish brown or brownish black. Limits of variation shown. Ovipositor with ventral extensions of papillae anales. Adult captured June 24. MI. Larva feeds on *Helianthus*. Syst: Heinrich (1923b). Biol: Brown *et al.* (1983). (1 N, 1 Gm, T, top photo specimen Kossuth Co., IA, bottom, Winnipeg, MB)



*Eucosma nandana* Kearfott

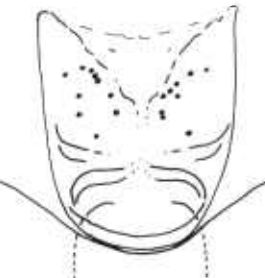
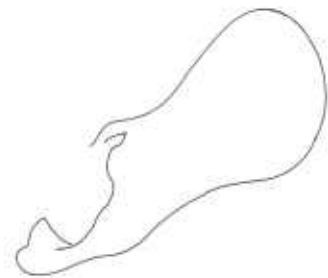
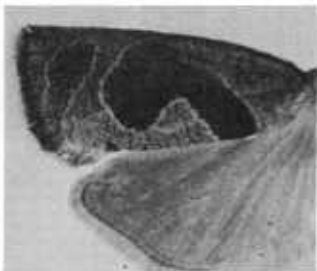
Forewing 12.0 mm long, dark areas yellowish brown. Sides of female corpus bursae sclerotized near ductus bursae. Adult captured Aug 19. MN. Syst: Heinrich (1923b). (1 N, 1 Gm, T)





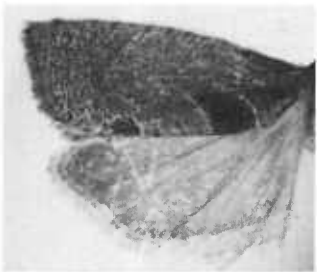
*Eucosma dorsisignatana*  
(Clemens)

Forewing 8.5 to 11.0 mm long, dorsal markings brown. Adults captured July 30–October 12. MI, MN. Larva feeds in *Solidago* roots. Syst and Biol: Miller (1985c). (63 N, 10 Gm, 9 Gf, T)



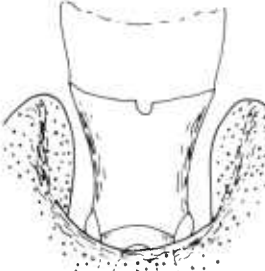
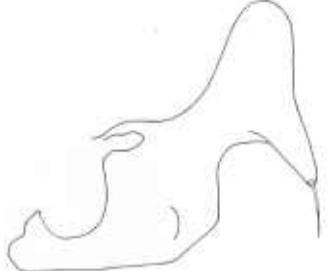
*Eucosma similiana* (Clemens)

Forewing 6.5 to 10.0 mm long, variable pattern; dorsal markings brown. Adults captured July 28–September 14. Limits of variation shown. MI, WI, MN. Univoltine. Larva feeds in *Solidago* rootstalks. Syst: Miller (1985c). Biol: Capek (1971). (45 N, 11 Gm, 9 Gf, T)



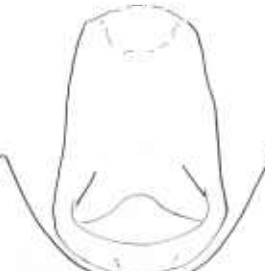
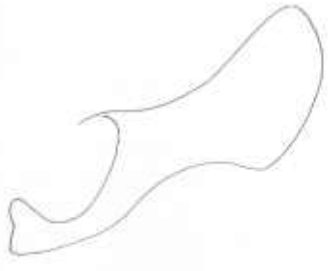
*Eucosma derelicta* Heinrich

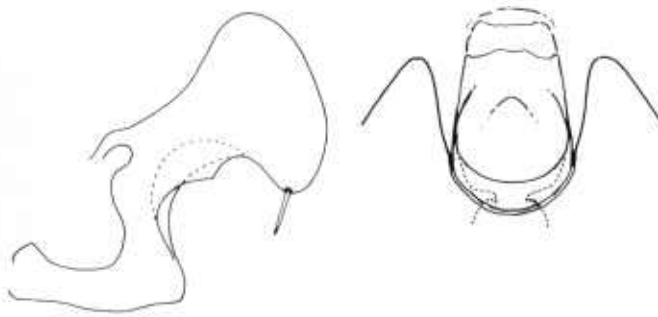
Forewing 5.5 to 8.5 mm long, dark areas yellowish brown. Adults captured July 6–August 26. MI, WI, MN. Larva feeds in *Solidago* rootstalks. Syst: Heinrich (1929). Biol: Capek (1971). (82 N, 10 Gm, 5 Gf, T)



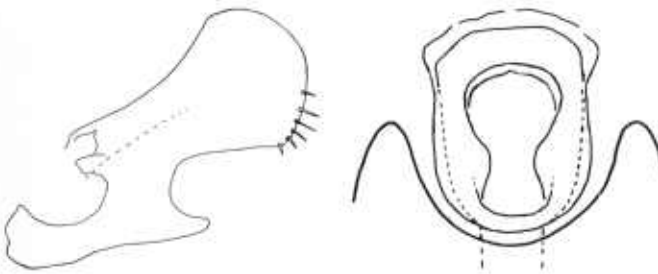
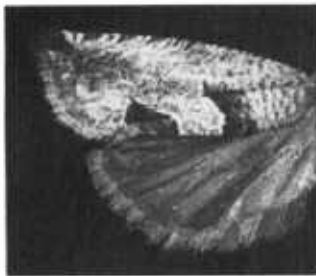
*Eucosma fulminana*  
(Walsingham)

Forewing 11.0 mm long, brown with silver specks. No capture date. MI. Syst: Heinrich (1923b). (1 N, 1 Gm, T)

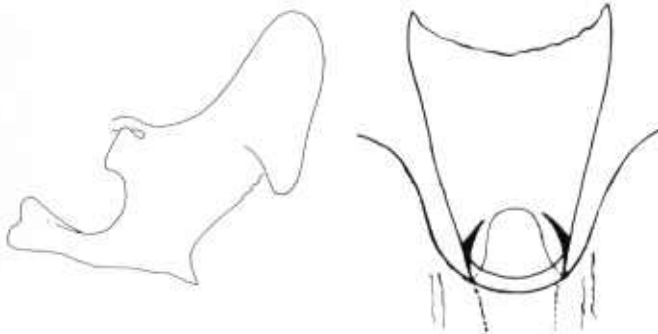




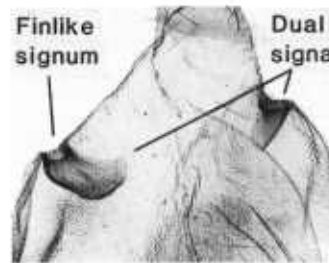
*Eucosma sombreana* Kearfott  
Forewing 7.0 to 12.5 mm long, grayish brown or yellowish brown. Adults captured June 26–August 27. MI, WI, MN. Univoltine. Larva feeds in *Helianthus* rootstalks. Syst and Biol: Heinrich (1923b). (43 N, 10 Gm, 3 Gf, T)



*Eucosma consobrinana* Heinrich  
Forewing 6.0 to 9.0 mm long, dark markings brownish black. Adults captured June 7–September 1. MI, WI, MN. Syst: Heinrich (1923b). (85 N, 9 Gm, 10 Gf, T)



*Eucosma cataclystiana* (Walker)  
Forewing 6.0 to 8.5 mm long, dark areas orange yellow or yellowish brown. Adults captured June 22–September 7. MI, WI, MN. Larva feeds in *Solidago* rootstalks. Syst: Heinrich (1923b). Biol: Putman (1942). (36 N, 8 Gm, 3 Gf, T)



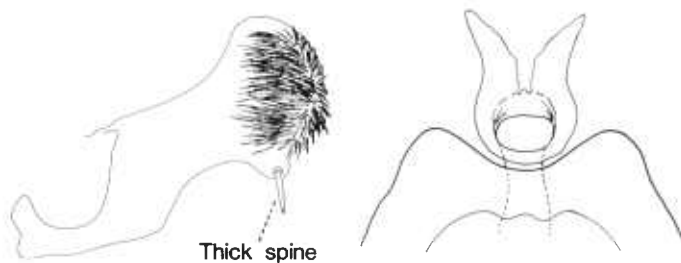
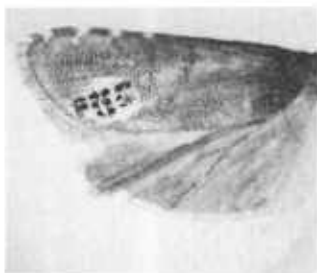
## Genus *Pelochrista*

**Both sexes.** Forewing without raised scale tufts, apex not falcate, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate,  $M_2$  and  $M_3$  separate,  $Cu_2$  not originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing with costal fold. Hindwing without upper-surface melanic sex scaling. Socius finger- or ribbonlike, not heavily sclerotized; valva not divided, sacculus not densely clothed with spinelike setae, without rudimentary clasper, outer surface lacking spinelike setae, with thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 emarginate posteriorly or inflected and overlapping ostium bursae. Lamella antevaginalis not conical. Ductus bursae sclerotized less than two-thirds its length from genital opening; corpus bursae with two thorn- or finlike signa.

**Comments.** More than 20 Nearctic species of *Pelochrista* are known.



*Pelochrista scintillana* (Clemens)

Forewing 7.5 to 11.5 mm long, medium pale areas orange yellow. Ovipositor with ventral extensions of papillae anales. Adults captured May 29–August 4. MI, WI, MN. Larva feeds on *Helianthus*. Syst: Heinrich (1923b). Biol: Walker (1936). (23 N, 8 Gm, 3 Gf, T)



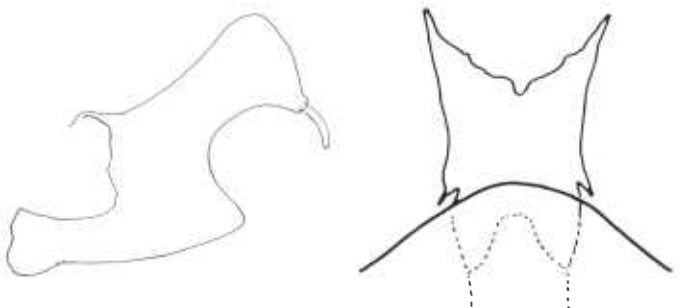
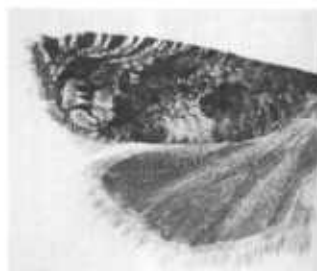
*Pelochrista corosana* (Walsingham)

Forewing 6.5 mm long, dark areas and markings grayish brown or brownish black. Adult captured July 23. MN. Syst: Heinrich (1923b). (1 N, 1 Gm, T, photo specimen Fillmore Co., NE)



*Pelochrista zomonana* (Kearfott)

Forewing 5.0 to 6.5 mm long, dark areas and markings grayish brown or brownish black. Sides of female corpus bursae sclerotized near ductus bursae. Adults captured June 10–September 8. MI. Larva feeds in *Chrysanthemum* roots and stems. Syst: Heinrich (1923b). Biol: MacKay (1959). (20 N, 11 Gm, 2 Gf, T)



*Pelochrista womonana* (Kearfott)

Forewing 6.5 to 9.0 mm long, dark areas and markings grayish yellowish brown or brownish black. Sides of female corpus bursae sclerotized near ductus bursae. Adults captured June 10–August 11. MI, MN. Larva feeds in *Helianthus* roots. Syst: Heinrich (1923b). Biol: Rogers *et al.* (1979). (11 N, 9 Gm, 1 Gf, T)

## Genus *Epiblema*

**Both sexes.** Forewing without raised scale tufts, termen straight or slightly concave, apex not falcate,  $R_1$  originating well before middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate,  $M_2$  and  $M_3$  separate. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

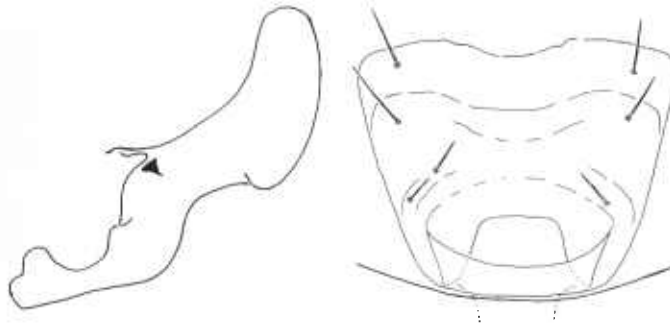
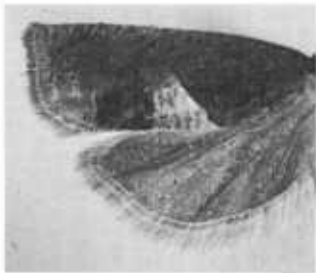
**Male.** Antenna not notched near base. Forewing with costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius finger- or ribbonlike, not heavily sclerotized; valva not divided, sacculus not densely clothed with spinelike setae, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin; vesica with all cornuti deciduous.

**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Lamella antevaginalis not developed. Ductus bursae sclerotized less than two-thirds its length from genital opening; corpus bursae without sclerotized sides, with two thorn- or finlike signa.

**Comments.** Among the species treated here, males of all except *Epiblema tandana* have a rudimentary valval clasper.

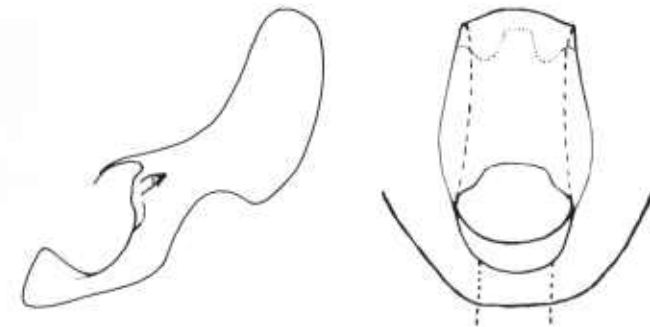
Nearly 40 Nearctic species of *Epiblema* are known. Brown (1973) gives a generic treatment.

Because specific distinctness between *E. strenuana* and *E. minutana* (Kearfott) is doubtful (Miller and Pogue 1984), these two are considered the *E. strenuana* complex here.



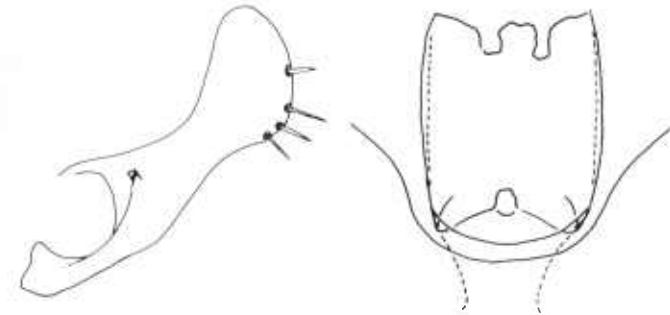
*Epiblema boxcana* (Kearfott)

Forewing 5.5 to 7.5 mm long, dark areas yellowish brown. Female ductus bursae sclerotized near middle. Adults captured May 28–July 19. MI, MN. Syst: Heinrich (1923b). (34 N, 6 Gm, 5 Gf, T)



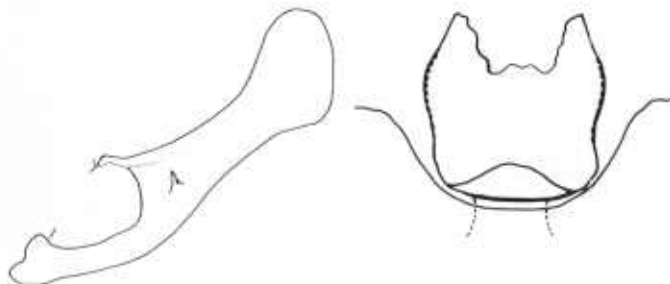
*Epiblema strenuana* (Walker) complex. Ragweed borer

Forewing 4.5 to 9.0 mm long, dark areas grayish brown. Female ductus bursae sclerotized near middle. Adults captured May 28–September 23. MI, WI, MN. Bivoltine. Larva bores in stems and branches of *Ambrosia*, *Xanthium*, *Chenopodium*. Syst and Biol: Miller and Pogue (1984). (93 N, 36 Gm, 27 Gf, T)



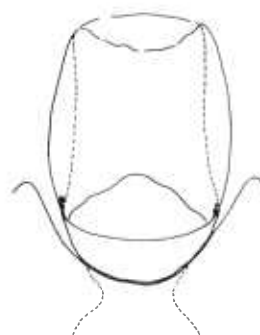
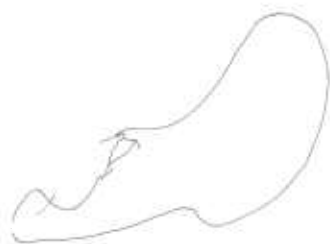
*Epiblema abruptana* (Walsingham)

Forewing 6.0 to 6.5 mm long, dark areas yellowish brown. Female ductus bursae not sclerotized near middle. Adults captured June 21–July 10. MI. Syst: Heinrich (1923b). (2 N, 2 Gm, T, photo specimen Clay Co., MO)



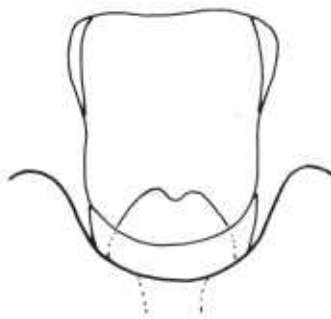
*Epiblema numerosana* (Zeller)

Forewing 6.0 to 8.5 mm long, dark areas grayish yellowish brown. Female ductus bursae not sclerotized near middle. Adults captured June 24–August 6. MN. Syst: Miller (1985a). (25 N, 4 Gm, 6 Gf, T)



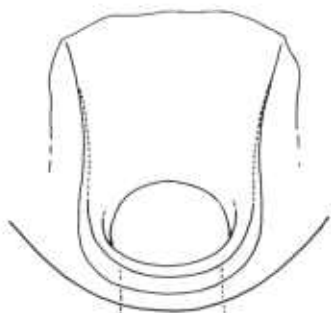
*Epiblema luctuosissima*  
Blanchard

Forewing 5.0 to 6.0 mm long, dark areas grayish brown. Female ductus bursae not sclerotized near middle. Adults captured June 17–August 16. MI, MN. Syst: Blanchard (1984). (10 N, 7 Gm, 3 Gf)



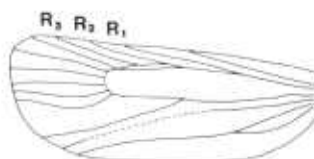
*Epiblema tripartitana* (Zeller)

Forewing 5.5 to 6.5 mm long, basal patch brownish black. Adults captured June 30–July 2. MI, WI. Larva feeds in flower heads and stems of *Rudbeckia*. Syst: Heinrich (1923b). Biol: Bottimer (1926). (6 N, 2 Gm, 1 Gf, T)



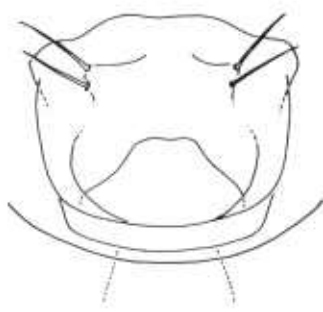
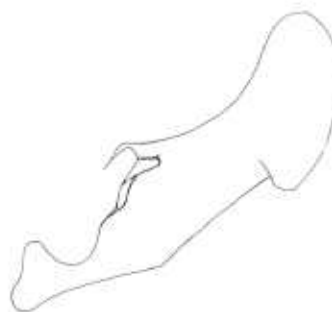
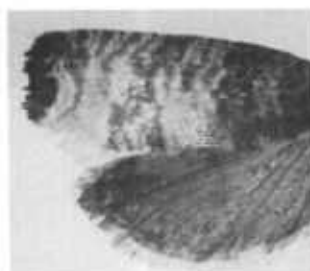
*Epiblema scudderiana*  
(Clemens)

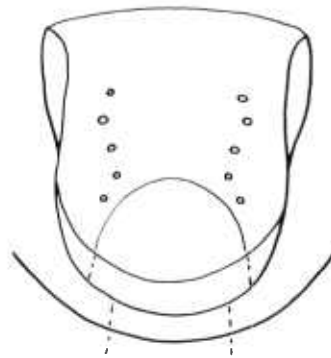
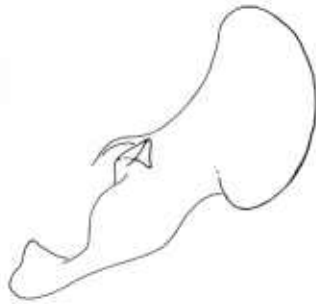
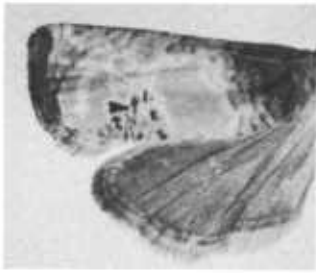
Forewing 7.0 to 10.5 mm long, dark areas and markings grayish brown or brownish black. Adults captured May 9–July 27. MI, WI, MN. Univoltine. Larva feeds on *Solidago*, mainly in stems. Syst and Biol: Miller (1976a). (145 N, 12 Gm, 5 Gf, T)



*Epiblema obfuscana* (Dyar)

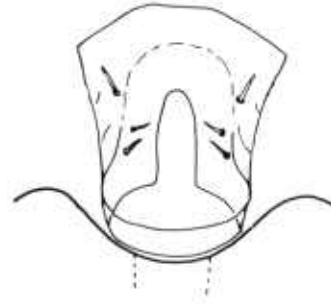
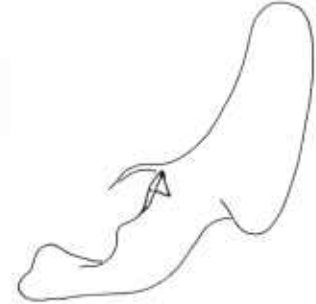
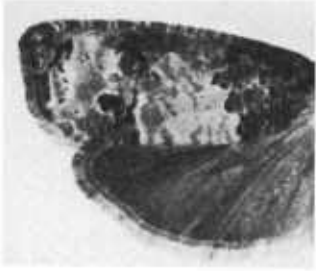
Forewing 5.5 to 9.5 mm long, dark areas and markings grayish brown or brownish black. Adults captured May 18–June 14. MI, WI. Larva feeds in *Solidago* stems. Syst: Heinrich (1923b). Biol: Putman (1942). (13 N, 5 Gm, 3 Gf, T)





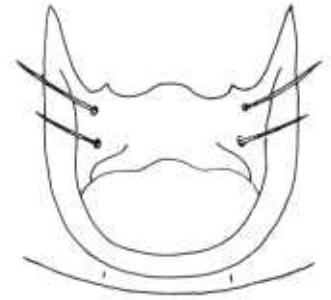
*Epiblema desertana* (Zeller)

Forewing 7.0 to 8.5 mm long, dark areas and markings grayish brown or brownish black. Adults captured May 24–June 25. MI. Univoltine. Larva feeds on *Solidago*, mainly in stems. Syst and Biol: Miller (1976a). (28 N, 6 Gm, 4 Gf, T)



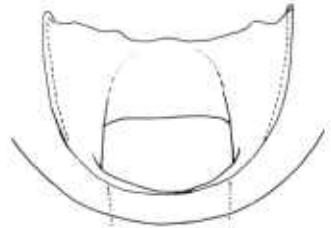
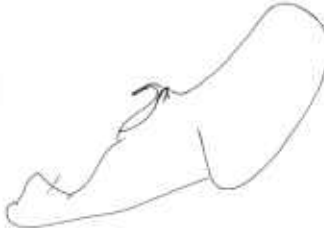
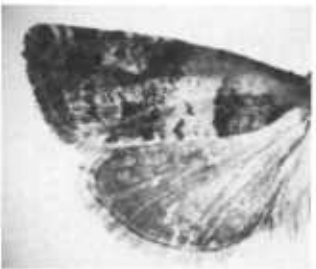
*Epiblema carolinana* (Walsingham)

Forewing 8.5 to 11.0 mm long, dark areas and markings grayish brown or brownish black. Adults captured July 4–August 6. MI, WI, MN. Univoltine. Larva feeds on *Rudbeckia*, mainly in roots. Syst: Heinrich (1923b). Biol: Thompson (1928). (11 N, 2 Gm, 2 Gf, T)



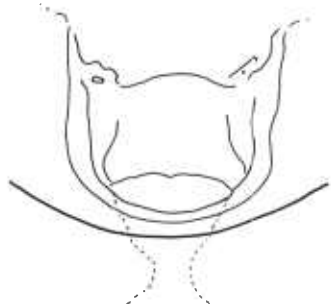
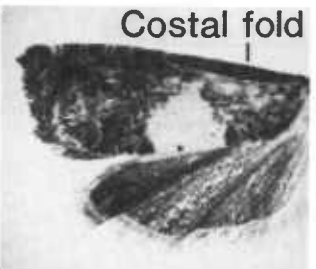
*Epiblema walsinghami* (Kearfott)

Forewing 8.0 to 9.0 mm long, dark areas and markings yellowish brown or brownish black. Adults captured June 30–July 18. MI. Syst: Heinrich (1923b). (10 N, 7 Gm, 1 Gf, T)



*Epiblema iowana* McDunnough

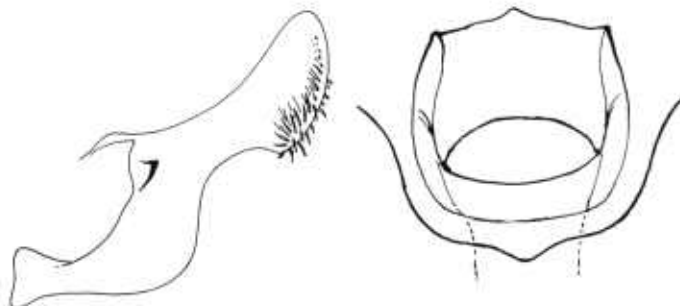
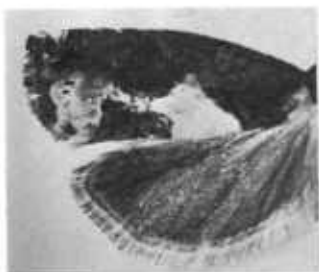
Forewing 6.5 mm long, dark markings brownish black. No capture date. MN. Larva feeds in *Ratibida* roots. Syst: Miller (1985a). Biol: Godfrey *et al.* (1987). (1 N, 1 Gm, photo specimen Boone Co., MO)



*Epiblema infelix* Heinrich

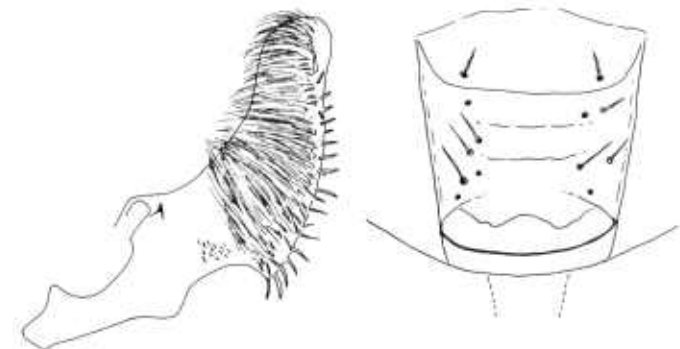
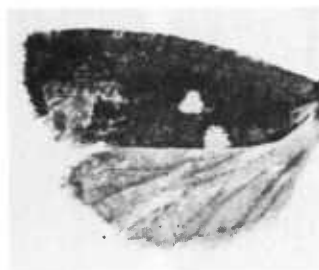
Forewing 5.5 to 7.5 mm long, dark areas and markings grayish brown or brownish black. Adults captured June 25–30. MI. Syst: Heinrich (1923b). (10 N, 9 Gm, T)





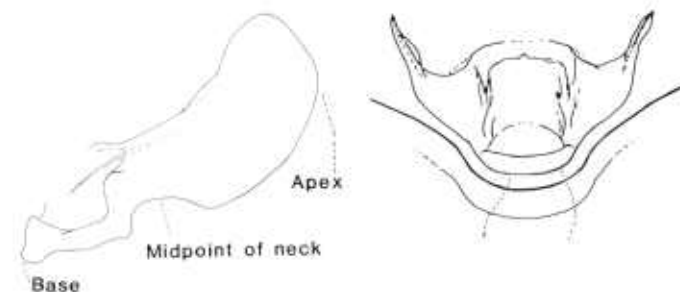
*Epiblema otiosana*  
(Clemens). *Bidens* borer

Forewing 6.0 to 9.0 mm long, dark areas grayish brown or brownish black. Variants shown are common (top) and rare (bottom). Adults captured May 30–August 30. MI, WI, MN. Larva feeds on *Bidens*, mainly in stems. Syst: Heinrich (1923b). Biol: Decker (1932). (80 N, 5 Gm, 7 Gf, T)



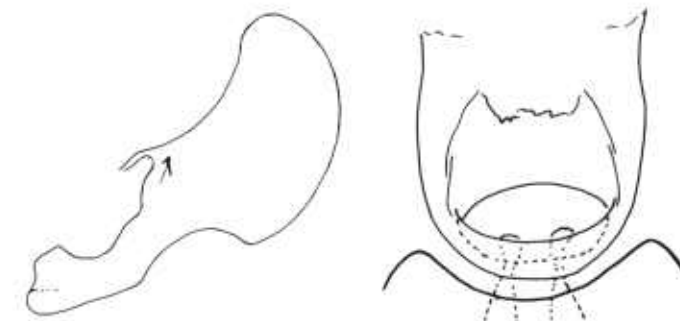
*Epiblema brightonana* (Kearfott)

Forewing 5.5 to 7.0 mm long, dark markings brownish black. Adults captured June 21–August 2. MI. Syst: Heinrich (1923b). (12 N, 4 Gm, 1 Gf, T)



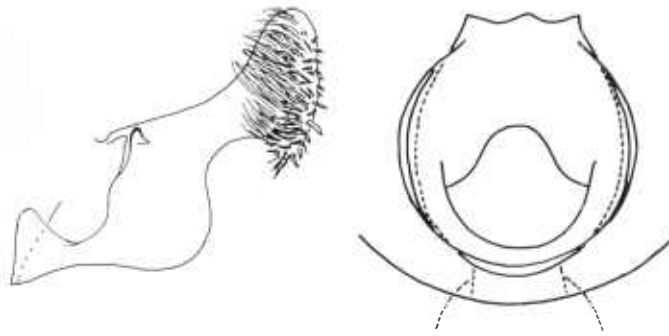
*Epiblema tandana* (Kearfott)

Forewing 8.0 to 10.0 mm long, dark markings brownish black. Adults captured June 5–July 21. MI, MN. Larva feeds in roots of *Rudbeckia*. Syst: Heinrich (1923b). Biol: Godfrey et al. (1987). (9 N, 5 Gm, T, photo specimen Bergen Co., NJ)



*Epiblema resumptana* (Walker)

Forewing 5.5 to 6.0 mm long, dark areas grayish brown or brownish black. Adults captured May 27–June 2. MI. Larva probably feeds on *Anaphalis margaritacea*. Syst and Biol: McDunnough (1959). (2 N, 2 Gm, T)



*Epiblema dorsisuffusana*  
(Kearfott)

Forewing 8.0 to 9.0 mm long,  
dark areas brownish black.  
Adults captured June 20–  
July 25. MI. Syst: Heinrich  
(1923b).  
(3 N, 2 Gm, 1 Gf, T)

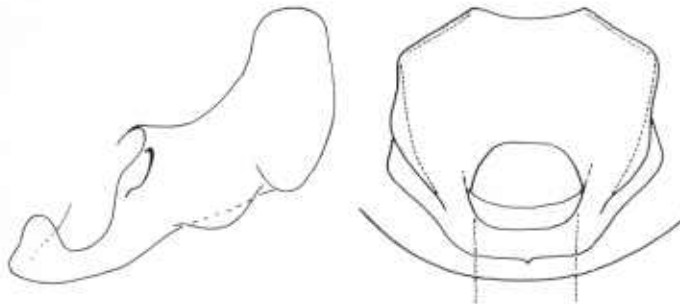
## Genus *Notocelia*

**Both sexes.** Forewing without raised scale tufts, termen straight, apex not falcate,  $R_1$  originating well before middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate,  $M_2$  and  $M_3$  separate. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing with costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius finger- or ribbonlike, not heavily sclerotized; valva not divided, sacculus not densely clothed with spinelike setae, with rudimentary clasper, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin; vesica with deciduous and nondeciduous cornuti.

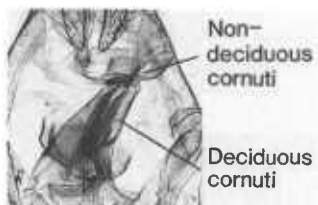
**Female.** Sternum 7 not deeply emarginate around ostium bursae, neither inflected nor overlapping ostium bursae. Lamella antevaginalis not conical, not wider than lamella postvaginalis in anteroposterior orientation. Ductus bursae sclerotized less than two-thirds its length from genital opening; corpus bursae without sclerotized sides, with two thorn- or finlike signa.

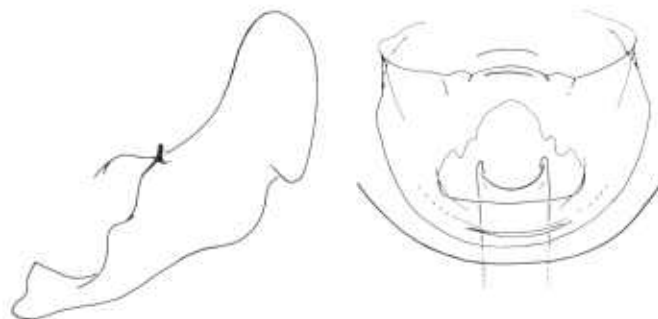
**Comments.** Four Nearctic species of *Notocelia* are known.



*Notocelia illotana* (Walsingham)

Forewing 6.5 to 8.5 mm long,  
dark areas and markings  
grayish brown or brownish  
black. Adults captured May 25–  
June 22. MI. Syst: Brown  
(1979a).  
(12 N, 3 Gm, 3 Gf, T)





*Notocelia culminana*  
(Walsingham)

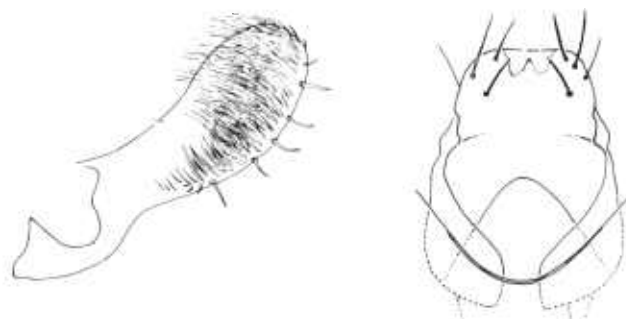
Forewing 7.0 to 8.5 mm long, dark areas and markings grayish brown or brownish black. Adults captured July 27--August 26. MI. Larva feeds in tied *Rosa* leaves. Syst: Brown (1979a). Biol: MacKay (1959). (5 N, 2 Gm, 2 Gf, T)

**Both sexes.** Forewing without raised scale tufts, termen concave, apex not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  united,  $M_2$  and  $M_3$  separate at base,  $M_2$ ,  $M_3$ , and  $Cu_1$  approximate at termen,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  united.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius present; valva not divided, without rudimentary clasper, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin.

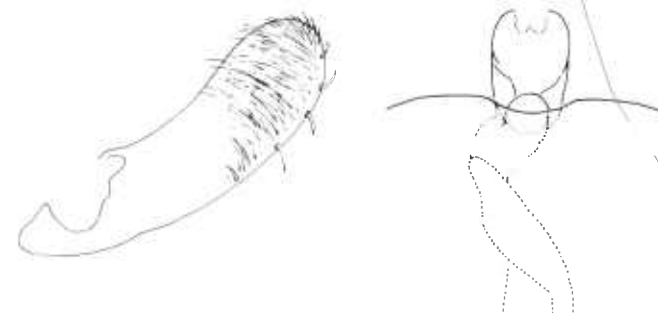
**Female.** Sternum 7 not deeply emarginate around ostium bursae, neither inflected nor overlapping ostium bursae. Ductus bursae sclerotized near middle; corpus bursae without sclerotized sides, with two finlike signa.

**Comments.** Six Nearctic species of *Suleima* are known.



*Suleima helianthana*  
(Riley). Sunflower bud moth

Forewing 6.5 to 7.0 mm long, dark areas and markings grayish yellowish brown or brownish black. Adults captured July 18--30. MI. Bivoltine. Larva feeds in buds, stems, and receptacles of *Helianthus*, *Coreopsis* Syst: Heinrich (1923b). Biol: Satterthwait (1948). (2 N, 2 Gm, T)



*Suleima cinerodorsana* Heinrich

Forewing 6.5 mm long, dark areas brownish black. Adult captured July 24. MI. Larva feeds in *Helianthus* stems. Syst: Heinrich (1923b). Biol: Putman (1942). (1 N, 1 Gm, T, photo specimen Allegheny Co., PA)

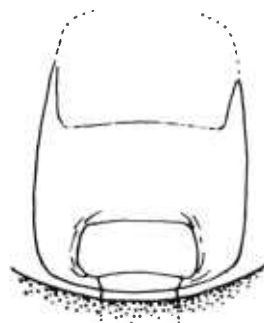
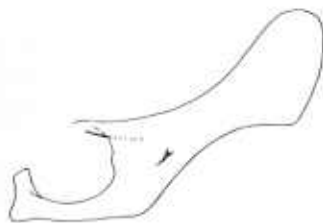
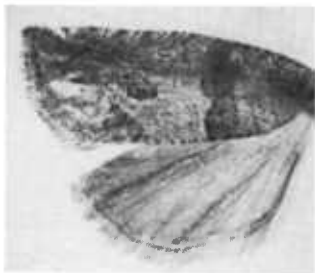
## Genus *Sonia*

**Both sexes.** Forewing without raised scale tufts, termen concave, apex not falcate,  $R_1$  originating well before middle of discal cell, upper internal vein of discal cell, originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  united,  $M_2$  and  $M_3$  separate at base,  $M_2$ ,  $M_3$ , and  $Cu_1$  approximate at termen,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing with costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius present; valva not divided, with rudimentary clasper, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 not deeply emarginate around ostium bursae, neither inflected nor overlapping ostium bursae. Ductus bursae with a sclerotized patch near middle; corpus bursae without sclerotized sides, with two finlike signa.

**Comments.** Six Nearctic species of *Sonia* are known.



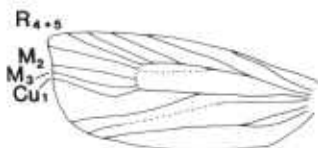
*Sonia paraplesiana* Blanchard

Forewing 6.0 to 7.0 mm long, dark areas and markings brown. Adults captured July 7–August 10. MI, MN. Syst: Blanchard (1979). (6 N, 5 Gm, 1 Gf)



*Sonia canadana* McDunnough

Forewing 6.0 to 8.5 mm long, dark areas brown. Adults captured July 8–August 23. MI. Larva feeds in rootstalks of *Aster*, *Solidago*. Syst: McDunnough (1925b). Biol: Capek (1971). (20 N, 10 Gm, 4 Gf, T)



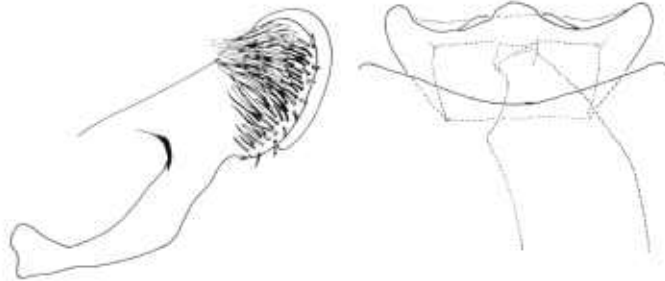
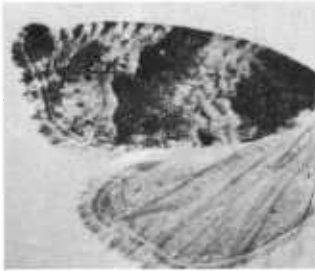
## Genus *Gypsonoma*

**Both sexes.** Forewing without raised scale tufts, termen slightly concave, apex not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_2$  and  $R_3$ ,  $R_4$  and  $R_5$  separate or approximate,  $M_2$  and  $M_3$  not connate,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  stalked,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius present; valva not divided, with rudimentary clasper, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin.

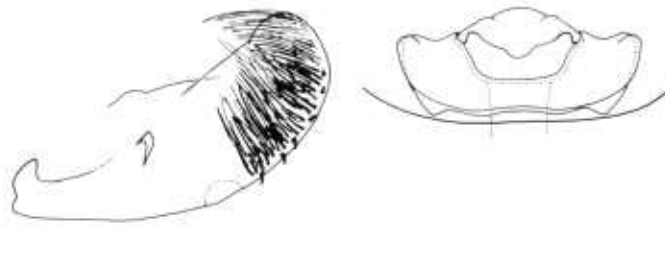
**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Lamella antevaginalis wider than lamella postvaginalis in anteroposterior orientation. Ductus bursae sclerotized less than three-quarters its length from genital opening; corpus bursae without sclerotized sides, with two thornlike signa.

**Comments.** Seven Nearctic species of *Gypsonoma* are known.



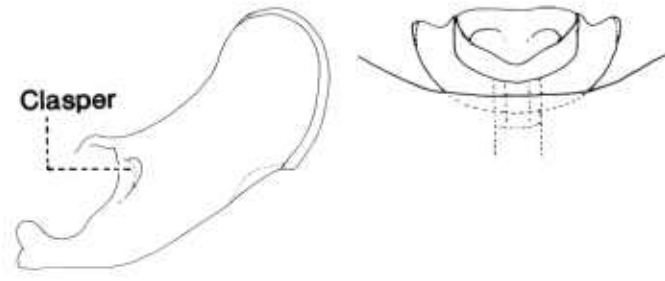
*Gypsonoma fasciolana*  
(Clemens)

Forewing 6.0 to 8.0 mm long, dark areas and markings grayish brown or brownish black. Adults captured May 29–July 23. MI, WI, MN. Larva feeds in rolled leaves of *Salix*, *Populus*. Syst: Heinrich (1923b). Biol: Prentice (1966). (107 N, 19 Gm, 5 Gf, T)



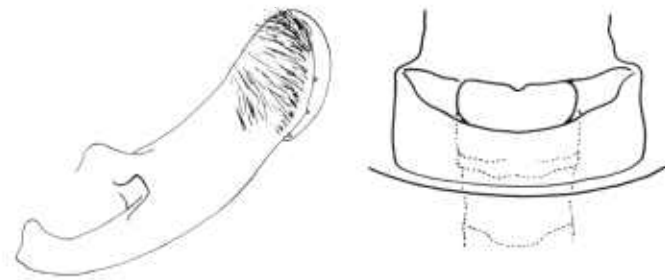
*Gypsonoma haimbachiana*  
(Kearfott). Cottonwood twig borer

Forewing 5.5 to 7.5 mm long, dark areas and markings grayish brown or brownish black. Female ductus bursae not sclerotized. Adults captured June 26–August 12. MI, WI, MN. Larva feeds on *Populus deltoides*, primarily in shoots. Syst: Heinrich (1923b). Biol: Morris (1967). (16 N, 3 Gm, 3 Gf, T)



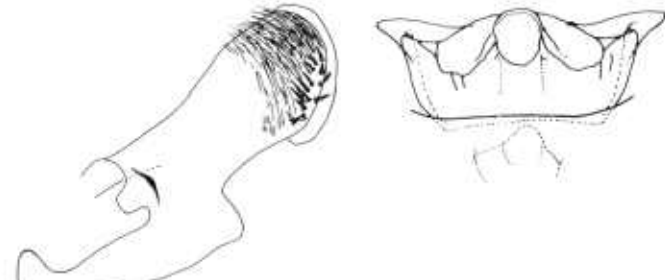
*Gypsonoma substitutionis*  
Heinrich

Forewing 4.5 to 6.0 mm long, dark areas and markings grayish yellowish brown or brownish black. Adults captured June 7–August 12. MI, WI. Larva feeds in rolled leaves of *Populus*, *Quercus*, *Acer*. Syst: Heinrich (1923b). Biol: Prentice (1966). (17 N, 9 Gm, 1 Gf, T)



*Gypsonoma salicicolana*  
(Clemens)

Forewing 5.5 mm long, dark areas and markings grayish brown. Adult captured June 20. MI. Larva feeds in rolled *Salix* leaves. Syst: Heinrich (1923b). Biol: Prentice (1966). (1 N, 1 Gm, T)



*Gypsonoma adjuncta* Heinrich

Forewing 5.5 to 7.0 mm long, dark areas and markings black. Adults captured June 24–July 17. MI, WI, MN. Larva feeds in rolled *Populus* leaves. Syst: Heinrich (1924). Biol: Prentice (1966). (9 N, 5 Gm, 3 Gf, T)

## Genus *Proteoteras*

**Both sexes.** Forewing with raised scale tufts near lower margin of discal cell, termen concave or straight, forewing not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate,  $M_2$  and  $M_3$  separate at base,  $M_2$ ,  $M_3$ , and  $Cu_1$  approximate at termen,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing with upper-surface melanic sex scaling or hair pencil near costa. Socius present; valva not divided, without rudimentary clasper, outer surface with long, flat spinelike setae, without thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 deeply emarginate around ostium bursae, neither inflected nor overlapping ostium bursae, with narrow, recessed lamella postvaginalis, ostium bursae elongate. Ductus bursae sclerotized near middle; corpus bursae without sclerotized sides, with two finlike signa.

**Comments.** Eight Nearctic species of *Proteoteras* are known.

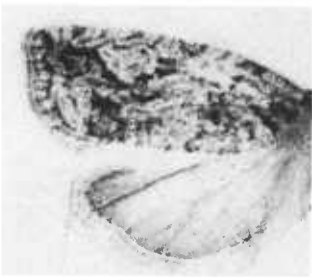


*Proteoteras aesculana* Riley

Forewing 6.0 to 9.0 mm long, dark areas and markings grayish olive green or olive black. Adults captured April 12–October 2. MI, WI, MN. Larva feeds in leafstalks, seeds, and shoots of *Acer negundo*, *Aesculus*. Syst: Wong *et al.* (1983). Biol: Powell (1962). (63 N, 13 Gm, 6 Gf, T)

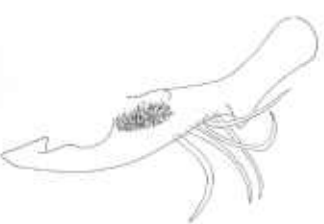


Melanic sex scaling



*Proteoteras willingana* (Kearfott). Boxelder twig borer

Forewing 7.0 to 8.5 mm long, dark areas and markings grayish yellowish brown or brownish black. Adults captured June 19–July 23. MI, MN. Univoltine. Larva feeds on leaves, and in buds and shoots of *Acer negundo*. Syst: Wong *et al.* (1983). Biol: Peterson (1958). (27 N, 6 Gm, 7 Gf, T)



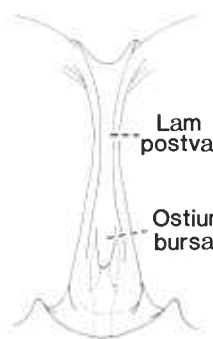
*Proteoteras crescentana* Kearfott

Forewing 7.0 to 9.0 mm long, pale area within dark crescent yellowish brown or grayish yellowish brown. Adults captured June 8–July 15. MI, WI, MN. Larva feeds in *Acer negundo* shoots. Syst: Wong *et al.* (1983). Biol: Prentice (1966). (10 N, 4 Gm, 2 Gf, T)



*Proteoteras naracana* Kearfott

Forewing 7.0 to 8.0 mm long, dark areas and markings brown or brownish black. Adults captured June 6–26. MI. Larva feeds on *Acer*. Syst: Heinrich (1923b). Biol: Godfrey *et al.* (1987). (8 N, 3 Gm, 2 Gf, T)



*Proteoteras moffatiana* Fernald

Forewing 7.0 to 9.5 mm long, pale areas green. Adults captured July 5–August 30. MI, WI, MN. Univoltine. Larva feeds in buds and shoots of *Acer saccharum*, *A. rubrum*, *A. saccharinum*. Syst: Heinrich (1923b). Biol: Prentice (1966). (58 N, 18 Gm, 12 Gf, T)



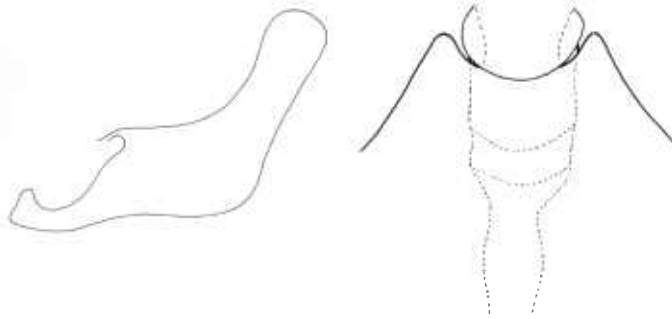
## Genus *Zeiraphera*

**Both sexes.** Forewing without raised scale tufts, termen straight or slightly concave, apex not falcate,  $R_1$  originating well before middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  not united,  $M_2$  and  $M_3$  separate. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Socius not finger- or ribbonlike; valva not divided, sacculus not densely clothed with spinelike setae, without rudimentary clasper, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at anal angle.

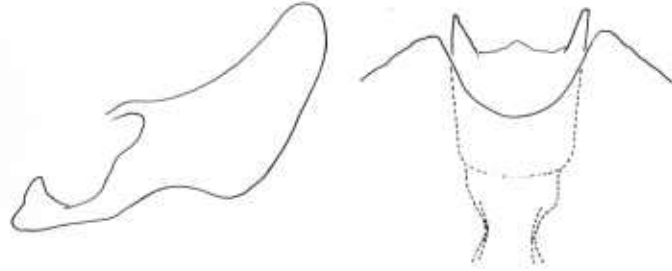
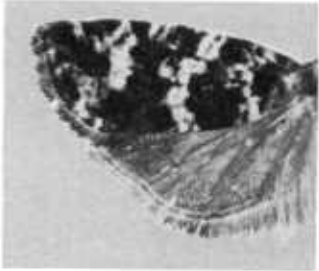
**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Ductus bursae with a sclerotized patch near middle; corpus bursae without sclerotized sides, with two thorn- or finlike signa.

**Comments.** Eight Nearctic species of *Zeiraphera* are known. Mutuura and Freeman (1966) reviewed the species of *Zeiraphera*.



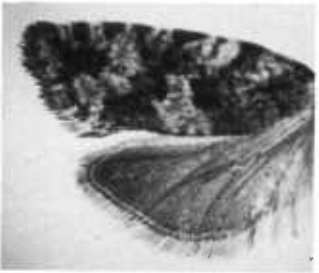
*Zeiraphera canadensis*  
Mutuura and Freeman.  
Spruce bud moth

Forewing 6.0 to 6.5 mm long, pale areas orange yellow. Adults captured July 18–24. MI, WI. Univoltine. Larva feeds on buds and foliage of *Picea glauca*, *P. mariana*, *Abies balsamea*. Syst: Mutuura and Freeman (1966). Biol: Pilon (1965). (7 N, 2 Gm, 3 Gf, T)



*Zeiraphera fortunana* (Kearfott)

Forewing 5.5 mm long, dark markings grayish brown or brownish black. Adult captured July 6. WI. Univoltine. Larva feeds progressively on buds and needles of *Picea*, *Abies*. Syst: Mutuura and Freeman (1966). Biol: Prentice (1966). (1 N, 1 Gm, T, photo specimen Carleton Co., ON)



*Zeiraphera unfortunana* Powell

Forewing 7.0 mm long, dark markings black or grayish yellowish brown. Adults captured July 22–August 8. MI, MN. Larva feeds on *Picea*, *Abies*. Syst: Powell (1983). Biol: Mutuura and Freeman (1966). (2 N, 2 Gm, photo specimen Thunder Bay Co., ON)

## Genus *Pseudexentera*

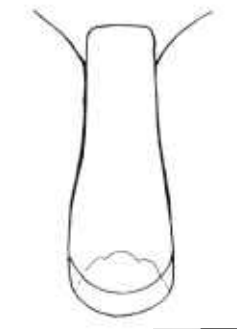
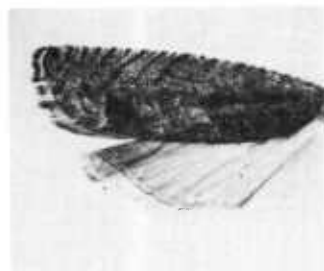
**Both sexes.** Forewing without raised scale tufts, termen concave, apex not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  not united,  $M_2$  and  $M_3$  not connate,  $M_2$ ,  $M_3$ , and  $Cu_1$  approximate at termen,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius not finger- or ribbonlike; valva not divided, saccus densely clothed with spinelike setae, without rudimentary clasper, outer service lacking spinelike setae, without thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 deeply emarginate around ostium bursae, neither inflected nor overlapping ostium bursae. Lamella antevaginalis not conical, lamella postvaginalis not recessed, ostium bursae not elongate. Ductus bursae sclerotized near middle; corpus bursae without sclerotized sides, with two finlike signa.

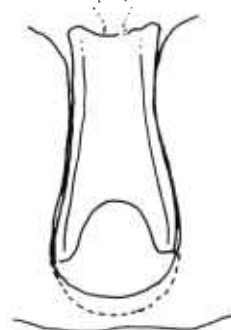
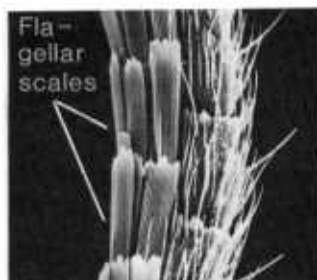
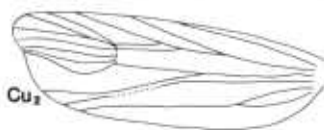
**Comments.** Nearly 20 Nearctic species of *Pseudexentera* are known. Miller (1986b) reviewed the species of *Pseudexentera*.





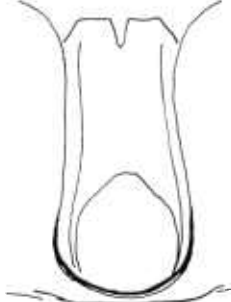
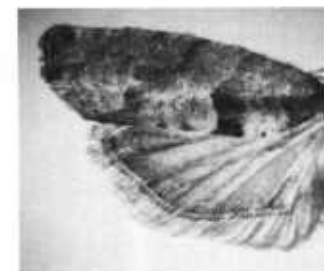
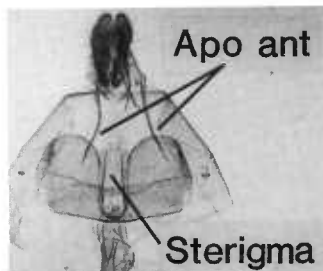
*Pseudexentera cressoniana*  
(Clemens)

Forewing 8.0 to 10.5 mm long, pale areas brownish gray. Adults captured March 31–May 31. MI, WI. Larva feeds on *Carya*. Syst and Biol: Miller (1986b). (55 N, 13 Gm, 13 Gf, T)



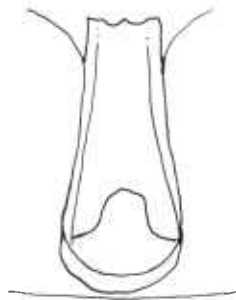
*Pseudexentera faracana*  
(Kearfott)

Forewing 7.0 to 9.5 mm long, variable pattern; dark markings medium brown or dark brown. Most divergent variants shown. Adults captured April 6–May 7. MI, WI. Univoltine. Larva feeds in rolled *Castanea* leaves. Syst and Biol: Miller (1986b). (4 N, 1 Gm, 3 Gf, T, top photo specimen Putnam Co., IL, bottom, Allegheny Co., PA)



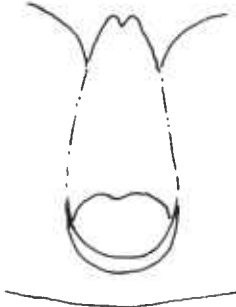
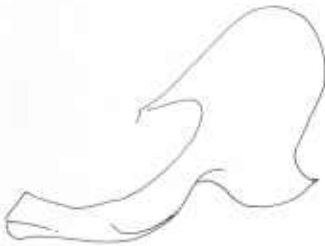
*Pseudexentera sepia* Miller

Forewing 7.0 mm long, dark markings dark brown. Adults captured April 25–May 20. WI. Syst: Miller (1986b). (2 N, 1 Gm, 1 Gf, T)



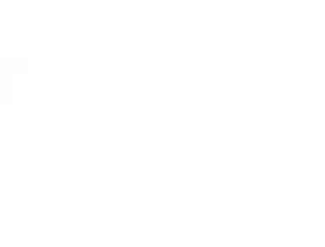
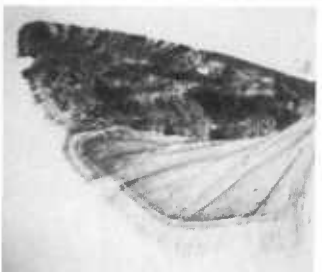
*Pseudexentera haracana*  
(Kearfott)

Forewing 8.0 mm long, dark markings brownish black. Adults captured May 2–30. MI, WI, MN. Syst: Miller (1986b). (3 N, 3 Gm, T, photo specimen Boone Co., MO)



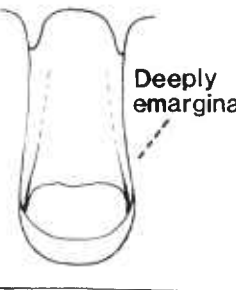
*Pseudexentera spoliata*  
(Clemens)

Forewing 7.0 to 9.0 mm long, variable pattern; dark markings brownish black. Limits of variation shown. Adults captured March 1–May 30. MI, WI. Univoltine. Larva feeds in rolled *Quercus* leaves. Syst and Biol: Miller (1986b). (59 N, 9 Gm, 16 Gf, T)



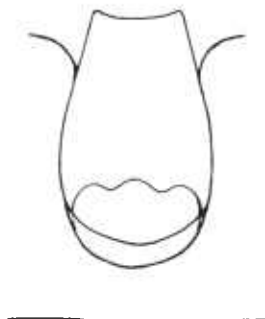
*Pseudexentera mali* Freeman

Forewing 7.0 to 8.0 mm long, variable pattern; pale areas brown or brownish gray. Limits of variation shown. Adults captured April 6–May 5. MI, WI. Univoltine. Larva feeds in buds, folded leaves, and young fruits of *Pyrus*, *Crataegus*. Syst: Freeman (1942). Biol: Chapman and Lienk (1971). (13 N, 5 Gm, 6 Gf, T)



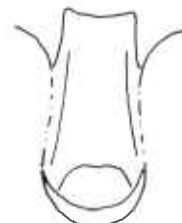
Deeply  
emarginate





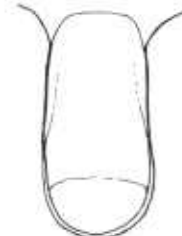
*Pseudexentera oregonana*  
(Walsingham)

Forewing 7.5 to 9.5 mm long, variable pattern; pale areas grayish brown. Limits of variation shown. Adults captured March 26–May 30. MI, WI. Univoltine. Larva feeds in rolled leaves of *Populus tremuloides*, *Salix*. Syst: Miller (1986b). Biol: Wong and Melvin (1967). (31 N, 5 Gm, 2 Gf, T, bottom photo specimen Aweme, MB)



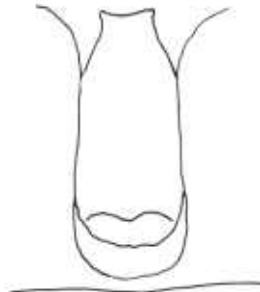
*Pseudexentera kalmiana*  
McDunnough

Forewing 5.5 to 6.0 mm long, dark areas dark grayish brown. Adults captured May 3–25. MI. Larva feeds on *Kalmia*. Syst and Biol: Miller (1986b). (7 N, 4 Gm, 3 Gf, T)



*Pseudexentera maracana*  
(Kearfott)

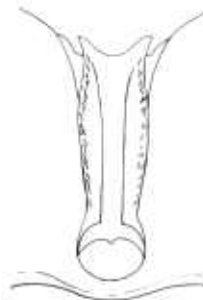
Forewing 7.0 to 7.5 mm long, dark markings dark grayish brown or brownish black. Adults captured April 2–May 15. MI, MN. Larva feeds on *Crataegus*. Syst and Biol: Miller (1986b). (8 N, 3 Gm, 5 Gf, T)



*Pseudexentera vaccinii* Miller

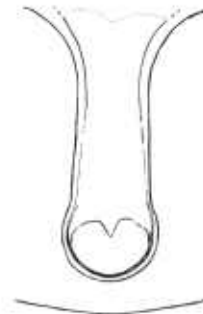
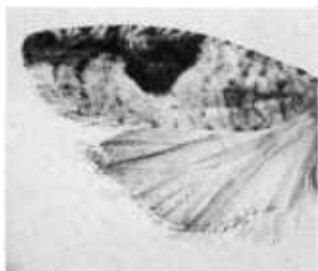
Forewing 6.5 to 8.0 mm long, dark markings grayish brown. Adults captured April 8–May 25. MI, MN. Larva feeds on *Vaccinium*. Syst and Biol: Miller (1986b). (19 N, 10 Gm, 8 Gf, T)





*Pseudexentera costomaculana*  
(Clemens)

Forewing 7.0 to 9.5 mm long, pale areas orange yellow. Adults captured April 23–July 18. MI. Larva feeds in terminals and folded leaves of *Hamamelis*. Syst and Biol: Miller (1986b). (28 N, 4 Gm, 4 Gf, T)



*Pseudexentera virginiana*  
(Clemens)

Forewing 7.5 to 9.0 mm long, pale areas yellowish brown. Adults captured March 25–May 11. MI. Syst: Miller (1986b). (25 N, 4 Gm, 3 Gf)

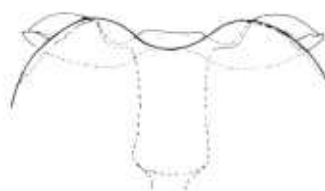
## Genus *Gretchena*

**Both sexes.** Forewing with raised scale tufts near dorsal margin, apex not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  not united,  $M_2$  and  $M_3$  not connate,  $M_2$ ,  $M_3$ , and  $Cu_1$  approximate at termen,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus not developed; socius not finger- or ribbonlike; valva not divided, sacculus densely clothed with spinelike setae, without rudimentary clasper, outer surface lacking spinelike setae.

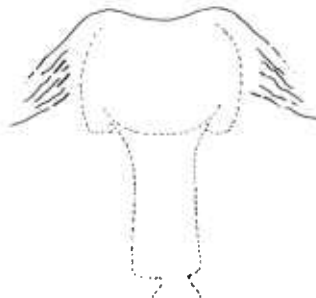
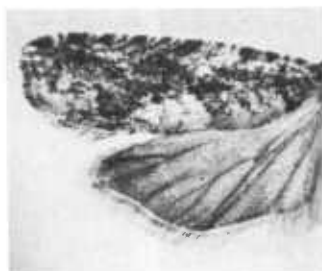
**Female.** Sternum 7 inflected and overlapping ostium bursae, without median bilobed projection. Ductus bursae sclerotized near middle; corpus bursae without sclerotized sides, with two thornlike signa.

**Comments.** Eleven Nearctic species of *Gretchena* are known. Brown (1982) gives a generic diagnosis.



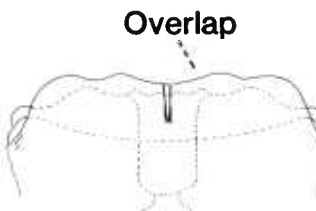
*Gretchena deludana* (Clemens)

Forewing 7.0 to 8.0 mm long, dark markings brownish black. Adults captured May 6–31. MI. Syst: Heinrich (1923b). (21 N, 5 Gm, 3 Gf)



*Gretchena concubitana* Heinrich

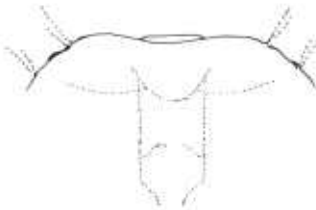
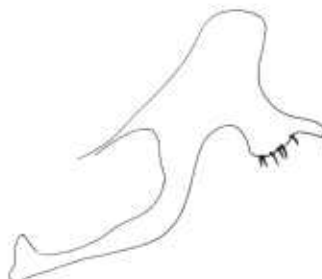
Forewing 7.0 to 9.0 mm long, dark markings brownish black. Adults captured May 4–June 3. MI. Larva feeds on *Carya*. Syst and Biol: Heinrich (1923b). (17 N, 5 Gm, 7 Gf, T)



Overlap

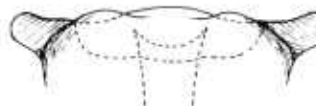
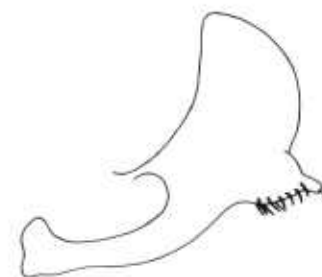
*Gretchena watchungana* (Kearfott)

Forewing 5.5 to 8.0 mm long, dark markings brownish black. Adults captured April 26–June 7. MI, WI, MN. Larva feeds on *Alnus*. Syst: Heinrich (1923b). Biol: Krauth et al. (1977). (37 N, 9 Gm, 7 Gf, T)



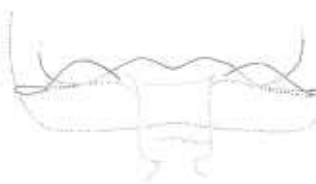
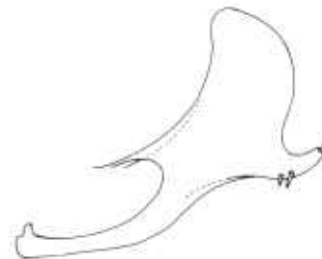
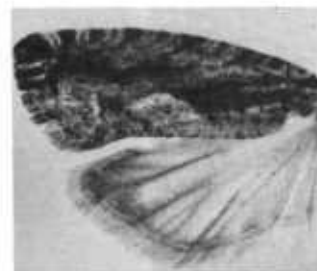
*Gretchena bolliana* (Slingerland). Pecan bud moth

Forewing 7.5 to 8.5 mm long, dark areas and markings black or brownish black. Adults captured June 25–October 4. MI, MN. Larva feeds on buds, foliage, and fruits of *Carya illinoensis*, *Juglans*. Syst: Heinrich (1923b). Biol: Moznette et al. (1931). (6 N, 4 Gm, 1 Gf, T)



*Gretchena amatana* Heinrich

Forewing 7.5 to 8.5 mm long, dark areas and markings brownish black. Adults captured May 11–August 3. MI, WI. Syst: Heinrich (1923b). (14 N, 6 Gm, 4 Gf, T)



*Gretchena delicatana* Heinrich

Forewing 7.0 to 8.0 mm long, dark areas and markings brownish black. Adults captured May 6–June 19. MI, WI, MN. Syst: Heinrich (1923b). (15 N, 6 Gm, 5 Gf, T)

## Genus *Rhopobota*

**Both sexes.** Forewing without raised scale tufts, termen concave, apex not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_2$  and  $R_3$ ,  $R_4$  and  $R_5$  not connate,  $M_2$  and  $M_3$  not connate,  $M_2$ ,  $M_3$ , and  $Cu_1$  approximate at termen. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus bifurcate; socius present; valva not divided, without thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Corpus bursae with sides sclerotized, with two thornlike signa.

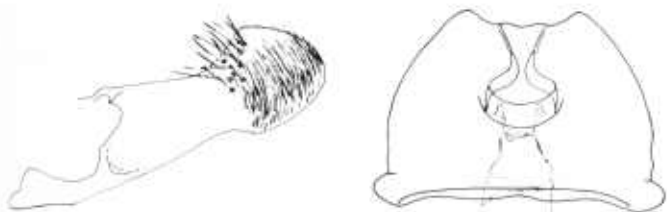
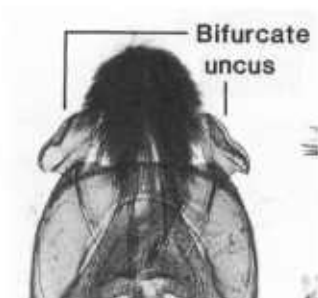
**Comments.** Three Nearctic species of *Rhopobota* are known. Among the species treated here, *R. naevana* also occurs in the Palearctic.

Brown (1983) gives a synopsis of the genus.



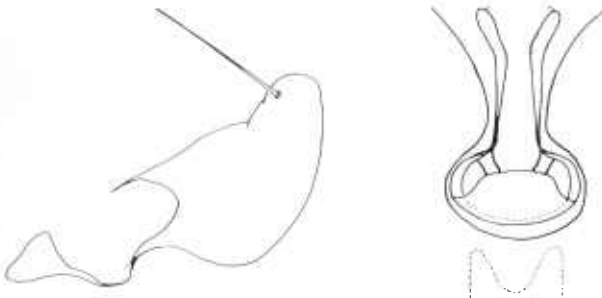
*Rhopobota naevana* (Hübner). Blackheaded fireworm

Forewing 5.0 to 7.0 mm long, dark markings brownish black or grayish brown. Adults captured May 27–August 24. MI, WI. Multivoltine. Larva feeds on *Vaccinium* buds, leaves, flowers, and fruit. Syst: Brown (1983). Biol: Plank (1922). (17 N, 5 Gm, 11 Gf)



*Rhopobota dietziana* (Kearfott)

Forewing 5.5 to 6.0 mm long, dark markings brownish black. Adults captured May 28–August 12. MI. Larva feeds on *Ilex* leaves. Syst: Brown (1983). Biol: Ferguson (1975). (4 N, 2 Gm, 1 Gf, T)



*Rhopobota finitimana* (Heinrich)

Forewing 4.5 to 5.0 mm long, dark areas and markings brownish black. Adults captured June 12–21. MI. Larva feeds on leaves of *Nemopanthus mucronata*, *Ilex*. Syst: Brown (1983). Biol: Ferguson (1975). (7 N, 2 Gm, 5 Gf, T)

## Genus *Epinotia*

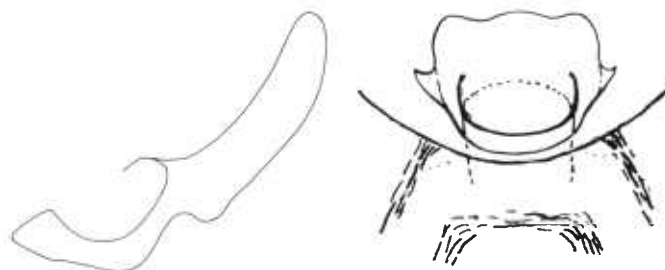
**Both sexes.** Forewing apex not falcate, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  not united,  $M_2$  and  $M_3$  not connate. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Hindwing without upper-surface melanic sex scaling. Uncus developed; socius not finger- or ribbonlike; valva not divided, sacculus densely spined, without rudimentary clasper, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin; anellus tightly surrounding aedeagus.

**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Lamella antevaginalis not conical. Ductus bursae sclerotized near middle; corpus bursae without sclerotized sides, with two thorn- or finlike signa.

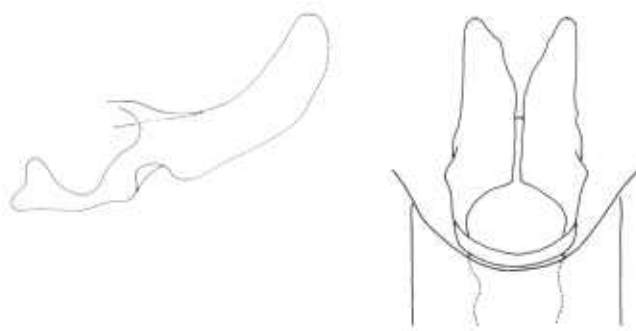
**Comments.** More than 75 Nearctic species of *Epinotia* are known. Among the species treated here, *E. solandriana*, *E. nisella*, *E. cruciana*, and *E. nanana* also occur in the Palearctic.

Brown (1980a, 1986) gives a generic treatment.



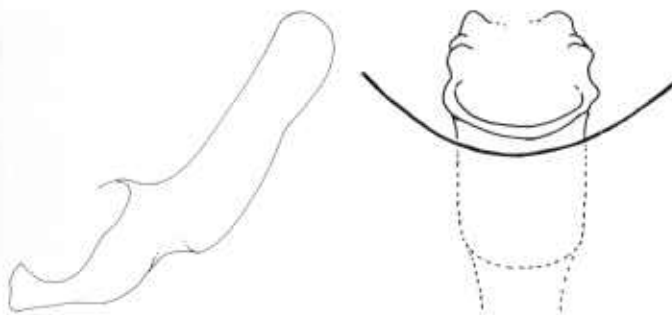
*Epinotia solandriana* (Linnaeus)

Forewing 8.0 to 10.5 mm long, variable pattern; dark areas and markings brown, grayish brown, or brownish black. Common variants shown. Adults captured July 5–September 5. MI, WI, MN. Univoltine. Larva feeds in rolled leaves of *Betula*, *Populus*, *Alnus*, others. Syst: Robinson and Nielsen (1983). Biol: Lindquist and Macleod (1967). (56 N, 3 Gm, 13 Gf)



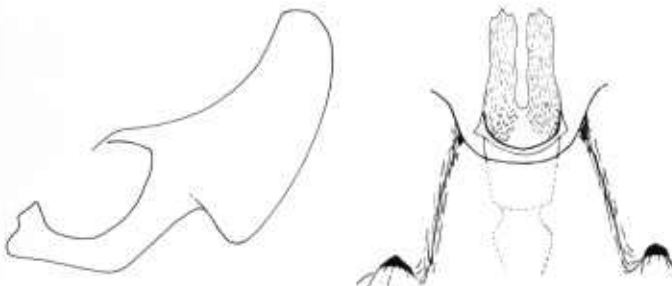
*Epinotia medioviridana* (Kearfott)

Forewing 7.0 to 8.0 mm long, pale areas green. Adults captured August 10–September 8. MI, WI. Larva feeds in tied *Rubus* leaves. Syst: Brown (1980a). Biol: MacKay (1953). (4 N, 1 Gm, 2 Gf, T)



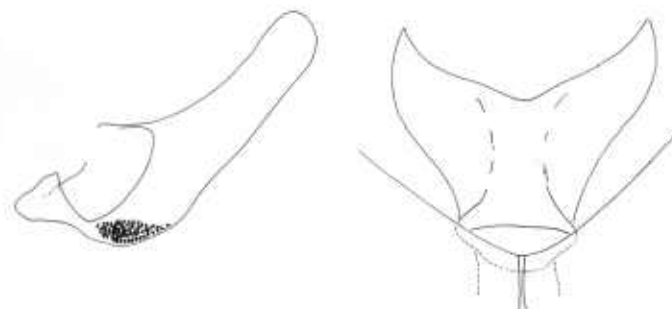
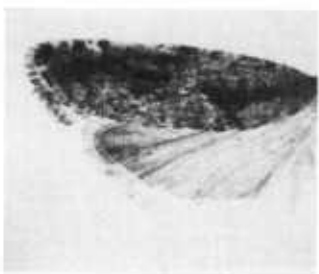
*Epinotia radicana* (Heinrich)

Forewing 7.0 mm long, dark markings brown. Adults captured June 24–25. MN. Univoltine. Larva feeds on foliage of *Picea glauca*, *P. mariana*, *Abies balsamea*, others. Syst: Brown (1983). Biol: Blais (1961). (2 N, 1 Gm, 1 Gf)



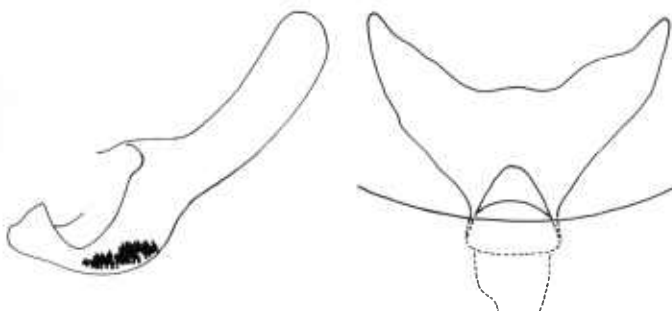
*Epinotia madderana* (Kearfott)

Forewing 6.0 to 6.5 mm long, terminal markings brown. Adults captured June 24–July 15. MI, MN. Syst: Brown (1980a). (7 N, 2 Gm, 2 Gf, T)



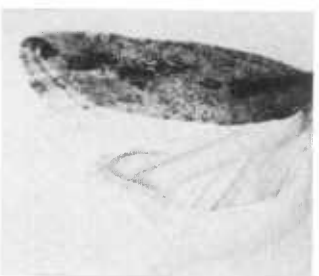
*Epinotia laracana* (Kearfott)

Forewing 6.5 to 7.5 mm long, dark areas and markings brownish black. Adults captured April 20–May 20. MI, WI. Larva feeds on *Celtis*. Syst and Biol: Brown (1986). (17 N, 2 Gm, 15 Gf)

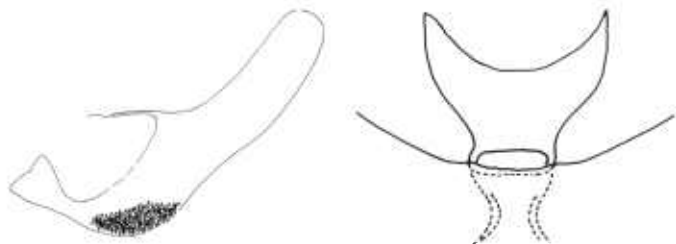


*Epinotia zandana* (Kearfott)

Forewing 6.0 to 8.0 mm long, variable pattern; dark markings brownish black. Common variants shown. Adults captured March 23–April 21. MI. Larva feeds on *Crataegus*. Syst and Biol: Brown (1986). (25 N, 9 Gm, 12 Gf, T)

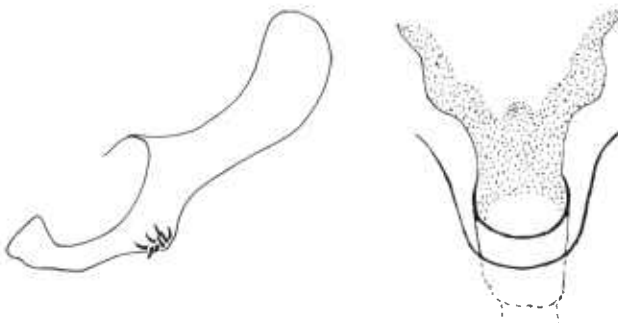
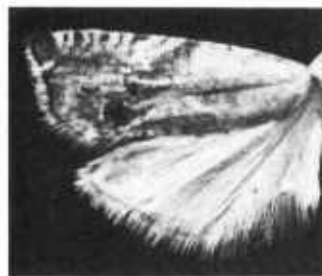






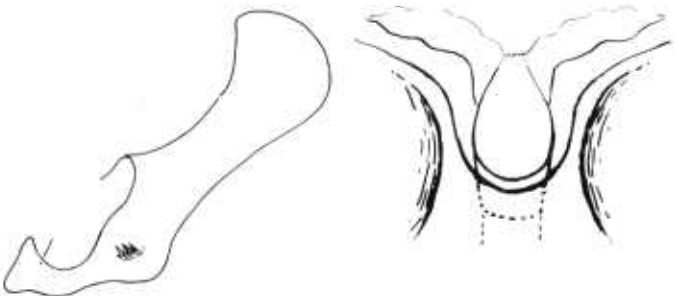
*Epinotia vertumnana* (Zeller)

Forewing 6.0 to 7.0 mm long, grayish yellowish brown. Adults captured March 23–April 26. MI. Larva feeds on *Crataegus*. Syst and Biol: Brown (1986). (31 N, 25 Gm, 6 Gf)



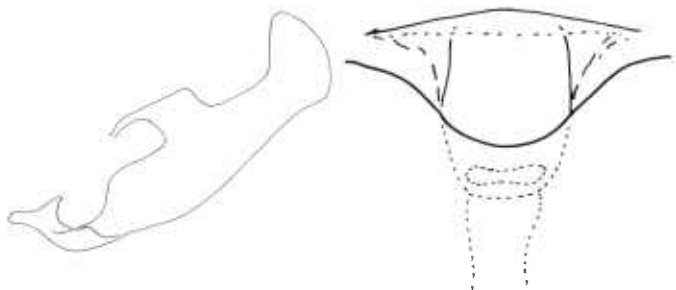
*Epinotia corylana* McDunnough

Forewing 5.0 to 6.5 mm long, dark markings yellowish brown, brown, or brownish black. Adults captured May 31–August 14. MI, MN. Larva feeds in *Corylus* catkins. Syst: McDunnough (1925a). Biol: MacKay (1959). (11 N, 7 Gm, 4 Gf, T, photo specimen Victoria Co., ON)



*Epinotia solicitana* (Walker)

Forewing 5.5 to 7.0 mm long, dark markings grayish brown or brownish black. Adults captured May 21–August 16. MI, WI, MN. Univoltine. Larva feeds in terminals or folded leaves of *Betula papyrifera*, *P. populifolia*. Syst: Brown (1980a). Biol: Smith (1946). (34 N, 12 Gm, 9 Gf, T)



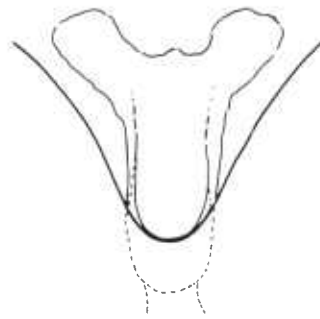
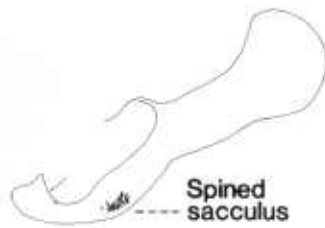
*Epinotia nisella* (Clerck)

Forewing 6.0 to 7.5 mm long, variable pattern; dark markings grayish brown or brownish black, sometimes orange in dorsal area. Adults captured July 26. MN. Larva feeds in female catkins and terminals of *Populus tremuloides* and *P. balsamifera*, respectively. Syst: Robinson and Nielsen (1983). Biol: Miller (1986a). (16 N, 2 Gm, 3 Gf)

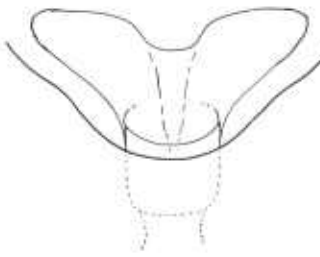


*Epinotia criddleana* (Kearfott)

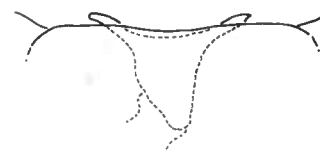
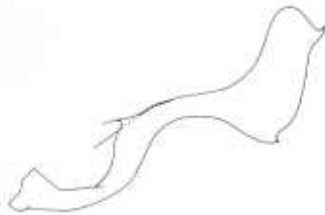
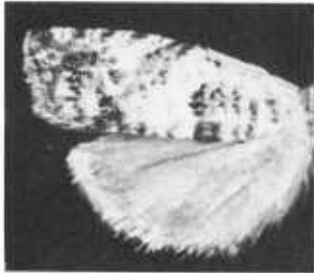
Forewing 6.5 to 7.5 mm long, variable pattern; dark markings gray or brownish black, sometimes orange in dorsal area. Adults captured July 18–August 26. MI, MN. Larva feeds in buds and tied leaves of *Populus*, *Salix*, *Quercus*. Syst and Biol: Kusch (1967). (10 N, 2 Gm, 5 Gf, T)



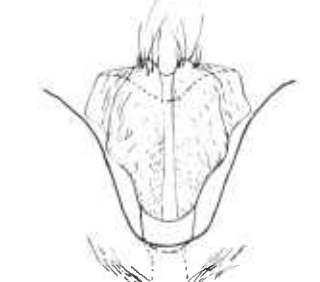
*Epinotia transmissana* (Walker)  
Forewing 6.0 to 8.0 mm long, dark areas grayish yellowish brown, grayish brown, or brownish black. Adults captured June 16–August 4. MI, WI, MN. Larva feeds on *Salix*. Syst: Brown (1980a). Biol: Prentice (1966). (26 N, 8 Gm, 7 Gf, T)



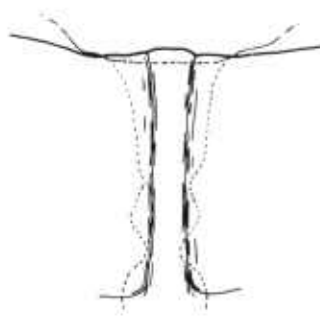
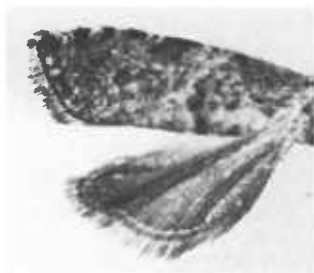
*Epinotia momonana* (Kearfott)  
Forewing 5.5 to 6.5 mm long, dark markings grayish brown or brownish black. Adults captured July 4–September 5. MI, WI, MN. Larva feeds on *Picea*. Syst: McDunnough (1935). Biol: Heinrich (1923b). (11 N, 5 Gm, 6 Gf, T)



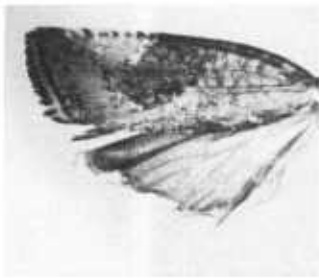
*Epinotia huroniensis* Brown  
Forewing 6.0 mm long, dark markings grayish brown or brownish black. Adults captured July 2–August 2. MI, MN. Syst: Brown (1980b). (7 N, 5 Gm, 1 Gf)



*Epinotia nonana* (Kearfott)  
Forewing 8.5 to 11.0 mm long, dark markings grayish brown or brownish black. Adults captured August 3–October 1. MI, MN. Syst: Heinrich (1929). (34 N, 6 Gm, 3 Gf, T)

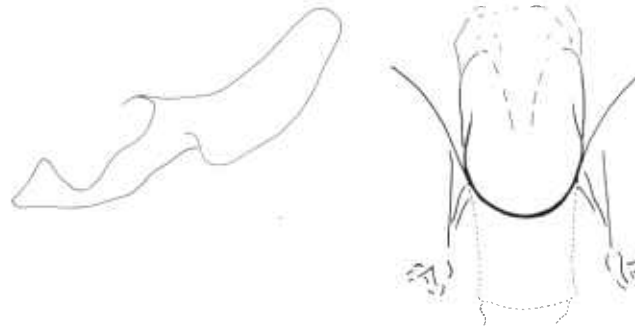


*Epinotia nanana* (Treitschke)  
Forewing 4.5 to 5.0 mm long, dark markings grayish yellowish brown or brownish black. Adults captured April 30–June 13. MI, WI. Univoltine. Larva feeds in *Picea* needles. Syst: Kuznetsov (1986). Biol: Daviault and Ducharme (1966). (76 N, 6 Gm, 4 Gf)



*Epinotia medioplagata*  
(Walsingham)

Forewing 5.5 to 7.5 mm long, dark markings brown. Adults captured July 16. MI, WI, MN. Larva feeds in rolled *Salix* leaves. Syst: Brown (1980a). Biol: Prentice (1966). (3 N, 2 Gm, 1 Gf, T)



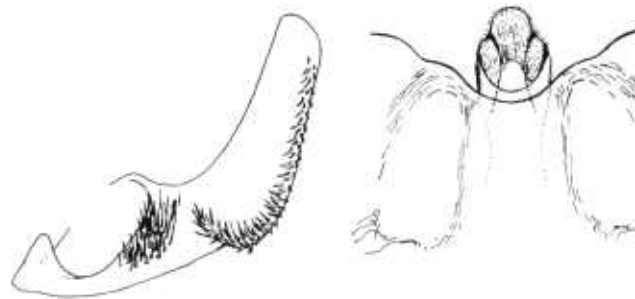
*Epinotia cruciana* (Linnaeus)

Forewing 5.0 to 5.5 mm long, dark areas brown. No capture dates. MN. Larva feeds in tied *Salix* leaves. Syst: Robinson and Nielsen (1983). Biol: Prentice (1966). (7 N, 2 Gm, 2 Gf)



*Epinotia septemberana* (Kearfott)

Forewing 7.5 mm long, dark areas brownish orange or brown. Adult captured August 23. MN. Larva feeds on *Rhododendron*, *Ledum groenlandicum*. Syst: Brown (1980a). Biol: Ferguson (1975). (1 N, 1 Gm, photo specimen Hancock Co., ME)



*Epinotia lindana* (Fernald)

Forewing 7.5 to 9.0 mm long, dark areas grayish brown or brownish black. Adults captured August 25–October 12. MI, WI, MN. Larva feeds in tied leaves of *Cornus*. Syst: Brown (1980a). Biol: McDunnough (1933). (32 N, 3 Gm, 3 Gf, T)

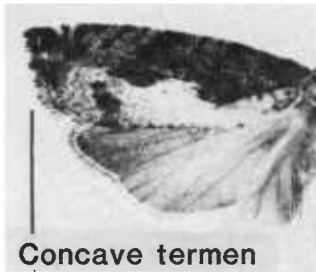
## Genus *Catastega*

**Both sexes.** Forewing termen concave, apex not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate or approximate,  $M_2$  and  $M_3$  approximate at base,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus bifid; socius not finger- or ribbonlike; valva not divided, sacculus densely clothed with spinelike setae, without rudimentary clasper, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin; anellus loosely surrounding aedeagus.

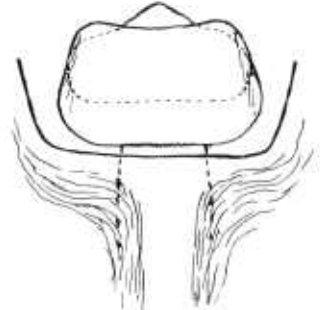
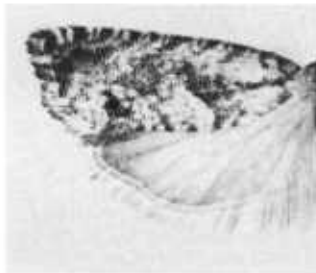
**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Lamella antevaginalis conical; lamella postvaginalis absent. Ductus bursae sclerotized one-third its length from genital opening; corpus bursae without sclerotized sides, with two thorn- or finlike signa.

**Comments.** Three Nearctic species of *Catastega* are known. Brown (1980a, 1986) gives a generic treatment.



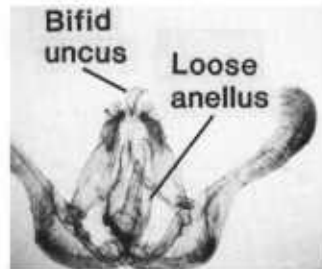
*Catastega timidella* Clemens

Forewing 7.0 to 9.0 mm long, dark markings grayish brown or brownish black. Adults captured May 13–July 10. MI, WI, MN. Larva feeds in serpentine tube on underside of *Quercus*, *Juglans*, *Betula* leaves. Syst: Brown (1986). Biol: Prentice (1966). (39 N, 9 Gm, 5 Gf)



*Catastega aceriella* Clemens. Maple trumpet skeletonizer

Forewing 6.5 to 8.5 mm long, dark markings grayish brown or brownish black. Adults captured May 20–July 20. MI, WI, MN. Univoltine. Larva feeds in serpentine tube on underside of leaves of *Acer*, *Crataegus*, *Fagus grandifolia*. Syst: Brown (1986). Biol: Côté and Allen (1973). (41 N, 14 Gm, 7 Gf)



## Genus *Ancylis*

**Both sexes.** Forewing without raised scale tufts, apex falcate, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate,  $M_2$  and  $M_3$  not connate,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked or united.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Socius present, not finger- or ribbon-like; valva not divided, without rudimentary clasper, sacculus not densely clothed with spinelike setae, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 not deeply emarginate around ostium bursae, neither inflected nor overlapping ostium bursae. Lamella antevaginalis not conical. Ductus bursae sclerotized less than two-thirds its length from genital opening; corpus bursae without sclerotized sides, with two bladelike signa.

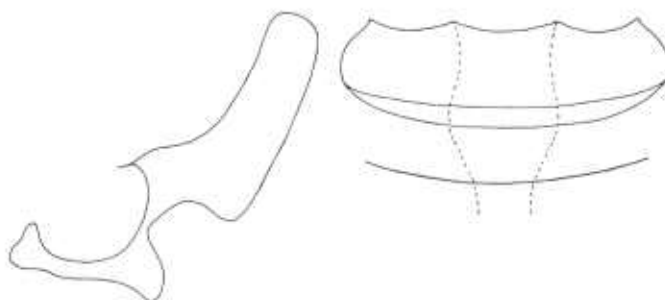
**Comments.** In the former *Anchylopera* species (Heinrich 1923b), hindwing  $M_3$  and  $Cu_1$  are united; in the remaining *Ancylis* species, these veins are stalked.

More than 30 Nearctic species of *Ancylis* are known. Among the species treated here, the following also occur in the Palearctic: *A. comptana*, *A. diminutana*, *A. unguicella*, and *A. tineana*.

I was unable to confirm Heinrich's (1923b) report of *Ancylis goodelliana* (Fernald) in the region.

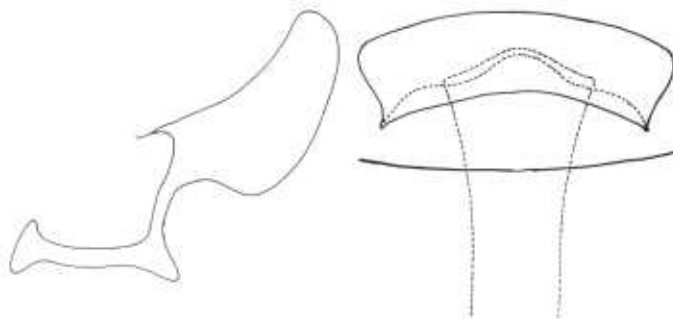
The *A. subaequana* complex here includes *A. galeamatana* (McDunnough) and *A. sheppardana* (McDunnough). The latter two are based on slightly differing specimens from one locality each (McDunnough 1956). The range of variability among representatives of the complex in this study made it difficult to rule out any constituent species.

The *A. burgessiana* complex here includes *A. laciniana* (Zeller) and *A. spiraeifoliana* (Clemens). Species limits in this group are ill defined at present, and identities of the latter two rely heavily on host data (McDunnough 1955). Host data were lacking for representatives of the complex in the Michigan-Wisconsin-Minnesota sample, and I could not satisfactorily distinguish the constituent species.



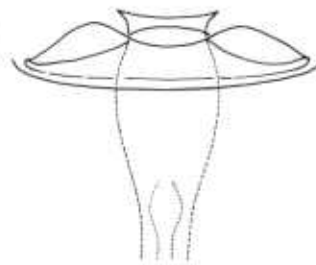
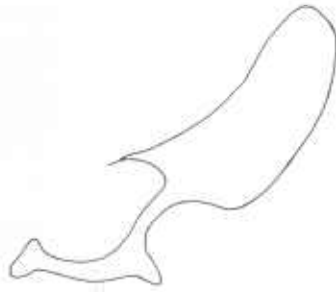
*Ancylis nubeculana* (Clemens)

Forewing 7.0 to 8.0 mm long, dark dorsal area grayish brown or brownish black. Adults captured April 24–July 5. MI, WI. Univoltine. Larva feeds in folded leaves of *Pyrus*, *Amelanchier*, *Crataegus*, others. Syst: Heinrich (1923b). Biol: Chapman and Lienk (1971). (36 N, 4 Gm, 3 Gf, T)



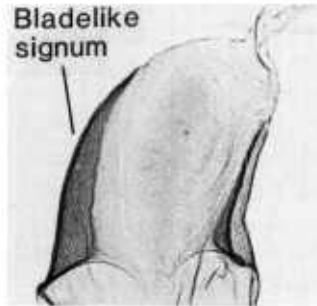
*Ancylis subaequana* (Zeller) complex

Forewing 6.0 to 7.5 mm long, dorsal spot brownish black. Male vesica long, subequal in length to valva. Adults captured May 20–August 3. MI, WI, MN. Syst: McDunnough (1956). (67 N, 13 Gm, 4 Gf, T)



*Ancylys metamelana* (Walker)

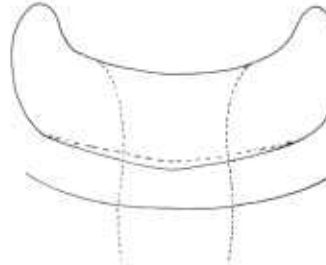
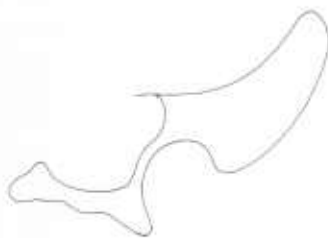
Forewing 4.5 to 5.5 mm long, dark areas yellowish brown or brownish black. Female ductus bursae not sclerotized. Adults captured May 8–September 2. MI, WI. Multivoltine. Larva feeds in folded or tied leaves of *Trifolium repens*, *T. pratense*, *T. hybridum*. Syst: McDunnough (1955). Biol: Wehrle (1929). (51 N, 4 Gm, 6 Gf, T)



Bladelike  
signum

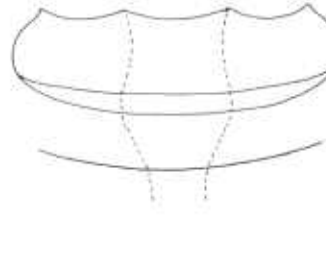
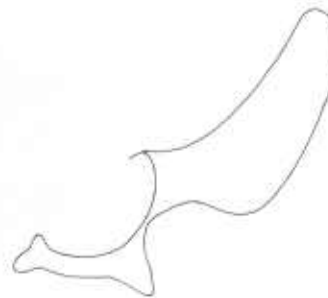


Falcate apex



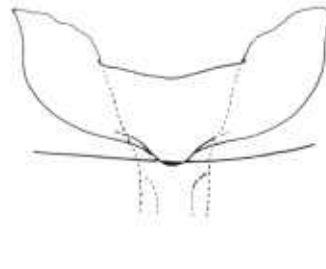
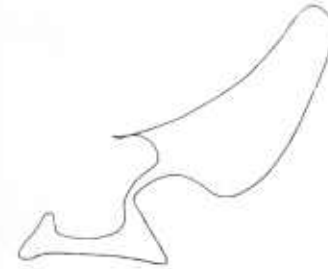
*Ancylys semiovana* (Zeller)

Forewing 6.5 to 7.5 mm long, dark areas brownish black. Adults captured May 29–August 30. MI, MN. Larva feeds on *Ceanothus*. Syst: Heinrich (1923b). Biol: MacKay (1959). (46 N, 3 Gm, 2 Gf, T)



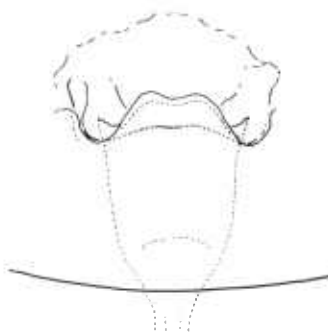
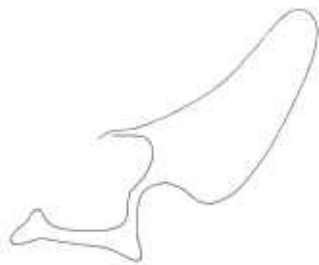
*Ancylys burgessiana* (Zeller)  
complex

Forewing 5.5 to 8.0 mm long, dorsal spot brown. Adults captured May 22–July 31. MI, WI, MN. Larva feeds in folded leaves of *Quercus*, *Corylus*, *Prunus*. Syst: Heinrich (1923b). Biol: Prentice (1966). (117 N, 11 Gm, 8 Gf, T)



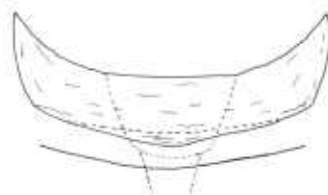
*Ancylys platanana* (Clemens)

Forewing 7.5 mm long, unmarked areas orange yellow. Adult captured June 13. MI. Larva feeds on *Platanus occidentalis* leaves. Syst: Heinrich (1923b). Biol: Denmark (1960). (1 N, 1 Gm, T)



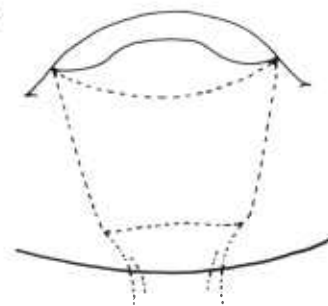
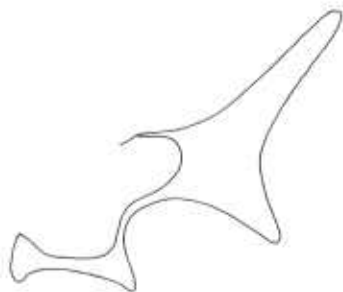
*Ancylys comptana* (Frölich).  
Strawberry leafroller

Forewing 4.5 to 7.0 mm long, dark areas grayish brown, yellowish brown, or brownish black. Adults captured May 3–August 8. MI, WI, MN. Multivoltine. Larva feeds in folded *Fragaria*, *Rubus* leaves. Syst: Miller (1973b). Biol: Neiswander (1944). (39 N, 8 Gm, 5 Gf)



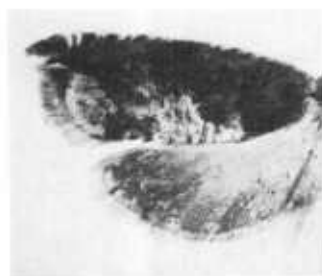
*Ancylys divisana* (Walker)

Forewing 4.5 to 7.0 mm long, dark areas reddish brown. Female ductus bursae not sclerotized. Adults captured June 4–August 7. MI, WI. Larva feeds in rolled leaves of *Quercus*, *Platanus occidentalis*. Syst: Heinrich (1923b). Biol: Forbes (1923). (9 N, 2 Gm, 2 Gf, T)



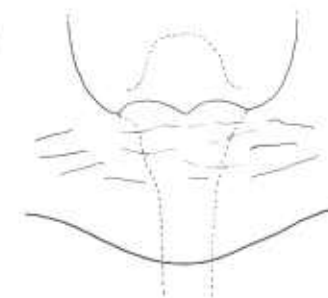
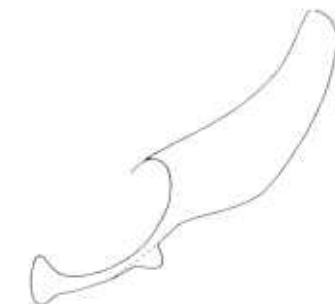
*Ancylys apicana* (Walker)

Forewing 4.5 to 7.0 mm long, dark areas grayish reddish brown. Female ductus bursae not sclerotized. Adults captured May 19–June 24. MI, WI, MN. Larva feeds in folded leaves of *Rubus*, *Pyrus*, *Betula*, others. Syst: Heinrich (1923b). Biol: Chapman and Lienk (1971). (21 N, 2 Gm, 4 Gf, T)



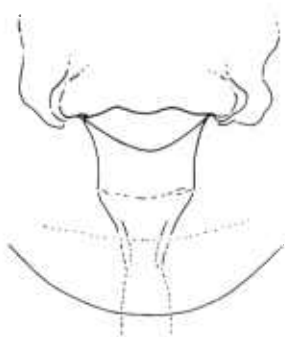
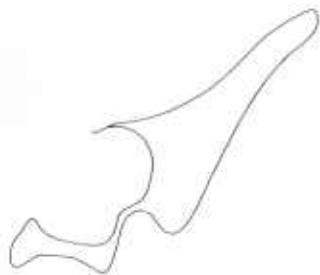
*Ancylys muricana* (Walsingham)

Forewing 4.0 mm long, dark basal area grayish reddish brown. Female ductus bursae not sclerotized. Adult captured July 20. MI. Larva feeds on *Fragaria*, *Cornus*, *Rubus*, others. Syst: Heinrich (1923b). Biol: Putman (1942). (1 N, 1 Gm, T, photo specimen Erie Co., NY)



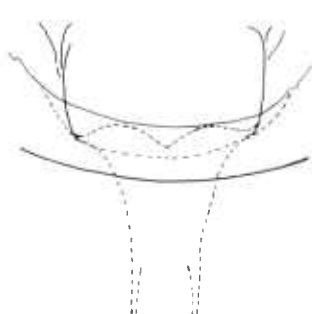
*Ancylys carbonana* Heinrich

Forewing 6.0 to 6.5 mm long, dark areas grayish brown. Adults captured May 23–June 21. MI. Syst: Heinrich (1923b). (8 N, 5 Gm, 1 Gf, T)



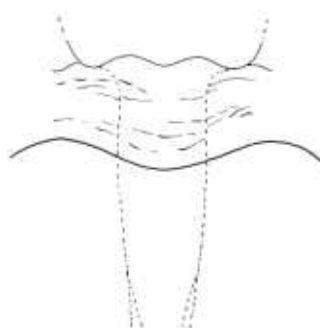
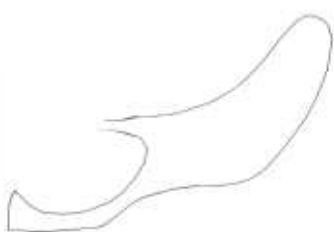
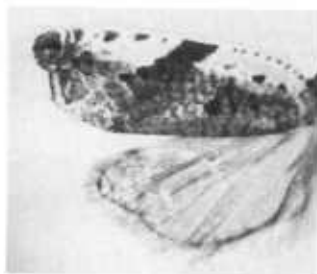
*Ancylys diminutana* (Haworth)

Forewing 6.0 to 8.0 mm long, dark areas yellowish brown or grayish yellowish brown. Female ductus bursae not sclerotized. Adults captured May 20–August 18, MI, WI. Larva feeds on *Salix*. Syst: Heinrich (1923b). Biol: MacKay (1959). (13 N, 5 Gm, 3 Gf, photo specimen Jasper Co., MO)



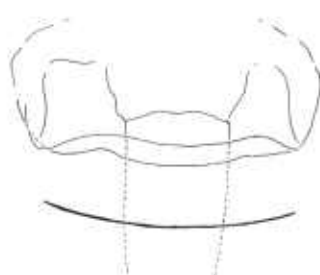
*Ancylys unguicella* (Linnaeus)

Forewing 6.5 to 8.5 mm long, dark markings yellowish brown, grayish yellowish brown, or brownish black. Adults captured May 18–July 13, MI. Syst: Robinson and Nielsen (1983). (3 N, 3 Gm)



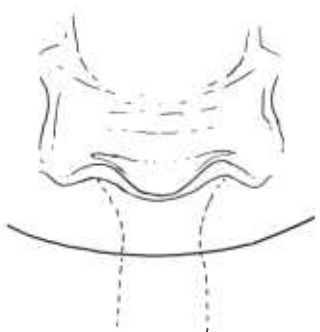
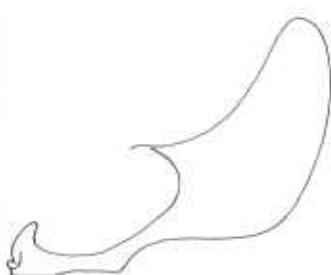
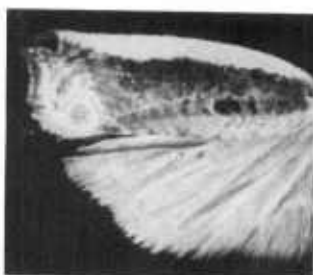
*Ancylys mediofasciana* (Clemens)

Forewing 7.0 to 9.5 mm long, dark areas and markings yellowish brown, grayish yellowish brown, or brownish black. Female ductus bursae not sclerotized. Adults captured May 20–June 6, MI, MN. Larva feeds in rolled *Pyrus* leaves. Syst: Heinrich (1923b). Biol: Ferguson (1975). (17 N, 3 Gm, 4 Gf, T)



*Ancylys tineana* (Hübner)

Forewing 6.5 to 7.0 mm long, dark areas yellowish brown or grayish yellowish brown. Adults captured May 20–July 1, MI. Larva feeds on *Populus*, *Prunus*. Syst: Heinrich (1923b). Biol: Ferguson (1975). (3 N, 1 Gm, 1 Gf, photo specimen Aweme, MB)



*Ancylys albacostana* Kearfott

Forewing 7.5 to 8.5 mm long, dark areas grayish brown or brownish black. Adults captured May 29–June 30, MI, MN. Syst: Heinrich (1923b). (6 N, 2 Gm, T)



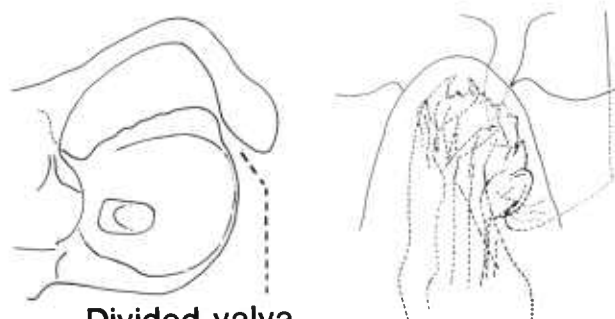
## Genus *Hystricophora*

**Both sexes.** Forewing without raised scale tufts, termen concave, apex not falcate,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_4$  and  $R_5$  separate,  $M_2$  and  $M_3$  separate at base,  $M_2$ ,  $M_3$ , and  $Cu_1$  not approximate at termen,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_2$  and  $M_3$  approximate,  $M_3$  and  $Cu_1$  stalked.

**Male.** Antenna not notched near base. Forewing without costal fold. Hindwing without upper-surface melanic sex scaling. Uncus developed; socius and hamus absent; valva divided, without rudimentary clasper, outer surface lacking spinelike setae, without thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 inflected and overlapping ostium bursae, without bilobed projection. Ductus bursae sclerotized two-thirds or more of its length from genital opening; corpus bursae without sclerotized sides, without signa.

**Comments.** More than 10 Nearctic species of *Hystricophora* are known.



Divided valva

*Hystricophora vestaliana* (Zeller)

Forewing 10.0 mm long, pale areas yellowish white or pale yellow. Adult captured June 30. Wl. Syst: Heinrich (1923b). (1 N, 1 Gf, T, photo specimen lectotype)

## Tribe Grapholitini

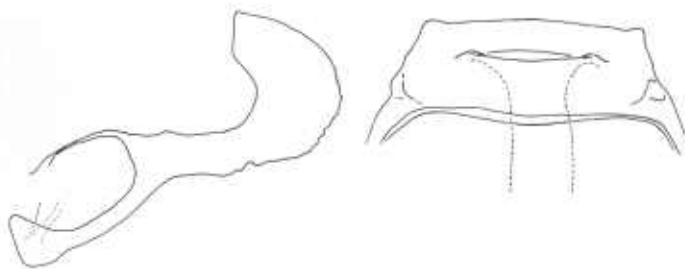
### Genus *Dichrorampha*

**Both sexes.** Thorax without posterior tuft. Forewing termen slightly concave,  $R_1$  originating well before middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  separate, cubital vein with pecten,  $M_3$  and  $Cu_1$  connate or stalked.

**Male.** Hindwing inner margin not modified. Hindmost tibia without dilated or tufted scaling. Abdominal segment 8 without paired lateral tufts. Uncus not developed; socius and hamus absent; valva without rudimentary clasper, without thick spinelike seta on cucullus at lower margin; vesica with cornuti.

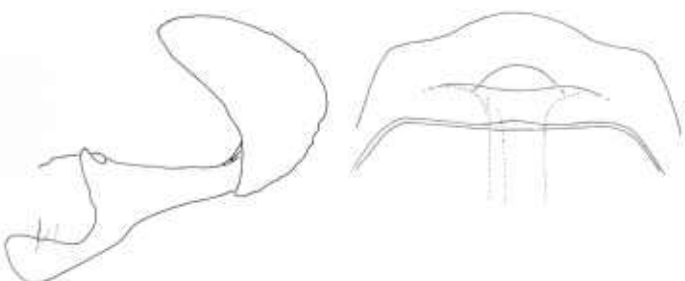
**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Ductus bursae sclerotized one-half or less its length from genital opening; corpus bursae with one thornlike signum.

**Comments.** Nine Nearctic species of *Dichrorampha* are known. Among the species treated here, *D. sedatana* also occurs in the Palearctic. Part of the genus was reviewed by Miller (1983d).



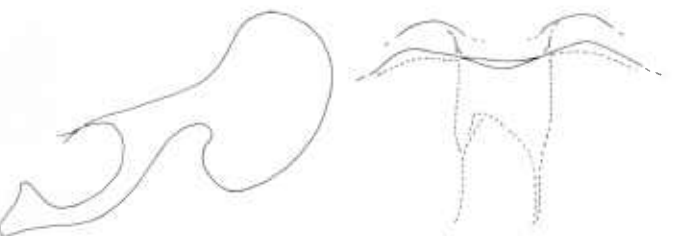
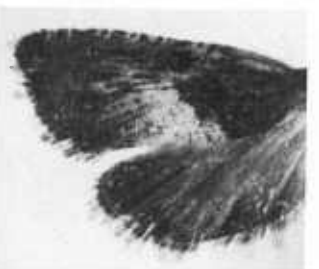
*Dichrorampha simulana*  
(Clemens)

Forewing 5.5 to 7.0 mm long, variable pattern; dark areas yellowish brown. Limits of variation shown. Adults captured June 26–August 7. MI, WI, MN. Female tentatively associated. Syst: Miller (1983d). (3 N, 1 Gm, 2 Gf, T, top photo specimen Moraine Lake, AB, bottom, Lake City, CO)



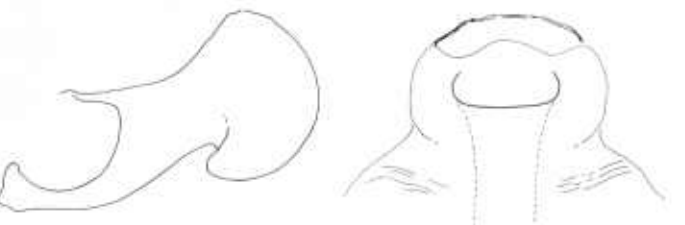
*Dichrorampha bittana* (Busck)

Forewing 5.0 to 6.5 mm long, variable pattern; dark areas yellowish brown. Limits of variation shown. Adults captured June 25–August 26. MI, WI. Female tentatively associated. Syst: Miller (1983d). (7 N, 7 Gm, T, bottom photo specimen Grundy Co., MO)



*Dichrorampha incanana*  
(Clemens)

Forewing 4.5 to 5.0 mm long, dark areas grayish brown or brownish black. Adults captured July 17–19. MI, WI. Syst: Heinrich (1926). (2 N, 2 Gm, T, photo specimen Putnam Co., IL)



*Dichrorampha sedatana* (Busck)

Forewing 5.0 to 6.0 mm long, dark areas yellowish brown. Adults captured June 8–13. MI. Syst: Miller (1983d). (5 N, 3 Gm, 1 Gf, T)

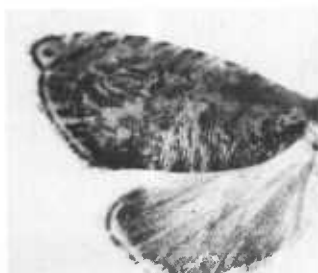
## Genus *Talponia*

**Both sexes.** Thorax without posterior tuft. Forewing termen concave,  $R_1$  originating at or nearer middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  not approximate,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  separate, cubital vein with pecten,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin not modified. Hindmost tibia without dilated or tufted scaling. Abdominal segment 8 with paired lateral tufts. Uncus not developed; socius present; valva without rudimentary clasper, without thick spinelike seta on cucullus at anal angle; vesica without cornuti.

**Female.** Sternum 7 inflected and overlapping ostium bursae. Ductus bursae sclerotized only near genital opening; corpus bursae with two thornlike signa.

**Comments.** *Talponia* appears to be monotypic.



*Talponia plummeriana* (Busck)

Forewing 5.5 mm long, distal half brown. Adult captured July 8. MI. Larva feeds in *Asimina* flowers. Syst: Heinrich (1926). Biol: MacKay (1959). (1 N, 1 Gm, T, photo specimen Clay Co., MO)

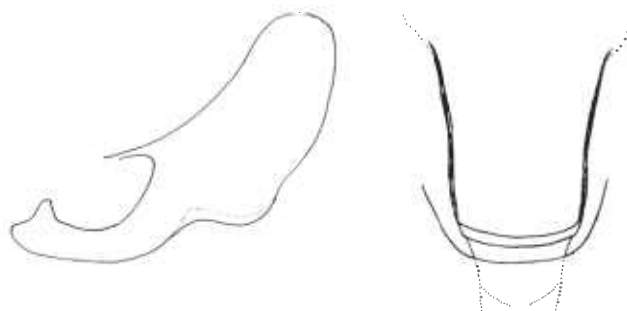
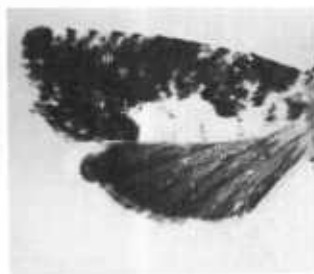
## Genus *Pammene*

**Both sexes.** Thorax without posterior tuft. Forewing termen straight or slightly concave,  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate, cubital vein with pecten,  $M_3$  and  $Cu_1$  connate or stalked.

**Male.** Hindwing inner margin not modified. Hindmost tibia without dilated or tufted scaling. Abdominal segments 6 and 7 with modified dorsal hair tufts beneath scaling. Abdominal segment 8 without paired lateral tufts. Uncus not developed; socius and hamus absent; valva without rudimentary clasper, without thick spinelike seta on cucullus at lower margin; vesica with cornuti.

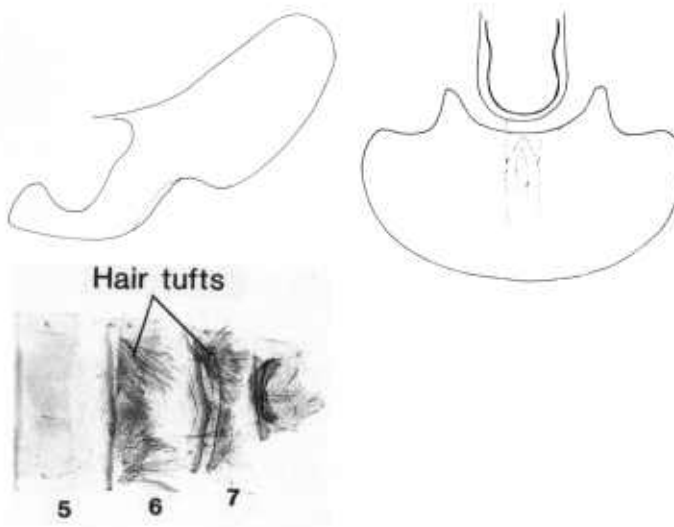
**Female.** Sternum 7 emarginate posteriorly, neither inflected nor overlapping ostium bursae. Ductus bursae sclerotized one-third its length from genital opening; corpus bursae with two thornlike signa.

**Comments.** Five Nearctic species of *Pammene* are known.



*Pammene felicitana* (Heinrich)

Forewing 5.0 to 6.0 mm long, dark areas grayish yellowish brown or brownish black. Adults captured June 2-12. MI. Syst: Heinrich (1926). (6 N, 1 Gm, 1 Gf, T)



*Pammene perstructana* (Walker)

Forewing 4.5 to 7.0 mm long, dark areas grayish yellowish brown or brownish black. Adults captured June 7–August 1. MI, WI, MN. Syst: Miller (1985e). (21 N, 9 Gm, 6 Gf, T)

### Genus *Eucosmomorpha*

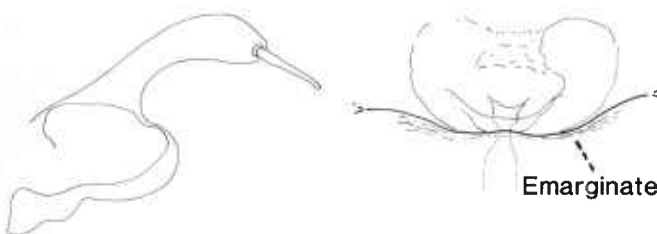
**Both sexes.** Thorax without posterior tuft. Forewing termen concave,  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  not approximate,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing  $R_s$  and  $M_1$  approximate or connate, cubital vein with pecten,  $M_3$  and  $Cu_1$  approximate or connate.

**Male.** Hindwing inner margin modified into a sclerotized edge. Hindmost tibia without dilated or tufted scaling. Abdominal segment 8 without paired lateral tufts. Uncus not developed; socius present; valva without rudimentary clasper, with thick spinelike seta on cucullus at lower margin; vesica with cornuti.

**Female.** Sternum 7 slightly emarginate posteriorly, neither inflected nor overlapping ostium bursae. Ductus bursae sclerotized near middle; corpus bursae with one thornlike signum.

**Comments.** Only one Nearctic species of *Eucosmomorpha* is known; *E. albersana* also occurs in the Palearctic. Obratzsov (1961) gives a generic treatment.

Placement of *Eucosmomorpha* in the tribe Grapholitini is based on the prevailing venational criteria (Heinrich 1923b, Obratzsov 1958). Use of other characters in determining higher category relations may affect the future placement of this and other genera (Kuznetsov and Stekolnikov 1977).



*Eucosmomorpha albersana* (Hübner)

Forewing 5.5 mm long, dark markings brown or grayish brown. Adult captured June 2. MI. Syst: Miller (1983c). (1 N, 1 Gm)

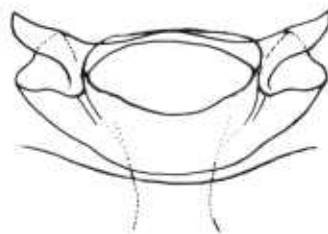
## Genus *Larisa*

**Both sexes.** Thorax without posterior tuft. Forewing termen convex,  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  not approximate,  $Cu_2$  originating at distal two-thirds or between distal two-thirds and three-quarters of discal cell. Hindwing Rs and  $M_1$  stalked, cubital vein with pecten,  $M_3$  and  $Cu_1$  stalked.

**Male.** Hindwing inner margin not modified. Hindmost tibia without dilated or tufted scaling. Abdominal segment 8 with paired lateral tufts. Uncus developed; hamus present; valva with rudimentary clasper, without thick spinelike seta on cucullus at lower margin; vesica with cornuti.

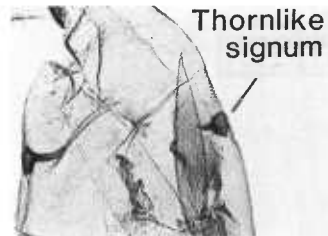
**Female.** Sternum 7 emarginate posteriorly, neither inflected nor overlapping ostium bursae. Ductus bursae sclerotized two-thirds its length from genital opening; corpus bursae with two thornlike signa.

**Comments.** *Larisa* appears to be monotypic. Miller (1978a) gives a generic treatment.



*Larisa subsolana* Miller

Forewing 4.5 to 5.0 mm long, dark areas brown or brownish black. Adults captured June 12–August 5. MI. Larva feeds on *Carya*. Syst: Miller (1978a). Biol: Brown *et al.* (1983). (4 N, 3 Gm, 1 Gf, T)



Thornlike signum

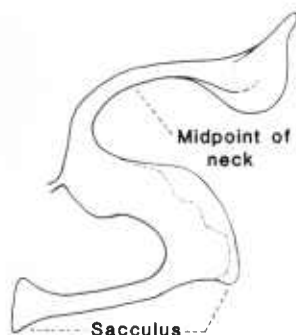
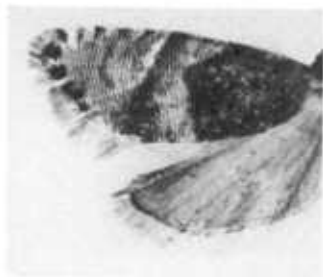
## Genus *Sereda*

**Both sexes.** Thorax without posterior tuft. Forewing termen slightly concave,  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing Rs and  $M_1$  approximate, cubital vein without pecten,  $M_3$  and  $Cu_1$  connate.

**Male.** Hindwing inner margin not modified. Hindmost tibia without dilated or tufted scaling. Abdominal segment 8 with paired lateral tufts. Uncus not developed; socius and hamus absent; valva without rudimentary clasper, without thick spinelike seta on cucullus at lower margin, cross-sectional width at mid-point of neck one-eighth or less of lineal sacculus width; vesica with cornuti.

**Female.** Sternum 7 emarginate posteriorly, neither inflected nor overlapping ostium bursae. Ductus bursae sclerotized two-thirds its length from genital opening; corpus bursae with two thornlike signa.

**Comments.** *Sereda* appears to be monotypic. The species name *tautana* has usually been misspelled *lautana* (Miller 1973a).



*Sereda tautana* (Clemens)

Forewing 5.0 to 5.5 mm long, basal half brown. Adults captured April 13–May 23. MI. Larva feeds on *Quercus*. Syst: Heinrich (1926). Biol: Prentice (1966). (11 N, 5 Gm, 3 Gf, T)

## Genus *Grapholita*

**Both sexes.** Thorax without posterior tuft. Forewing  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate, cubital vein with pecten,  $M_3$  and  $Cu_1$  connate or stalked.

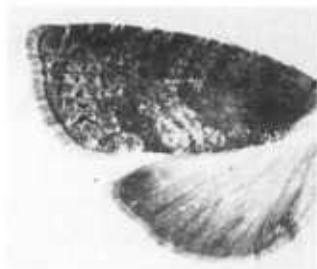
**Male.** Hindwing inner margin not modified. Hindmost tibia without dilated or tufted scaling. Abdominal segment 8 with paired lateral tufts. Uncus not developed; socius and hamus absent; valva without rudimentary clasper, without thick spinelike seta on cucullus at lower margin; vesica with cornuti.

**Female.** Sternum 7 neither inflected nor overlapping ostium bursae. Corpus bursae with two thornlike signa or without signa.

**Comments.** Nearly 20 Nearctic species of *Grapholita* are known. Among the species treated here, *G. molesta* and *G. delineana* also occur in the Palearctic, the former being accidentally introduced to the Nearctic sometime before 1915. The latter is a possible introduction (Miller 1982).

I was unable to confirm Beebe's (1954) report of *G. libertana* Heinrich in the region.

*Grapholita packardi* is considered a complex here because of diverse habits and hosts (Chapman and Lienk 1971).



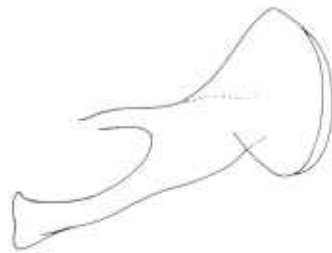
*Grapholita molesta* (Busck). Oriental fruit moth

Forewing 5.0 to 5.5 mm long, dark areas yellowish brown or grayish yellowish brown. Female corpus bursae with two signa. Adults captured May 6–July 4. MI. Multivoltine. Larva feeds in terminals and fruits of *Prunus*, *Pyrus*, *Rosa*, others. Syst: Heinrich (1926). Biol: Chapman and Lienk (1971). (5 N, 3 Gm, 2 Gf, T, photo specimen Story Co., IA)



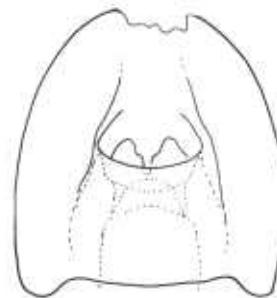
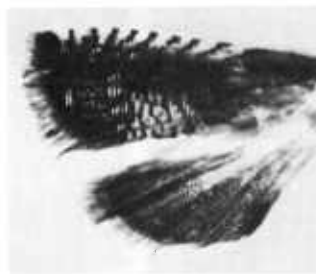
*Grapholita packardii* (Zeller) complex. Cherry fruitworm

Forewing 4.0 to 5.5 mm long, dark areas yellowish brown or grayish yellowish brown. Female corpus bursae with two signa. Sex scaling on underforewing and upper-hindwing surfaces of male. Adults captured May 23–August 24. MI, WI, MN. Uni- or bivoltine depending on host. Larva feeds in fruits and terminals of *Pyrus*, *Prunus*, *Crataegus*, others. Syst: Heinrich (1926). Biol: Balduf (1959). (37 N, 10 Gm, 8 Gf, T)



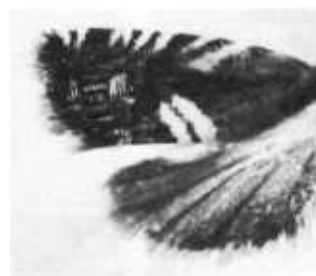
*Grapholita prunivora* (Walsh). Lesser appleworm

Forewing 4.0 to 5.0 mm long, dark areas yellowish brown or brownish black. Female corpus bursae with two signa. Adults captured May 27–August 11. MI, WI, MN. Bivoltine. Larva feeds in fruits of *Pyrus*, *Prunus*, *Crataegus*. Syst: Heinrich (1926). Biol: Chapman and Lienk (1971). (119 N, 5 Gm, 4 Gf, T)



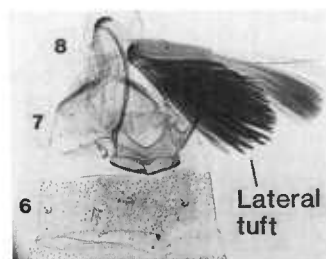
*Grapholita angleseana* (Kearfott)

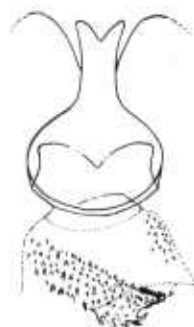
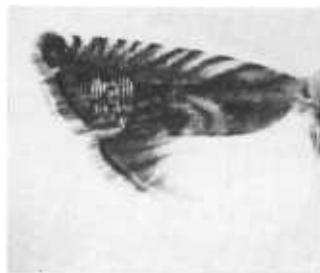
Forewing 5.0 mm long, dark areas and markings grayish yellowish brown or brownish black. Female corpus bursae with two signa. Adult captured June 20. MI. Larva feeds on seeds of *Fragaria*. Syst and Biol: Heinrich (1926). (1 N, 1 Gm, T)



*Grapholita fana* (Kearfott)

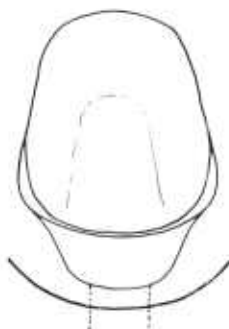
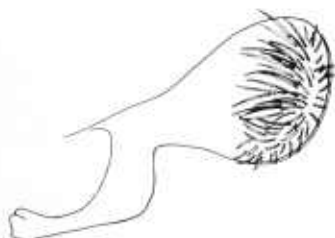
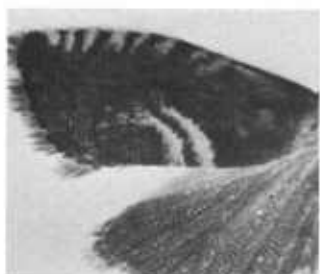
Forewing 4.5 to 5.0 mm long, dark areas grayish yellowish brown. Female corpus bursae with two signa. Adults captured May 24–June 16. MI. Larva feeds in *Desmodium* buds and flowers. Syst and Biol: Heinrich (1926). (3 N, 2 Gm, 1 Gf, T)





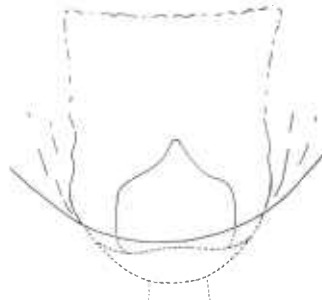
*Grapholita eclipsana* (Zeller)

Forewing 4.0 to 6.0 mm long, dark areas and markings yellowish brown or brownish black. Female corpus bursae with two signa. Adults captured April 21–June 10. MI, WI, MN. Larva feeds on *Amorpha*. Syst: Heinrich (1926). Biol: Godfrey et al. (1987). (23 N, 6 Gm, 3 Gf, T)



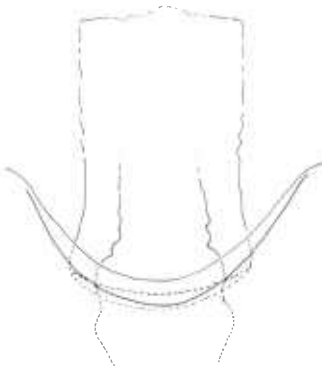
*Grapholita interstinctana* (Clemens). Clover head caterpillar

Forewing 4.0 to 5.0 mm long, dark areas yellowish brown. Signa absent from female corpus bursae. Adults captured May 30–September 6. MI, WI, MN. Multivoltine. Larva feeds on *Trifolium* heads and leaves. Syst: Heinrich (1926). Biol: Wehrle (1924). (46 N, 6 Gm, 6 Gf, T)



*Grapholita tristrigana* (Clemens)

Forewing 4.5 to 6.0 mm long, dark areas grayish yellowish brown. Signa absent from female corpus bursae or no larger than specks. Adults captured May 24–July 3. MI. Larva feeds in seeds and stems of *Baptisia*, *Lupinus*. Syst and Biol: Miller (1982). (2 N, 2 Gm, T, photo specimen Gentry Co., MO)



*Grapholita delineana* Walker

Forewing 4.5 to 6.0 mm long, dark areas grayish yellowish brown. Female corpus bursae with two signa. Adults captured June 19–July 22. WI, MN. Larva feeds in stems and seeds of *Cannabis sativa*. Syst and Biol: Miller (1982). (13 N, 5 Gm, 2 Gf, photo specimen Putnam Co., IL)

Genus *Corticivora*

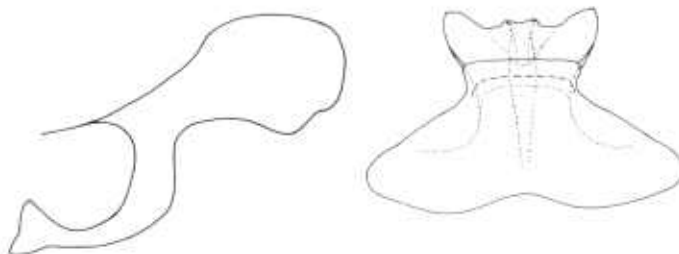
**Both sexes.** Thorax without posterior tuft. Forewing termen slightly convex,  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $R_3$  and  $R_4$  approximate,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  stalked, cubital vein with pecten,  $M_3$  and  $Cu_1$  stalked.

**Male.** Hindwing inner margin not modified. Hindmost tibia without dilated or tufted scaling. Abdominal segment 8 without paired lateral tufts. Uncus not developed; socius present; valva without rudimentary clasper, without thick spinelike seta on cucullus at lower margin; vesica without cornuti.



**Female.** Sternum 7 inflected and overlapping ostium bursae. Ductus bursae sclerotized one-third its length from genital opening; corpus bursae with two scobinate signa.

**Comments.** Three Nearctic species of *Corticivora* are known. Brown (1984) gives a generic treatment and reviews the species of *Corticivora*.



*Corticivora clarki* Clarke

Forewing 4.0 to 5.0 mm long, dark areas grayish yellowish brown. Adults captured July 5–August 10. MI. Larva feeds on *Pinus*. Syst: Brown (1984). Biol: Clarke (1951). (5 N, 4 Gm, T)

## Genus *Cydia*

**Both sexes.** Thorax without posterior tuft. Forewing  $R_1$  originating at or near middle of discal cell,  $R_2$  originating nearer  $R_3$  than  $R_1$ , upper internal vein of discal cell originating between  $R_1$  and  $R_2$ ,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate,  $M_3$  and  $Cu_1$  connate, stalked, or united.

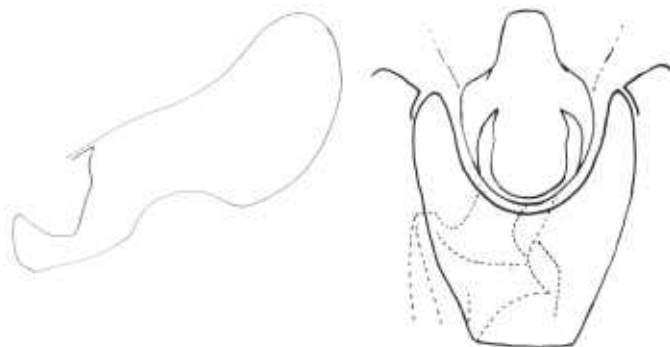
**Male.** Abdominal segments 6 and 7 without modified dorsal hair tufts beneath scaling. Abdominal segment 8 without paired lateral tufts. Uncus not developed; socius and hamus absent; valva without rudimentary clasper, without thick spinelike seta on cucullus at lower margin.

**Female.** Sternum 7 emarginate posteriorly or deeply emarginate around ostium bursae, neither inflected nor overlapping ostium bursae. Corpus bursae with two thornlike signa.

**Comments.** More than 40 Nearctic species of *Cydia* are known. Among the species treated here, *C. pomonella* is cosmopolitan, and *C. rusticella* (formerly *C. nigricana*) and *C. strobilella* occur in the Palearctic. The first two were accidentally introduced to the Nearctic: the former before 1819, the latter before 1900.

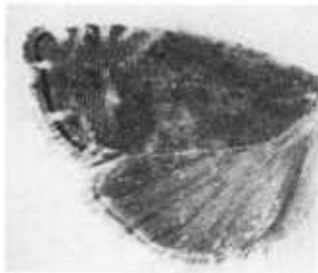
Brown (1983) discusses some generic characters.

*Cydia toreuta* and *C. latiferreana* are considered complexes here. The former may consist of different species on different *Pinus* hosts (Abrahamson and Kraft 1965). In the latter, genitalic variation may represent additional but as yet unrecognized species (Brown 1983).



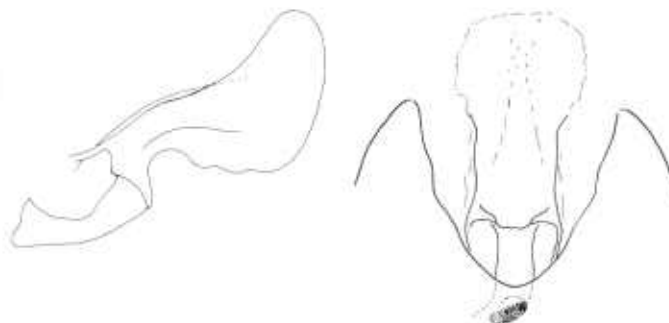
*Cydia laricana* (Busck)

Forewing 5.0 mm long, dark areas and markings grayish brown or brownish black. Adult captured August 24. WI. Larva feeds beneath *Larix* bark. Syst and Biol: Heinrich (1926). (1 N, 1 Gm, T, photo specimen Missoula Co., MT)



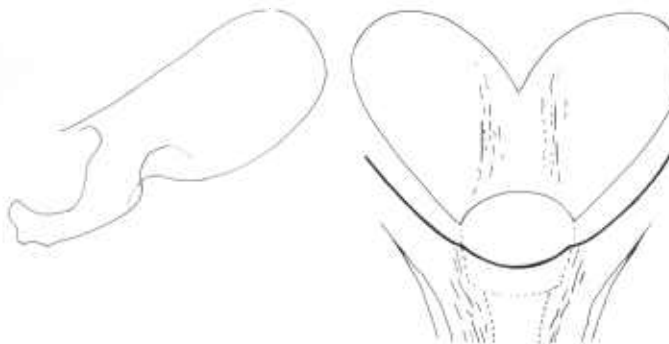
*Cydia inopiosa* Heinrich

Forewing 4.0 to 5.0 mm long, dark areas and markings grayish brown or brownish black. No capture dates; all adults reared. MI, WI. Larva feeds beneath *Pinus* bark. Syst and Biol: Brown and W.E. Miller (1983). (17 N, 3 Gm, 5 Gf, T)



*Cydia garacana* (Kearfott)

Forewing 5.5 to 6.5 mm long, dark areas grayish brown. Adults captured June 20–July 20. MI, MN. Larva feeds on *Populus*. Syst: Heinrich (1926). Biol: Brown et al. (1983). (9 N, 6 Gm, T)



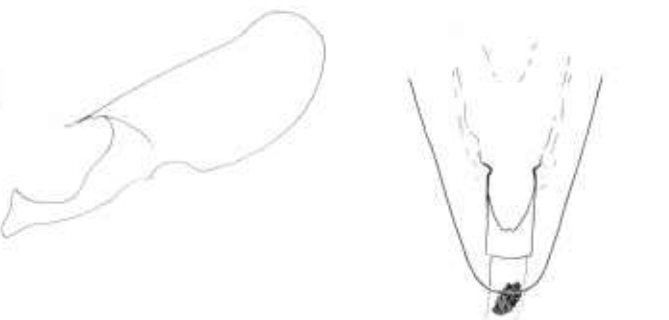
*Cydia multilineana* (Kearfott)

Forewing 6.0 to 7.0 mm long, dark areas grayish brown. Adults captured June 12–29. MI. Syst: Heinrich (1926). (3 N, 1 Gm, 2 Gf, T)



*Cydia albimaculana* (Fernald)

Forewing 5.0 mm long, dark areas and markings grayish brown or brownish black. Adults captured May 8–20. MI. Syst: Heinrich (1926). (2 N, 2 Gm, T)

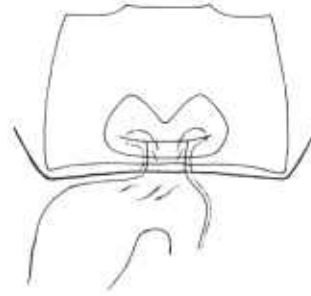


*Cydia populana* (Busck)

Forewing 5.0 to 7.5 mm long, dark areas grayish brown. Adults captured June 12–July 31. MI. Larva feeds beneath *Populus* bark. Syst and Biol: Heinrich (1926). (18 N, 5 Gm, 5 Gf, T)

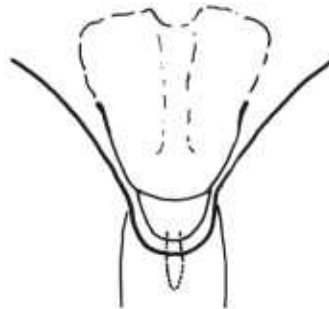


Male  
unknown



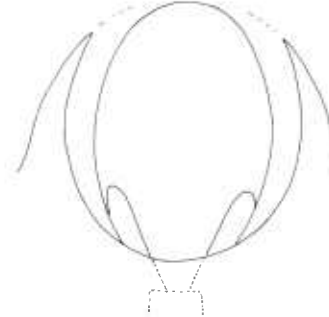
*Cydia lacustrina* (Miller)

Forewing 8.5 to 9.5 mm long, dark markings dark grayish brown. Adults captured June 6–15. MI. Syst: Miller (1976b). (5 N, 3 Gf, T)



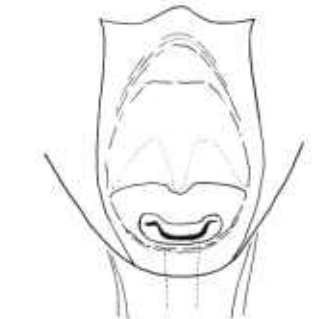
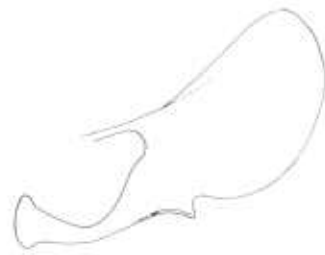
*Cydia flexiloqua* (Heinrich)

Forewing 6.5 to 7.5 mm long, dark markings dark grayish brown. Adults captured July 3–August 29. MI, WI, MN. Syst: McDunnough (1944). (6 N, 4 Gm, 2 Gf, T, photo specimen York Co., ON)



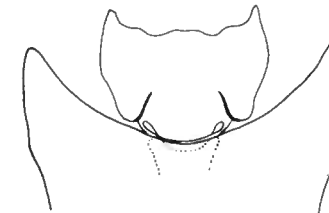
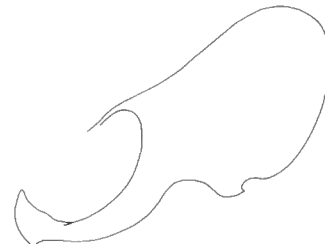
*Cydia strobilella* (Linnaeus).  
Spruce seed moth

Forewing 3.0 to 5.0 mm long, dark markings dark grayish brown. No capture dates; all adults reared. MI, MN. Larva feeds on *Picea* seeds and cones. Syst: Brown and W.E. Miller (1983). Biol: Tripp (1954). (9 N, 2 Gm, 2 Gf, photo specimen Carleton Co., ON)



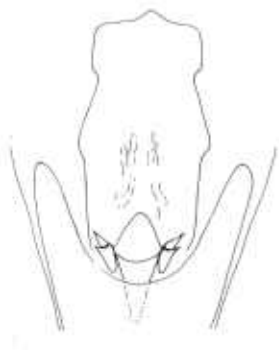
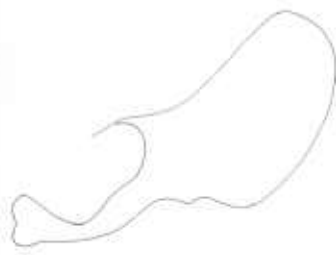
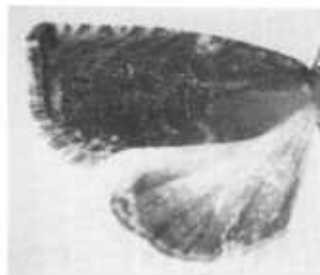
*Cydia rusticella* (Clerck). Pea moth

Forewing 5.5 to 6.5 mm long, dark areas yellowish brown. Adults captured June 22–August 20. MI, MN. Univoltine. Larva feeds in developing *Pisum* seed pods. Syst: Robinson and Nielsen (1983). Biol: Fluke (1921). (7 N, 3 Gm, 3 Gf)



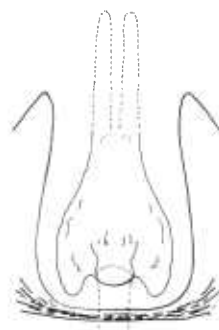
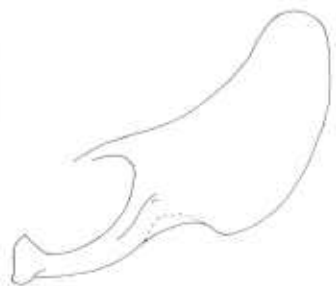
*Cydia candana* (Forbes)

Forewing 7.5 mm long, dark markings brown. Male hindwing cubital pecten in a pocket. Adult captured May 15. MN. Larva feeds in *Acer* seeds. Syst: Heinrich (1926). Biol: MacKay (1959). (1 N, 1 Gm, T, photo specimen Boone Co., MO)



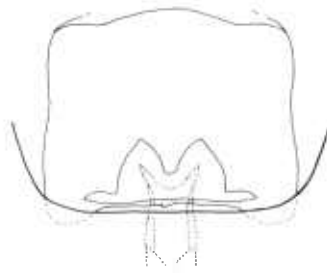
*Cydia caryana* (Fitch). Hickory shuckworm

Forewing 5.0 to 6.5 mm long, dark areas brownish black. Male lacks hindwing cubital pecten. Adults captured May 26–September 6. MI. Probably univoltine with protracted emergence period. Larva feeds in husks and fruits of *Carya*, *C. illinoensis*. Syst: Heinrich (1926). Biol: Payne and Heaton (1975). (8 N, 5 Gm, 2 Gf, T, photo specimen St. Louis Co., MO)



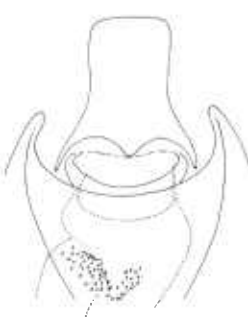
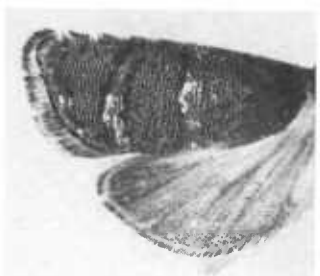
*Cydia latiferreana* (Walsingham) complex. Filbertworm

Forewing 6.0 to 9.5 mm long, pale areas orange or brown. Male with dilated scaling on hindmost tibia, without vesical cornuti, with hindwing cubital pecten in a pocket. Adults captured May 3–October 1. MI, WI, MN. Multivoltine. Varieties F and G of Heinrich (1926) confirmed. Larva feeds in *Quercus* acorns. Syst and Biol: Brown (1983). (50 N, 9 Gm, 3 Gf, T)



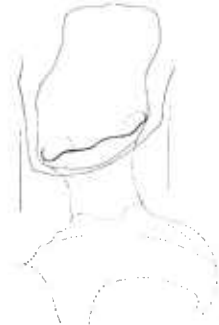
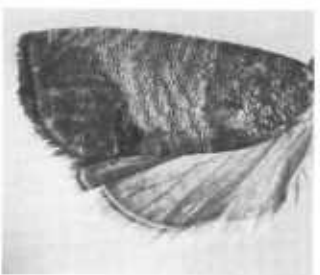
*Cydia gallaesaliciana* (Riley)

Forewing 5.0 mm long, dark areas brownish black. Adult captured July 8. MI. Larva feeds in *Salix* stem gall. Syst and Biol: Heinrich (1926). (1 N, 1 Gm, T, photo specimen Boone Co., MO)



*Cydia toreuta* (Grote) complex. Eastern pine seedworm

Forewing 5.0 to 7.0 mm long, dark markings bordering metallic bands brownish black. Adults captured May 24–July 2. MI, WI, MN. Larva feeds on seeds within cones of *Pinus resinosa*, *P. banksiana*. Syst: Heinrich (1926). Biol: Harbo and Kraft (1969). (84 N, 12 Gm, 7 Gf, T)



*Cydia pomonella* (Linnaeus). Codling moth

Forewing 6.5 to 10.0 mm long, dark apical area yellowish brown. Male hindwing cubital pecten in a pocket. Adults captured April 2–September 12. MI, WI, MN. Bivoltine. Larva feeds on *Pyrus* seeds and fruit. Syst: Brown (1979b). Biol: Putman (1963). (128 N, 13 Gm, 8 Gf)

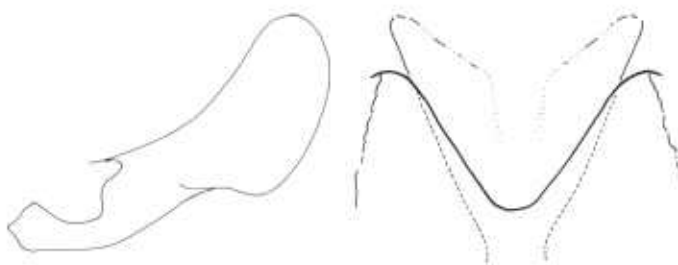
## Genus *Ecdytolopha*

**Both sexes.** Thorax with posterior tuft. Forewing termen convex,  $R_1$  originating at or near middle of discal cell, upper internal vein of discal cell originating between  $R_2$  and  $R_3$ ,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate, cubital vein with pecten,  $M_3$  and  $Cu_1$  connate or stalked.

**Male.** Hindwing inner margin modified into a pocket with or without enclosed hair pencil. Abdominal segment 8 without paired lateral tufts. Uncus not developed; socius and hamus absent; valva without rudimentary clasper, without thick spinelike seta on cucullus at lower margin; vesica with cornuti.

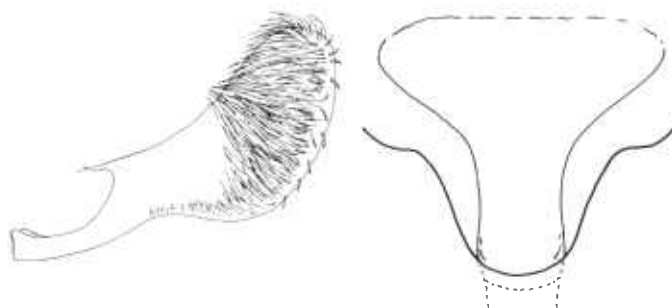
**Female.** Sternum 7 deeply emarginate around ostium bursae, neither inflected nor overlapping ostium bursae. Ductus bursae sclerotized one-third its length from genital opening; corpus bursae with two thornlike signa.

**Comments.** Five Nearctic species of *Ecdytolopha* are known.



*Ecdytolopha punctidiscanum*  
(Dyar)

Forewing 7.5 to 11.5 mm long, dark areas grayish yellowish brown or brownish black. Male with dilated scaling and hair pencil on hindmost tibia. Adults captured June 12–August 17. MI, WI. Larva feeds in *Robinia* stems. Syst: Heinrich (1926). Biol: Prentice (1966). (46 N, 2 Gm, 4 Gf, T)



*Ecdytolopha insiticiana*  
Zeller. Locust twig borer

Forewing 8.0 to 12.0 mm long, dark areas and markings grayish yellowish brown or brownish black. Adults captured May 23–August 12. MI, WI, MN. Larva feeds in *Robinia* rudimentary stem gall. Syst: Heinrich (1926). Biol: Harman and Berisford (1979). (32 N, 5 Gm, 3 Gf, T)

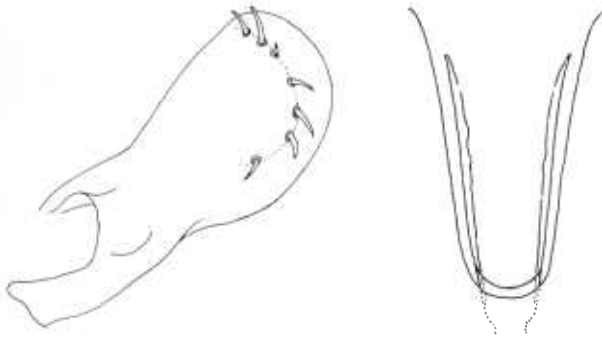
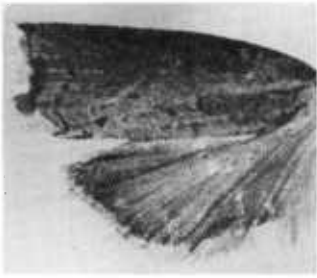
## Genus *Pseudogalleria*

**Both sexes.** Forewing termen concave,  $R_2$  originating equidistant between  $R_1$  and  $R_3$ , upper internal vein of discal cell originating between  $R_2$  and  $R_3$ ,  $R_3$  and  $R_4$  approximate,  $Cu_2$  originating before distal two-thirds of discal cell. Hindwing  $R_s$  and  $M_1$  approximate, cubital vein with pecten,  $M_3$  and  $Cu_1$  stalked.

**Male.** Hindwing inner margin not modified. Hindmost tibia without dilated or tufted scaling. Abdominal segment 8 without paired lateral tufts. Uncus not developed; socius and hamus absent; valva without rudimentary clasper, without thick spinelike seta on cucullus at lower margin; vesica with cornuti.

**Female.** Sternum 7 deeply emarginate around ostium bursae, neither inflected nor overlapping ostium bursae. Ductus bursae with a sclerotized patch near middle; corpus bursae with two thornlike signa.

**Comments.** *Pseudogalleria* appears to be monotypic.



*Pseudogalleria inimicella* (Zeller)

Forewing 7.5 to 11.5 mm long, dark areas reddish brown. Adults captured May 19–June 26. MI, MN. Larva feeds in *Smilax* stem bases. Syst: Heinrich (1923b). Biol: Putman (1942). (12 N, 3 Gm, 5 Gf, T)

## References Cited

- Abrahamson, L.P.; Kraft, K.J. (1965) A population study of the cone moth *Laspeyresia toreuta* Grote in *Pinus banksiana* stands. *Ecology*. 46:561–563; 1965.
- Adamski, D.; Peters, T.M. (1986) Review of nearctic *Apotomis* Hübner (Lepidoptera: Tortricidae: Olethreutini). *Canadian Entomologist*. 118:649–689; 1986.
- Baker, T.C.; Cardé, R.T. (1979) Courtship behavior of the oriental fruit moth (*Grapholitha molesta*): experimental analysis and consideration of the role of sexual selection in the evolution of courtship pheromones in the Lepidoptera. *Annals of the Entomological Society of America*. 72:173–188; 1970.
- Baldur, W.V. (1959) Obligatory and facultative insects in rose hips: their recognition and bionomics. *Illinois Biological Monographs*. 26:1–194; 1959.
- Barras, S.J.; Norris, D.M. (1969) Bionomics of *Eucosma monitorana* (Lepidoptera: Tortricidae) attacking red pine cones in Wisconsin. *Annals of the Entomological Society of America*. 62:1284–1290; 1969.
- Beebe, R. (1954) A new North American record and a second rarity. *The Lepidopterists' News*. 8:26; 1954.
- Benander, P. (1950) Vecklarfjärilar, Tortricina. *Svensk Insektafauna*. 39:1–173; 1950.
- Bennett, S.E. (1961) The strawberry leaf roller complex in Tennessee. *Journal of the Tennessee Academy of Sciences*. 36:320–360; 1961.
- Bentinck, G.A.; Graaf, Diakonoff, A. (1968) De Nederlandse Bladrollers (Tortricidae). *Monografieën van de Nederlandse Entomologische Vereniging*. 3:1–201; 1968.
- Blais, J.R. (1961) Notes on the biology of *Griselda radicana* (Wlsm.) (Lepidoptera: Olethreutidae). *Canadian Entomologist*. 93:648–653; 1961.
- Blanchard, A. (1979) New status for *Epiblema minutana* (Kearfott) and new species of *Epiblema* Hübner and *Sonia* Heinrich (Tortricidae). *Journal of the Lepidopterists' Society*. 33:179–188; 1979.
- Blanchard, A. (1984) *Epiblema luctuosana* A. Blanchard, a homonym, is changed to *Epiblema luctuosissima*, new name. *Journal of the Lepidopterists' Society*. 38:245; 1984.
- Borror, D.J.; DeLong, D.M.; Triplehorn, C.A. (1981) An introduction to the study of insects. 5th ed. New York: Holt, Rinehart and Winston; 1981. 827 p.
- Bottimer, L.J. (1926) Notes on some Lepidoptera from eastern Texas. *Journal of Agricultural Research*. (Washington DC) 33:797–819; 1926.
- Bradley, J.D.; Tremewan, W.G.; Smith, A. (1979) British tortricoid moths. Tortricidae: Olethreutinae. London: The Ray Society; 1979. 336 p.
- Braun, A.F. (1951) The *Aesculus*-feeding species of *Exartema* with description of a new species (Lepidoptera, Eucosmidae). *Ohio Journal of Science*. 51:353–357; 1951.
- Brown, R.L. (1973) Phylogenetic systematics: its application to the genus *Epiblema* (Lepidoptera). Fayetteville, AR: University of Arkansas; 1973. 179 p. M.S. thesis.
- Brown, R. L. (1979a) Nomenclatorial changes in Eucosmini (Tortricidae). *Journal of the Lepidopterists' Society*. 33:21–28; 1979.
- Brown, R.L. (1979b) The valid generic and tribal names for the codling moth, *Cydia pomonella* (Olethreutinae: Tortricidae). *Annals of the Entomological Society of America*. 72:565–567; 1979.
- Brown, R.L. (1980a) A revision of the genus *Epinotia* (Hübner) (Tortricidae: Eucosmini), part 1: The North American species of the *stroemiana* lineage. Ithaca, NY: Cornell University; 1980. 470 p. Ph.D. dissertation. [Abst. Int. 41/9 (B):3302. Microfilm No. 8102899.]
- Brown, R.L. (1980b) A new species of *Epinotia* Hübner (Lepidoptera: Tortricidae). *Proceedings of the Entomological Society of Washington*. 82:504–509; 1980.
- Brown, R.L. (1982) Notes on *Gretchena*: a new species and the synonymy of *Gwendolina* (Lepidoptera: Tortricidae). *Proceedings of the Entomological Society of Washington*. 84:594–602; 1982.
- Brown, R.L. (1983) Taxonomic and morphological investigations of Olethreutinae: *Rhopobota*, *Griselda*, *Melissopus*, and *Cydia* (Lepidoptera: Tortricidae). *Entomography*. 2:97–120; 1983.
- Brown, R.L. (1984) Review of *Corticivora* (Lepidoptera: Tortricidae) with analysis of its tribal relationships and descriptions of new species. *Proceedings of the Entomological Society of Washington*. 86:278–286; 1984.
- Brown, R.L. (1986) Resurrection of *Catastega* Clemens and revision of the *Epinotia vertumnana* (Zeller) species-group (Tortricidae: Olethreutinae). *Journal of the Lepidopterists' Society*. 40:327–346; 1986.
- Brown, R.L.; Miller, P.R. (1983) Studies of Lepidoptera hindwings with emphasis on ultrastructure of scales in *Cydia caryana* (Fitch) (Tortricidae). *Entomography*. 2:261–295; 1983.
- Brown, R.L.; Miller, W.E. (1983) Valid names of the spruce seed moth and a related *Cydia* species (Lepidoptera: Tortricidae). *Annals of the Entomological Society of America*. 76:110–111; 1983.
- Brown, R.L.; Clarke, J.F.G.; Habeck, D.H. (1983) New host records for Olethreutinae (Tortricidae). *Journal of the Lepidopterists' Society*. 37:224–227; 1983.
- Butcher, J.W.; Hodson, A.C. (1949) Biological and ecological studies on some lepidopterous bud and shoot insects of jack pine (Lepidoptera-Olethreutidae). *Canadian Entomologist*. 81:161–173; 1949.
- Čapek, M. (1971) The possibility of biological control of imported weeds of the genus *Solidago* L. in Europe. *Acta Instituti Forestalis Zvolensis*. 429–441; 1971.
- Chapman, P.J.; Lienk, S.E. (1971) Tortricid fauna of apple in New York (Lepidoptera: Tortricidae); including an account of apple's occurrence in the state, especially as a naturalized plant. Spec. Publ. Geneva, NY: New York Agricultural Experiment Station; 1971. 122 p.
- Clarke, J.F.G. (1941) The preparation of slides of the genitalia of Lepidoptera. *Bulletin of the Brooklyn Entomological Society*. 36:149–161; 1941.
- Clarke, J.F.G. (1951) A new genus and species of North American Olethreutidae (Lepidoptera: Laspeyresinae). *Journal of the Washington Academy of Sciences*. 41:46–47; 1951.
- Clarke, J.F.G. (1953) New species of Olethreutidae from Illinois (Lepidoptera). *Journal of the Washington Academy of Sciences*. 43:226–231; 1953.
- Clarke, J.F.G. (1958) Catalogue of the type specimens of Microlepidoptera in the British Museum (Natural History) described by Edward Meyrick. Vol. 3. London: The British Museum (Natural History); 1958. 600 p.

- Clarke, J.F.G. (1973) The genus *Eumarozia* Heinrich (Olethreutidae). *Journal of the Lepidopterists' Society*. 27:268-274; 1973.
- Côté, W.A.; Allen, D.C. (1973) Biology of the maple trumpet skeletonizer, *Epinotia aceriella* (Lepidoptera: Olethreutidae), in New York. *Canadian Entomologist*. 105:463-470; 1973.
- Davault, L.; Ducharme, R. (1966) Life history and habits of the green spruce leaf miner, *Epinotia nanana* (Treitschke) (Lepidoptera: Tortricidae). *Canadian Entomologist*. 98:693-699; 1966.
- DeBoo, R.F.; Sippell, W.L.; Wong, H.R. (1971) The eastern pine-shoot borer, *Eucosma gloriola* (Lepidoptera: Tortricidae), in North America. *Canadian Entomologist*. 103:1473-1486; 1971.
- Decker, G.C. (1932) Biology of the bidens borer, *Epiblema otiosana* (Clemens) (Lepidoptera, Olethreutidae). *Journal of the New York Entomological Society*. 40:503-509; 1932.
- Denmark, H.A. (1960) Some observations on the biology of *Anchylopera platanana* Clemens (Lepidoptera, Olethreutidae) in Florida. *Florida Entomologist*. 43:81-87; 1960.
- Diakonoff, A. (1964) Further records and descriptions of the species of *Bactra* Stephens (Lepidoptera, Tortricidae). *Zoologische Verhandelingen Rijksmuseum van Natuurlijke Historie (Leiden)* 70:1-81; 1964.
- Diakonoff, A. (1973) The South Asiatic Olethreutini (Lepidoptera, Tortricidae). *Zoologische Monographiën Rijksmuseum van Natuurlijke Historie (Leiden)* 1:1-700; 1973.
- Ferguson, D.C. (1975) Host records for Lepidoptera reared in eastern North America. *Tech. Bull.* 1521. Washington, DC: U.S. Department of Agriculture; 1975. 49 p.
- Fernald, M.L. (1970) Gray's manual of botany. 8th ed. New York: D. Van Nostrand; 1970. 1632 p.
- Fernekes, V. (1906) List of Lepidoptera occurring in Milwaukee County. *Bulletin of the Wisconsin Natural History Society*. 4:39-58; 1906.
- Fluke, C.L. (1921) The pea moth in Wisconsin. *Journal of Economic Entomology*. 14:94-98; 1921.
- Forbes, W.T.M. (1923) The Lepidoptera of New York and neighboring states. *Mem.* 68. Ithaca, NY: Cornell University Agricultural Experiment Station; 1923. 729 p.
- Freeman, T.N. (1941) New species of Canadian Lepidoptera. *Canadian Entomologist*. 73:123-127; 1941.
- Freeman, T.N. (1942) A new species of *Pseudexentera* from apple, with notes on allied species (Olethreutidae, Lepidoptera). *Canadian Entomologist*. 74:212-215; 1942.
- Freeman, T.N. (1957) A new species of the genus *Aphania* (Olethreutidae). *The Lepidopterists' News*. 11:27-28; 1957.
- Frick, K.E.; Garcia, C. (1975) *Bactra verutana* as a biological control agent for purple nutsedge. *Annals of the Entomological Society of America*. 68:7-14; 1975.
- Godfrey, G.L.; Cashatt, E.D.; Glenn, M.O. (1987) Microlepidoptera from the Sandy Creek and Illinois River region: an annotated checklist of the suborders Dacnonypha, Monotrysis, and Di-trysia (in part) (Insecta). *Spec. Publ.* 7. Champaign, IL: Illinois Natural History Survey. [In press.]
- Grant, G.G. (1978) Morphology of the presumed male pheromone glands on the forewings of tortricid and phyticid moths. *Annals of the Entomological Society of America*. 71:423-431; 1978.
- Hannemann, H.J. (1961) Die Tierwelt Deutschlands und der angrenzenden Meeresteile 48, Kleinschmetterlinge oder Microlepidoptera I, Die Wickler (s. str.) (Tortricidae). Jena: VEB Gustav Fischer Verlag; 1961. 233 p.
- Harbo, J.F.; Kraft, K.J. (1969) A study of *Phanerotoma toreuta*, a parasite of the pine cone moth *Laspeyresia toreuta*. *Annals of the Entomological Society of America*. 62:214-220; 1969.
- Hare, J.D. (1977) The biology of *Phaneta imbridana* (Lepidoptera: Tortricidae), a seed predator of *Xanthium strumarium* (Compositae). *Psyche*. 84:179-182; 1977.
- Harman, D.M.; Berisford, C.W. (1979) Host relationships and determination of larval instars of the locust twig borer *Ecdytolopha insiticiana*. *Environmental Entomology*. 8:19-23; 1979.
- Heinrich, C. (1923a) New Olethreutidae from eastern United States (Lepidoptera). *Proceedings of the Entomological Society of Washington*. 25:105-122; 1923.
- Heinrich, C. (1923b) Revision of the North American moths of the subfamily Eucosminae of the family Olethreutidae. *United States National Museum Bulletin*. 123:1-298; 1923.
- Heinrich, C. (1924) North American Eucosminae, notes and new species (Lepidoptera). *Journal of the Washington Academy of Sciences*. 14:385-393; 1924.
- Heinrich, C. (1926) Revision of the North American moths of the subfamilies Laspeyresinae and Olethreutinae. *United States National Museum Bulletin*. 132:1-216; 1926.
- Heinrich, C. (1929) Notes on some North American moths of the subfamily Eucosminae. *United States National Museum Proceedings*. 75:1-23; 1929.
- Hetz, M.W.; Werner, F.G. (1979) Insects associated with roots of some rangeland Compositae in southern Arizona. *Southwestern Entomologist*. 4:285-288; 1979.
- Horak, M. (1984) Assessment of taxonomically significant structures in Tortricinae (Lep., Tortricidae). *Mitteilungen der Schweizerischen Entomologischen Gesellschaft*. 57:3-64; 1984.
- Huang, Y.-M. (1962) Wisconsin Laspeyresinae. Madison, WI: University of Wisconsin; 1962. 57 p. M.S. paper.
- Jubb, G.L. (1973) Catches of *Episimus argutus* in grape berry moth sex-pheromone traps in Pennsylvania. *Journal of Economic Entomology*. 66:1345-1346; 1973.
- Kelly, K.L.; Judd, D.B. (1955) The ISCC-NBS method of designating colors and a dictionary of color names. *NBS Circ.* 553. Washington, DC: U.S. Department of Commerce, National Bureau of Standards; 1955. 158 p.
- Klots, A.B. (1942) Type material of North American Microlepidoptera other than Aegeriidae in the American Museum of Natural History. *Bulletin of the American Museum of Natural History*. 79:391-424; 1942.
- Klots, A.B. (1970) Lepidoptera. In: Tuxen, S.L., ed. *Taxonomist's glossary of genitalia in insects*. 2d ed. Copenhagen: Munksgaard; 1970. 115-130.
- Krauth, S.J.; Hall, D.J.; Coppel, H.C.; Shenefelt, R.D. (1977) New Wisconsin records of Lepidoptera reared in conjunction with the gypsy moth (*Lymantria dispar* (L.)) parasitoid recovery program. *For. Res. Notes* 208. Madison, WI: University of Wisconsin; 1977. 5 p.
- Küchler, A.W. (1964) Potential natural vegetation of the conterminous United States. *Spec. Publ.* 36. New York: American Geographical Society; 1964. 39 p.
- Kusch, D.S. (1967) Notes on the biology of *Epinotia criddleana* Kft. *Bi-Monthly Research Notes, Canadian Forestry Service*. 23(1):3; 1967.
- Kuznetsov, V.I. (1978) Taxonomic key to insects of the European USSR, 4, Lepidoptera, 21, Tortricidae [Russian]. Leningrad: "NAUKA"; 1978. 193-680.
- Kuznetsov, V.I. (1986) Type specimens of super tribe Eucosmidii (Lepidoptera, Tortricidae) from the collection of F. Treitschke in the Hungarian Natural History Museum (Budapest). *Entomological Review* [a translation of *Entomologicheskoye Obzreniye*]. 65:153-167; 1986.



- Kuznetsov, V.I.; Stekolnikov, A.A. (1977) Functional morphology of the male genitalia and phylogenetic relationships of some tribes in the family Tortricidae (Lepidoptera) of the fauna of the Far East. Transactions of the Zoological Institute of the Academy of Science USSR [Russian]. 70:65–97; 1977.
- Leraut, P. (1978) Quelques changements dans le nomenclature des Tortricidae de France. Alexanor. 10:338–341; 1978.
- Lindquist, O.H. (1961) A shoot moth on jack pine in Ontario. Bi-Monthly Progress Report, Canadian Forestry Service. 17(2):2; 1961.
- Lindquist, O.H.; Macleod, L.S. (1967) A biological study of *Epinotia solandriana* (Lepidoptera: Olethreutidae), a leaf roller on birch in Ontario. Canadian Entomologist. 99:1110–1114; 1967.
- MacKay, M.R. (1953) The last-instar larva of *Epinotia medioviridana* (Kft.) (Lepidoptera: Olethreutidae). Canadian Entomologist. 85:404–407; 1953.
- MacKay, M.R. (1959) Larvae of the North American Olethreutidae (Lepidoptera). Canadian Entomologist. 91(Suppl. 10):1–338; 1959.
- MacKay, M.R. (1962) Additional larvae of the North American Olethreutinae (1) (Lepidoptera: Tortricidae). Canadian Entomologist. 94:626–643; 1962.
- Marshall, G.E.; Musgrave, L.I. (1937) A progress report on the Microlepidoptera of southern Indiana, and their parasites. Canadian Entomologist. 69:100–106; 1937.
- Martin, J.L. (1960) Life history of the pine tip moth, *Rhyacionia adana* Heinrich, in Ontario (Lepidoptera: Olethreutidae). Canadian Entomologist. 92:724–728; 1960.
- McDowell, L.L.; Wong, H.R. (1962) Pine tip moths in southeastern Manitoba. Bi-Monthly Progress Report, Canadian Forestry Service. 18(4):2; 1962.
- McDunnough, J. (1925a) New Canadian Lepidoptera with notes. Canadian Entomologist. 57:11–23; 1925.
- McDunnough, J.H. (1925b) New Canadian Eucosminae (Lepidoptera). Canadian Entomologist. 57:115–116; 1925.
- McDunnough, J. (1929) Some apparently new Microlepidoptera. Canadian Entomologist. 61:266–271; 1929.
- McDunnough, J. (1931) A new *Argyroplote* species (Eucosmidae, Lepid.). Canadian Entomologist. 63:150–152; 1931.
- McDunnough, J. (1933) Notes on the biology of certain tortricid species with structural details of the larvae and pupae. Canadian Journal of Research. 9:502–517; 1933.
- McDunnough, J. (1935) New Canadian eucosmids with notes (Lepidoptera). Canadian Entomologist. 67:140–149; 1935.
- McDunnough, J. (1938) Some apparently new Eucosmidae (Lepid.). Canadian Entomologist. 70:90–100; 1938.
- McDunnough, J. (1942) Tortricid notes and descriptions. Canadian Entomologist. 74:63–71; 1942.
- McDunnough, J. (1944) New Microlepidoptera with notes. Canadian Entomologist. 76:153–156; 1944.
- McDunnough, J.H. (1954) New Microlepidoptera from the region of Halifax, Nova Scotia, with notes on other species. Am. Mus. Novit. 1686. New York: American Museum of Natural History; 1954. 15 p.
- McDunnough, J. (1955) Critical remarks on the synonymy of certain *Anchylopera* species, with descriptions of new species (Lepidoptera, Eucosmidae). Am. Mus. Novit. 1725. New York: American Museum of Natural History; 1955. 16 p.
- McDunnough, J.H. (1956) Microlepidoptera notes and new species. Am. Mus. Novit. 1789. New York: American Museum of Natural History; 1956. 17 p.
- McDunnough, J.H. (1959) On some changes in nomenclature of Microlepidoptera, with description of a new species. Am. Mus. Novit. 1954. New York: American Museum of Natural History; 1959. 9 p.
- McGregor, M.D. (1967) Biology and natural enemies of an aspen leaf tier, *Sciaphila duplex*, in the intermountain region. Journal of Economic Entomology. 60:1213–1216; 1967.
- Merz, R.W. (1979) Forest atlas of the midwest. St. Paul, MN: U.S. Department of Agriculture, Forest Service, North Central Forest Experiment Station; 1979. 48 p.
- Miller, W.E. (1967) The European pine shoot moth—ecology and control in the Lake States. Forest Science Monographs. 14:1–72; 1967.
- Miller, W.E. (1970) Fernald types of North American Olethreutinae (Lepidoptera: Tortricidae). Proceedings of the Entomological Society of Washington. 72:288–294; 1970.
- Miller, W.E. (1971) Identity of *Phaneta refusana* (Walker) with description of a new species (Tortricidae). Journal of the Lepidopterists' Society. 25:284–287; 1971.
- Miller, W.E. (1973a) Clemens types of Olethreutinae (Lepidoptera, Tortricidae). Transactions of the American Entomological Society. 99:205–234; 1973.
- Miller, W.E. (1973b) Two previously unrecognized scientific names for the strawberry leafroller (Lepidoptera: Tortricidae). Annals of the Entomological Society of America. 66:553–554; 1973.
- Miller, W.E. (1974) Identities of taxonomically confused moths of the *Eucosma agricola* group and description of a new species (Lepidoptera, Tortricidae). Annals of the Entomological Society of America. 67:601–604; 1974.
- Miller, W.E. (1976a) Biology and taxonomy of three gall forming species of *Epiblema* (Olethreutidae). Journal of the Lepidopterists' Society. 30:50–58; 1976.
- Miller, W.E. (1976b) A new species of *Laspeyresia* from Michigan (Lepidoptera: Olethreutidae). Great Lakes Entomologist. 9:171–172; 1976.
- Miller, W.E. (1977a) Uniform genitalia among wing color morphs of olethreutid moths. Journal of the Lepidopterists' Society. 31:118; 1977.
- Miller, W.E. (1977b) Wing measure as a size index in Lepidoptera: the family Olethreutidae. Annals of the Entomological Society of America. 70:253–256; 1977.
- Miller, W.E. (1978a) *Larisa subsolana*, a new genus and species of moth from eastern North America (Olethreutidae). Journal of the Lepidopterists' Society. 32:256–260; 1978.
- Miller, W.E. (1978b) *Petrova* pitch-blister moths of North America and Europe: two new species and synopsis (Olethreutidae). Annals of the Entomological Society of America. 71:329–340; 1978.
- Miller, W.E. (1979a) Identity corrections for two North American *Apotomis* moths (Tortricidae: Olethreutinae). Great Lakes Entomologist. 12:115–118; 1979.
- Miller, W.E. (1979b) The genus *Olethreutes*: identity corrections and description of a new species (Lepidoptera: Tortricidae: Olethreutinae). Annals of the Entomological Society of America. 72:232–236; 1979.
- Miller, W.E. (1982) *Grapholita delineana* (Walker), a Eurasian hemp moth, discovered in North America. Annals of the Entomological Society of America. 75:184–186; 1982.
- Miller, W.E. (1983a) Genus *Phaneta*: new synonymies and a new species (Lepidoptera: Tortricidae). Annals of the Entomological Society of America. 76:98–303; 1983.
- Miller, W.E. (1983b) Nearctic *Endothenia* species: a new synonymy, a misidentification, and a revised status (Lepidoptera: Tortricidae). Great Lakes Entomologist. 16:5–12; 1983.
- Miller, W. E. (1983c) *Eucosmomorpha albersana* (Hübner), a palaearctic species, collected in North America (Tortricidae, Grapholitini). Journal of the Lepidopterists' Society. 37:88–89; 1983.

- Miller, W.E. (1983d) New synonymies in nearctic *Dichrorampha* (Lepidoptera: Tortricidae). Proceedings of the Entomological Society of Washington. 85:727-733; 1983.
- Miller, W.E. (1985a) Nearctic *Epiblema*: a new synonymy, a revised identity, and two new species (Lepidoptera: Tortricidae). Great Lakes Entomologist. 18:33-38; 1985.
- Miller, W.E. (1985b) Nearctic *Rhyacionia* pine tip moths: a revised identity and a new species (Lepidoptera: Tortricidae). Great Lakes Entomologist. 18:119-122; 1985.
- Miller, W.E. (1985c) Nearctic *Eucosma* (Lepidoptera: Tortricidae): four new species and three new synonymies. Annals of the Entomological Society of America. 78:240-247; 1985.
- Miller, W.E. (1985d) Nearctic *Olethreutes*: five new synonymies, two revised statuses, and notes (Lepidoptera: Tortricidae). Proceedings of the Entomological Society of Washington. 87:408-417; 1985.
- Miller, W.E. (1985e) *Pammene perstructana* (Walker) (Lepidoptera: Tortricidae) identified after more than a century. Great Lakes Entomologist. 18:145-147; 1985.
- Miller, W.E. (1986a) *Epinotia nisella* (Clerck): an unrecorded host and mode of feeding (Lepidoptera: Tortricidae). Great Lakes Entomologist. 19:205-207; 1986.
- Miller, W.E. (1986b) The species of *Pseudexentera* (Tortricidae). Journal of the Lepidopterists' Society. 40:218-237; 1986.
- Miller, W.E.; Pogue, M.G. (1984) Ragweed borer (Lepidoptera: Tortricidae: Eucosmini): taxonomic implications of an allometric analysis of adult characters. Annals of the Entomological Society of America. 77:227-231; 1984.
- Morris, R.C. (1967) Biology of *Gypsonoma haimbachiana* (Lepidoptera: Olethreutidae), a twig borer in eastern cottonwood. Annals of the Entomological Society of America. 60:423-427; 1967.
- Mosher, D.G.; Wilson, L.F. (1974) Life history and some habits of a larch moth, *Paralobesia palliolana* (Lepidoptera: Tortricidae), in Michigan. Great Lakes Entomologist. 7:133-136; 1974.
- Moznette, G.F.; Bissell, T.L.; Adair, H.S. (1931) Insects of the pecan and how to combat them. Farm. Bull. 1654. Washington, DC: U.S. Department of Agriculture; 1931. 59 p.
- Mutuura, A.; Freeman, T.N. (1966) The North American species of the genus *Zeiraphera* Treitschke (Olethreutidae). Journal of Research on the Lepidoptera. 5:153-176; 1966.
- Neiswander, R.B. (1944) Insect pests of strawberries in Ohio. Bull. 651. Wooster, OH: Ohio Agricultural Experiment Station; 1944. 37 p.
- Oatman, E.R.; Legner, E.F.; Brooks, R.F. (1962) Bionomics of the eye-spotted bud moth, *Spilonota ocellana*, on cherry in Wisconsin. Journal of Economic Entomology. 55:930-934; 1962.
- Obraztsov, N.S. (1953) Classification of holartic species of the genus *Lobesia* Guenée, with description of *Paralobesia* gen. nov. (Lepidoptera, Tortricidae). Tijdschrift voor Entomologie. 96:85-94; 1953.
- Obraztsov, N.S. (1958) Die Gattungen der palaearktischen Tortricidae. II. Die Unterfamilie Olethreutinae. Tijdschrift voor Entomologie. 101:229-261; 1958.
- Obraztsov, N.S. (1961) Die Gattungen der palaearktischen Tortricidae. II. Die Unterfamilie Olethreutinae. Teil 4. Tijdschrift voor Entomologie. 104:51-70; 1961.
- Pankhurst, R.J. (1970) Key generation by computer. Nature. 227:1269-1270; 1970.
- Payne, J.A.; Heaton, E.K. (1975) The hickory shuckworm: its biology, effect upon nut quality, and control. Annual Report of the Northern Nut Growers Association. 66: 19-25; 1975.
- Peterson, L.O.T. (1958) The boxelder twig borer, *Proteoteras willingana* (Kearfott), (Lepidoptera: Olethreutidae). Canadian Entomologist. 90:639-646; 1958.
- Pettit, R.H. (1933) The principal grape insects of Michigan. Spec. Bull. 239. East Lansing, MI: Michigan Agricultural Experiment Station; 1933. 18 p.
- Pilon, J.G. (1965) Bionomics of the spruce budworm, *Zeiraphera ratzeburgiana* (Ratz.) (Lepidoptera: Olethreutidae). Phytoprotection. 46:5-13; 1965.
- Plank, H.K. (1922) The blackhead fireworm of cranberry on the Pacific coast. Bull. 1032. Washington, DC: U.S. Department of Agriculture; 1922. 46 p.
- Powell, J.A. (1962) Biological and taxonomic notes on two California species of *Proteoteras* (Lepidoptera: Tortricidae). Pan-Pacific Entomologist. 38:191-195; 1962.
- Powell, J.A. (1968) Host associations and taxonomy of nearctic conifer cone moths in the genus *Eucosma* (Lepidoptera: Tortricidae). Hilgardia. 39:1-36; 1968.
- Powell, J.A. (1980) Evolution of larval food preferences in Microlepidoptera. Annual Review of Entomology. 25:133-159; 1980.
- Powell, J.A. (1983) Tortricidae. In: Hodges, R.W., ed. Check list of the Lepidoptera of America north of Mexico. London: E.W. Classey and the Wedge Entomological Research Foundation; 1983: 31-41.
- Powell, J.A.; Miller, W.E. (1978) Nearctic pine tip moths of the genus *Rhyacionia*: biosystematic review (Lepidoptera: Tortricidae, Olethreutinae). Agric. Handb. 514. Washington, DC: U.S. Department of Agriculture; 1978. 51 p.
- Prentice, R.M., ed. (1966) Microlepidoptera. In: Forest Lepidoptera of Canada recorded by the Forest Insect Survey. Dept. For. Canada Publ. 1142. Ottawa: Department of Forestry, Canada; 1966: 543-840.
- Putman, W.L. (1935) Notes on the hosts and parasites of some lepidopterous larvae. Canadian Entomologist. 67:105-109; 1935.
- Putman, W.L. (1942) Host plants and parasites of some lepidopterous larvae. Canadian Entomologist. 74:219-224; 1942.
- Putman, W.L. (1963) The codling moth, *Carpocapsa pomonella* (L.) (Lepidoptera: Tortricidae): a review with special reference to Ontario. Proceedings of the Entomological Society of Ontario. 93:22-60; 1963.
- Razowski, J. (1976) Phylogeny and system of Tortricidae (Lepidoptera). Acta Zoologica Cracoviensia. 21:73-120; 1976.
- Roberts, D.W. (1966) Life history and parasites of *Evora hemidesma* (Zeller) (Lepidoptera: Olethreutidae). Boyce Thompson Institute Contributions. 23:165-170; 1966.
- Robinson, G.S.; Nielsen, E.S. (1983) The Microlepidoptera described by Linnaeus and Clerck. Systematic Entomology. 8:191-242; 1983.
- Rogers, C.E.; Thompson, T.E.; Jones, O.R. (1979) *Eucosma womonana* Kearfott (Lepidoptera: Olethreutidae): a new pest of sunflower in the southern plains. Journal of the Kansas Entomological Society. 52:373-376; 1979.
- Satterthwait, A.F. (1948) Important sunflower insects and their insect enemies. Journal of Economic Entomology. 41:725-731; 1948.
- Schaffner, J.V. (1959) Microlepidoptera and their parasites reared from field collections in the northeastern United States. Misc. Publ. 767. Washington, DC: U.S. Department of Agriculture; 1959. 97 p.
- Smith, C.C. (1946) Notes on the birch shoot borer, *Epinotia solicitana* Walker (Olethreutidae, Lepidoptera). Acadian Naturalist. 2:114-121; 1946.
- Strickler, K.; Whalon, M. (1985) Microlepidoptera species composition in Michigan apple orchards. Environmental Entomology. 14:486-495; 1985.

- Swatschek, B. (1958) Die Larvalsystematik der Wickler (Tortricidae und Carposinidae). Berlin: Akademie-Verlag; 1958. 269 p.
- Tashiro, H. (1974) Biology and control of the spruce needleminer. *Journal of Economic Entomology*. 67:89–92; 1974.
- Thompson, R.W. (1928) The golden-glow borer (*Epiblema carolinana* Walsingham). Annual Report of the Entomological Society of Ontario. (1927) 58:73–75; 1928.
- Tripp, H.A. (1954) Description and habits of the spruce seed-worm (*Laspeyresia youngana* (Kft.) (Lepidoptera: Olethreutidae)). *Canadian Entomologist*. 86:385–402; 1954.
- Turnock, W.J. (1953) Some aspects of the life history and ecology of the pitch nodule maker, *Petrova albicapitana* (Busck) (Lepidoptera: Olethreutidae). *Canadian Entomologist*. 85:233–243; 1953.
- Vincent, C.; Simard, L.G.; Paradis, R.O. (1985) *Olethreutes olivaceana* (Fern.) (Lepidoptera: Tortricidae), ravageur des fraisières dans le sud-ouest de Québec. *Revue d'Entomologie du Québec*. 30:28–34; 1985.
- Walker, F.H. (1936) Observations on sunflower insects in Kansas. *Journal of the Kansas Entomological Society*. 9:16–25; 1936.
- Wehrle, L.P. (1924) The clover-seed caterpillar. Bull. 428. Ithaca, NY: New York (Cornell) Agricultural Experiment Station; 1924. 34 p.
- Wehrle, L.P. (1929) The clover-leaf caterpillar (*Olethreutes cespitana* Hübner) and the clover-leaf tyer (*Anchylopera angulifasciana* Zeller). Bull. 489. Ithaca, NY: New York (Cornell) Agricultural Experiment Station; 1929. 27 p.
- Werner, F.G. (1982) Common names of insects and related organisms 1982. College Park, MD: Entomological Society of America; 1982. 132 p.
- Wheeler, A.G.; Hoebeke, E.R. (1985) The insect fauna of ninebark, *Physocarpus opulifolius* (Rosaceae). *Proceedings of the Entomological Society of Washington*. 87:356–370; 1985.
- Wong, H.R.; Drouin, J.A.; Szlabey, D.L.; Dang, P.T. (1983) Identification of three species of *Proteoteras* (Lepidoptera: Tortricidae) attacking shoots of Manitoba maple in the Canadian prairies. *Canadian Entomologist*. 115:333–339; 1983.
- Wong, H.R.; Melvin, J.C.E. (1967) The leaf roller *Pseudexentera oregonana* Wlsh. *Bi-Monthly Research Notes, Canadian Forestry Service*. 23(1):3–4; 1967.

## Species Index

- abruptana*, *Epiblema*, 56  
*aceriella*, *Catastega*, 78  
*adana*, *Rhyacionia*, 38  
*adjuncta*, *Gypsonoma*, 63  
*aemulana*, *Endopiza*, 16  
*aesculana*, *Proteoteras*, 64  
*affiliana*, *Endothenia*, 18  
*agilana*, *Olethreutes*, 33  
*agricolana*, *Eucosma*, 49  
*albacostana*, *Ancylis*, 82  
*albersana*, *Eucosmomorpha*, 86  
*albicapitana*, *Retinia*, 39  
*albiciliana*, *Olethreutes*, 33  
*albiguttana*, *Eucosma*, 50  
*albimaculana*, *Cydia*, 92  
*albolineana*, *Endothenia*, 19  
*amatana*, *Gretchena*, 71  
*ambodaidaleia*, *Phaneta*, 47  
*angleseana*, *Grapholita*, 89  
*apicana*, *Ancylis*, 81  
*appendicea*, *Olethreutes*, 31  
*approximana*, *Aterpia*, 19  
*argenticostana*, *Phaneta*, 46  
*argutanus*, *Episimus*, 14  
*astrologana*, *Olethreutes*, 34  
*atrodentana*, *Olethreutes*, 26  
*auricapitana*, *Olethreutes*, 33  
*autumnana*, *Phaneta*, 43  
*awemeana*, *Phaneta*, 42  
  
*barbara*, *Eucosma*, 49  
*bilineana*, *Eucosma*, 52  
*bipartitana*, *Olethreutes*, 25  
*bipunctella*, *Eucosma*, 52  
*bittana*, *Dichorampha*, 84  
*bolliana*, *Gretchena*, 71  
*boxcana*, *Epiblema*, 56  
*brightonana*, *Epiblema*, 59  
*buoliana*, *Rhyacionia*, 38  
*burgessiana* complex, *Ancylis*, 79, 80  
*busckana*, *Rhyacionia*, 38  
  
*canadana*, *Sonia*, 62  
*canadensis*, *Zeiraphera*, 66  
*canana*, *Eucosma*, 48  
*candana*, *Cydia*, 93  
*capreana*, *Apotomis*, 21  
*carbonana*, *Ancylis*, 81  
*carduana*, *Lobesia*, 17  
*carolana*, *Olethreutes*, 34  
*carolinana*, *Epiblema*, 58  
*caryana*, *Cydia*, 94  
*cataclystiana*, *Eucosma*, 54  
*cespitana*, *Olethreutes*, 25, 35  
*chionosema*, *Hedya*, 36  
*cinerodorsana*, *Suleima*, 61  
*clarki*, *Corticivora*, 91  
*clavana*, *Olethreutes*, 25, 28  
*clavana*, *Phaneta*, 46  
*comandrana*, *Olethreutes*, 26  
*comptana*, *Ancylis*, 79, 81  
  
*concinna*, *Olethreutes*, 32  
*concubitana*, *Gretchena*, 71  
*confixana*, *Phaenasiophora*, 24  
*connectus*, *Olethreutes*, 27  
*consobrinana*, *Eucosma*, 54  
*convergana*, *Phaneta*, 45  
*corosana*, *Pelochrista*, 55  
*coruscana*, *Olethreutes*, 33  
*corylana*, *Epinotia*, 75  
*costomaculana*, *Pseudexentera*, 70  
*crescentana*, *Proteoteras*, 64  
*cressoniana*, *Pseudexentera*, 67  
*criddleana*, *Epinotia*, 75  
*cruciana*, *Epinotia*, 73, 77  
*culminana*, *Notocelia*, 61  
*cyanana*, *Hedya*, 37  
  
*deceptana*, *Apotomis*, 21  
*delicatana*, *Gretchena*, 71  
*delineana*, *Grapholita*, 88, 90  
*deludana*, *Gretchena*, 70  
*derelecta*, *Eucosma*, 53  
*desertana*, *Epiblema*, 58  
*dietziana*, *Rhopobota*, 72  
*diminutana*, *Ancylis*, 79, 82  
*divisana*, *Ancylis*, 81  
*dorsiatomana*, *Phaneta*, 46  
*dorsisignatana*, *Eucosma*, 53  
*dorsisuffusana*, *Epiblema*, 60  
*duplex*, *Pseudosciaphila*, 23  
  
*eclipsana*, *Grapholita*, 90  
*electrofusus*, *Olethreutes*, 26  
*essexana*, *Phaneta*, 42  
*exaeresima*, *Olethreutes*, 32  
*exoletus*, *Olethreutes*, 25  
  
*fagigemmeana*, *Olethreutes*, 29  
*fana*, *Grapholita*, 89  
*faracana*, *Pseudexentera*, 67  
*fasciatana*, *Olethreutes*, 32  
*fasciolana*, *Gypsonoma*, 63  
*felicetana*, *Pammene*, 85  
*ferrolineana*, *Olethreutes*, 34  
*finitimana*, *Rhopobota*, 72  
*flexiloqua*, *Cydia*, 93  
*footiana*, *Olethreutes*, 26  
*formosana*, *Phaneta*, 42, 43  
*fortunana*, *Zeiraphera*, 66  
*fulminana*, *Eucosma*, 53  
*funerea*, *Apotomis*, 21  
*furfurana*, *Bactra*, 15  
  
*galaxana*, *Olethreutes*, 33  
*galeamatana*, *Ancylis*, 79  
*galevora*, *Olethreutes*, 30  
*gallaesaliciana*, *Cydia*, 94  
*garacana*, *Cydia*, 92  
*gemistrigulana*, *Retinia*, 40  
*gentianaeana*, *Endothenia*, 17  
*giganteana*, *Eucosma*, 52  
  
*glaciana*, *Olethreutes*, 35  
*gloriola*, *Eucosma*, 51  
*goodelliana*, *Ancylis*, 79  
*graciliana*, *Eucosma*, 50  
*granti*, *Rhyacionia*, 38  
  
*haimbachiana*, *Gypsonoma*, 63  
*haracana*, *Pseudexentera*, 68  
*heathiana*, *Eucosma*, 48  
*hebesana*, *Endothenia*, 17, 18  
*helianthana*, *Suleima*, 61  
*hemidesma*, *Evora*, 37  
*huroniensis*, *Epinotia*, 76  
  
*illotana*, *Notocelia*, 60  
*impudens*, *Endothenia*, 19  
*incanana*, *Dichrorampha*, 84  
*infelix*, *Epiblema*, 58  
*infida*, *Apotomis*, 21, 22  
*inimicella*, *Pseudogalleria*, 96  
*inopiosa*, *Cydia*, 92  
*inornatana*, *Olethreutes*, 27  
*insiticiana*, *Ecdytolopha*, 95  
*interruptolineana*, *Zomaria*, 20  
*interstinctana*, *Grapholita*, 90  
*iowana*, *Epiblema*, 58  
  
*kalmiana*, *Pseudexentera*, 69  
*kiskana*, *Phaneta*, 47  
  
*laciniana*, *Ancylis*, 79  
*lacustrina*, *Cydia*, 93  
*laracana*, *Epinotia*, 74  
*laricana*, *Cydia*, 91  
*latiferraeana* complex, *Cydia*, 94  
*latiferraeana*, *Cydia*, 91  
*lautana*, *Serda*, 87  
*libertana*, *Grapholita*, 88  
*lindana*, *Epinotia*, 77  
*luctuosissima*, *Epiblema*, 57  
  
*maculatana*, *Eucosma*, 50  
*madderana*, *Epinotia*, 74  
*mafica*, *Retinia*, 40  
*maiorina*, *Bactra*, 15  
*major*, *Olethreutes*, 34  
*malachitana*, *Eumaroza*, 20  
*malana*, *Olethreutes*, 31  
*mali*, *Pseudexentera*, 68  
*mappana*, *Barbara*, 41  
*maracana*, *Pseudexentera*, 69  
*marmontana*, *Phaneta*, 44  
*matutina*, *Eucosma*, 51  
*mediofasciana*, *Ancylis*, 82  
*medioplagata*, *Epinotia*, 77  
*medioviridana*, *Epinotia*, 73  
*melanomesa*, *Olethreutes*, 25, 30  
*merrickana*, *Olethreutes*, 29  
*metallica*, *Retinia*, 39  
*metallcana*, *Olethreutes*, 25, 34  
*metamelana*, *Ancylis*, 80

- minutana*, *Epiblema*, 56  
*modernana*, *Phaneta*, 45  
*moftatiana*, *Proteoteras*, 65  
*molesta*, *Grapholita*, 88  
*momonana*, *Epinotia*, 76  
*monitorana*, *Eucosma*, 51  
*montanana*, *Endothenia*, 18  
*montanana*, *Phaneta*, 47  
*morrisoni*, *Eucosma*, 49  
*multilineana*, *Cydia*, 92  
*muricana*, *Ancylis*, 81  
*murina*, *Olethreutes*, 34  
*mysteriana*, *Olethreutes*, 27
- naevana*, *Rhopobota*, 72  
*nanana*, *Epinotia*, 73, 76  
*nandana*, *Eucosma*, 52  
*naracana*, *Proteoteras*, 65  
*nigrana*, *Olethreutes*, 25, 28  
*nigricana*, *Cydia*, 91  
*nisella*, *Epinotia*, 73, 75  
*niveiguttana*, *Phaenocarpa*, 24  
*nonana*, *Epinotia*, 76  
*nubeculana*, *Ancylis*, 79  
*nubiferana*, *Hedya*, 36  
*nubilana*, *Endothenia*, 18  
*numerosana*, *Epiblema*, 56
- obfuscana*, *Epiblema*, 57  
*ocellana*, *Spilonota*, 41  
*ochrocephala*, *Phaneta*, 47  
*ochroleucana*, *Hedya*, 36  
*ochrosuffusana*, *Olethreutes*, 29  
*ochroterminana*, *Phaneta*, 44  
*olivaceana*, *Olethreutes*, 26  
*olivaceana*, *Phaneta*, 47  
*oregonana*, *Pseudexentera*, 69  
*ornatula*, *Phaneta*, 42, 45  
*otiosana*, *Epiblema*, 59
- packardi* complex, *Grapholita*, 89  
*packardi*, *Grapholita*, 88  
*palabundana*, *Eucosma*, 51  
*pallidicostana*, *Phaneta*, 46  
*palliola*, *Endopiza*, 16  
*pallipennis*, *Retinia*, 40  
*paraplesiana*, *Sonia*, 62  
*parmatana*, *Phaneta*, 44  
*permundana*, *Olethreutes*, 25, 31  
*perstructana*, *Pammene*, 86  
*platanana*, *Ancylis*, 80  
*plummeriana*, *Talponia*, 85  
*pomonella*, *Cydia*, 91, 94  
*populana*, *Cydia*, 92  
*prunivora*, *Grapholita*, 89  
*punctana*, *Olethreutes*, 27  
*punctidiscanum*, *Ecdytolopha*, 95
- quadrifidus*, *Olethreutes*, 27
- radiatana*, *Phaneta*, 42  
*radicana*, *Epinotia*, 74  
*raracana*, *Phaneta*, 43  
*removana*, *Apotomis*, 22  
*resumptana*, *Epiblema*, 59  
*ridingsana*, *Eucosma*, 48  
*rindgei*, *Eucosma*, 50
- robinsonana*, *Eucosma*, 48  
*rusticella*, *Cydia*, 91, 93
- salicicolana*, *Gypsonoma*, 63  
*scintillana*, *Pelochrista*, 55  
*scudderiana*, *Epiblema*, 57  
*sedatana*, *Dichrorampha*, 83, 84  
*semiovana*, *Ancylis*, 80  
*separatana*, *Hedya*, 36  
*sepia*, *Pseudexentera*, 67  
*septemberana*, *Epinotia*, 77  
*sericorana*, *Olethreutes*, 30  
*sheppardana*, *Ancylis*, 79  
*similiana*, *Eucosma*, 53  
*simulana*, *Dichrorampha*, 84  
*smithiana*, *Eucosma*, 49  
*solandriana*, *Epinotia*, 73  
*solicitana*, *Epinotia*, 75  
*sombreana*, *Eucosma*, 54  
*sonia*, *Rhyacionia*, 39  
*spiraeifolia*, *Ancylis*, 79  
*spiraeifolia*, *Endopiza*, 16  
*spoliata*, *Pseudexentera*, 68  
*strenuana* complex, *Epiblema*, 56  
*strenuana*, *Epiblema*, 56  
*striatana*, *Phaneta*, 46  
*strobilella*, *Cydia*, 91  
*subaequana* complex, *Ancylis*, 79  
*submissana*, *Olethreutes*, 31  
*subsolana*, *Larisa*, 87  
*substitutionis*, *Gypsonoma*, 63
- tandana*, *Epiblema*, 56, 59  
*tarandana*, *Phaneta*, 45  
*tautana*, *Serda*, 87, 88  
*tiliana*, *Olethreutes*, 25, 28  
*timidella*, *Catantopha*, 78  
*tineana*, *Ancylis*, 79, 82  
*tocullionana*, *Eucosma*, 51  
*tomonana*, *Phaneta*, 44  
*toreuta* complex, *Cydia*, 94  
*toreuta*, *Cydia*, 91  
*transmissana*, *Epinotia*, 76  
*trinitana*, *Olethreutes*, 35  
*tripartitana*, *Epiblema*, 57  
*tristrigana*, *Grapholita*, 90  
*trogodana*, *Olethreutes*, 32
- umbrastriana*, *Phaneta*, 42, 43  
*undulana*, *Orthotaenia*, 23  
*unfortunata*, *Zeiraphera*, 66  
*unguicella*, *Ancylis*, 79, 82
- vaccinii*, *Pseudexentera*, 69  
*vagana*, *Eucosma*, 50  
*valdana*, *Olethreutes*, 30  
*verna*, *Phaneta*, 43  
*vernalana*, *Phaneta*, 45  
*versicolorana*, *Olethreutes*, 30  
*vertumnana*, *Epinotia*, 75  
*verutana*, *Bactra*, 15  
*vestaliana*, *Hystricophora*, 83  
*viburnana*, *Olethreutes*, 25, 28  
*virginiana*, *Pseudexentera*, 70  
*viteana*, *Endopiza*, 16
- walsinghami*, *Epiblema*, 58  
*watchungana*, *Gretchena*, 71  
*willingana*, *Proteoteras*, 64  
*womonana*, *Pelochrista*, 55
- zandana*, *Epinotia*, 74  
*zomonana*, *Pelochrista*, 55

## Host Index

- Abies*, 51, 66  
*A. balsamea*, 41, 66, 74  
*Acer*, 28, 35, 63, 65, 78, 93  
*A. negundo*, 64  
*A. rubrum*, 65  
*A. saccharinum*, 65  
*A. saccharum*, 65  
*Aesculus*, 29, 64  
*Alnus*, 31, 33, 71, 73  
*Ambrosia*, 56  
*Amelanchier*, 31, 36, 79  
*Amorpha*, 90  
*Anaphalis margaritacea*, 59  
*Artemisia*, 46, 47, 49  
*Asimina*, 85  
*Aster*, 42, 44
- Baptisia*, 90  
*Betula*, 21, 23, 33, 35, 73, 78, 81  
*B. papyrifera*, 75  
*B. populifolia*, 75  
*Bidens*, 59
- Cannabis sativa*, 90  
*Carya*, 28, 29, 67, 71, 87, 94  
*C. illinoensis*, 71, 94  
*Castanea*, 67  
*Ceanothus*, 80  
*Celtis*, 27, 74  
*Chrysanthemum*, 49, 55  
*Cirsium*, 17, 37  
*Comandra*, 26  
*Comptonia*, 26  
*Coreopsis*, 61  
*Cornus*, 27, 30, 32, 77, 81  
*Corylus*, 21, 28, 29, 75, 80  
*Crataegus*, 36, 68, 69, 74, 75, 78, 79, 89  
*Cyperus esculentus*, 15
- Desmodium*, 89  
*Diospyros virginiana*, 20
- Euphorbia*, 14
- Fagus grandifolia*, 29, 78  
*Fragaria*, 26, 35, 81, 89
- Gaylussacia*, 20  
*Gutierrezia*, 48
- Hamamelis*, 14, 26, 29, 70  
*Helianthus*, 52, 54, 55, 61
- Ilex*, 72  
*Impatiens*, 33
- Juglans*, 71, 78  
*Juncus*, 15
- Kalmia*, 30, 69
- Lactuca*, 45  
*Larix*, 16, 91  
*Ledum groenlandicum*, 77  
*Lupinus*, 90  
*Lysimachia*, 19
- Myrica*, 25, 30
- Nemopanthus mucronata*, 72
- Ostrya virginiana*, 20, 29
- Physocarpus opulifolius*, 31  
*Picea*, 19, 41, 51, 66, 76, 93  
*P. glauca*, 66, 74  
*P. mariana*, 66, 74  
*Pinus*, 38, 39, 40, 48, 51, 91, 92  
*P. banksiana*, 38, 39, 94  
*P. resinosa*, 38, 51, 94  
*P. sylvestris*, 38  
*P. virginiana*, 51  
*Pisum*, 93  
*Platanus occidentalis*, 80, 81  
*Populus*, 21, 22, 23, 31, 32, 35, 63, 73, 75, 82, 92  
*P. balsamifera*, 75  
*P. deltoides*, 63  
*P. tremuloides*, 69, 75  
*Prunus*, 27, 31, 36, 41, 80, 82, 88, 89  
*Pseudotsuga*, 51  
*Pyrus*, 31, 36, 68, 79, 81, 82, 88, 89, 94  
*P. malus*, 41
- Quercus*, 26, 63, 68, 75, 78, 80, 81, 88, 94
- Ratibida*, 58  
*Rhododendron*, 77  
*Rhus*, 14  
*Robinia*, 95  
*Rosa*, 36, 37, 61, 88  
*Rubus*, 25, 31, 36, 41, 73, 81  
*Rudbeckia*, 57, 58, 59
- Salix*, 21, 22, 23, 32, 63, 69, 75, 76, 77, 82, 94  
*Sarracenia*, 18  
*Sassafras albidum*, 24  
*Scirpus*, 15  
*Silphium*, 52  
*Smilax*, 96  
*Solidago*, 42, 43, 44, 47, 50, 53, 54, 57, 58, 62  
*Spermolepis*, 35  
*Spiraea*, 16, 30, 33, 37  
*Stachys*, 18
- Tilia*, 28  
*Trifolium*, 35, 90  
*T. hybridum*, 80  
*T. pratense*, 80  
*T. repens*, 80
- Ulmus*, 27
- Vaccinium*, 20, 69, 72  
*Verbascum*, 18  
*Verbena*, 18  
*Viburnum*, 28  
*Vitis*, 16
- Xanthium*, 47, 56